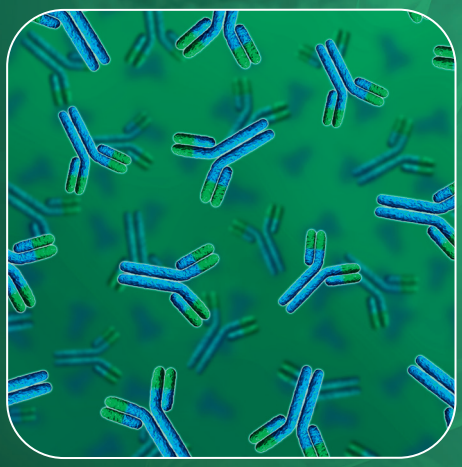
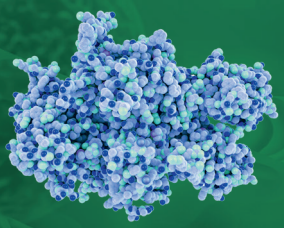
YMC

YMC Biochromatography Columns



RP
SEC
IEX
HIC

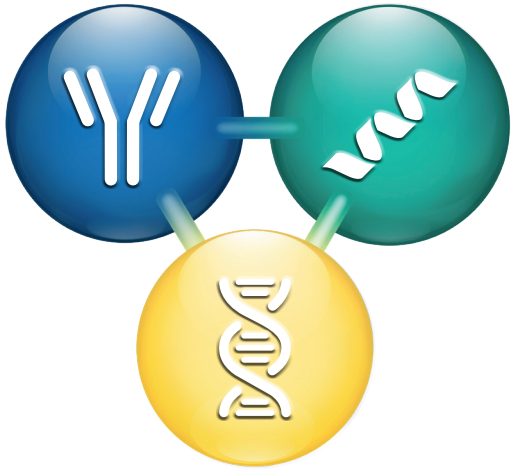


HPLC Columns for Biochromatography

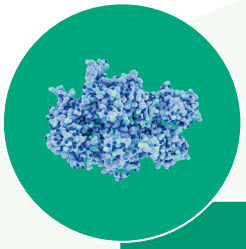
	Reversed Phase (RP)	Size Exclusion (SEC)	Ion Exchange (IEX)	Hydrophobic Interaction (HIC)
Separation principle	Hydrophobicity	Molecular size	Electric charge	Hydrophobicity
Max. MW	Up to about 150,000 Da	Up to about 1,000,000 Da	Up to several millions Da	Up to about 1,000,000 Da
Resolution	+++	++	+++	+++
Speed	+++	+	++/+++	++/+++
Loading	++	+	+++	+++
Stability	+/++	+++	+++	+++
Usage (e.g.)	• Peptide mapping • LC/MS • Nucleic acids and oligonucleotides	• Impurity analysis of antibody-drug conjugates • mAb separation	• Proteins/mAb • Charge variant analysis • Isoform analysis • Nucleic acids and oligonucleotides	• Drug-binding analysis of antibody-drug conjugates

Application data mainly by courtesy of YMC Co., Ltd.

Contents

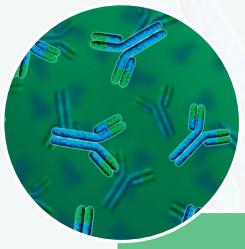


Phase selection guide



Proteins / Peptides

RP	IEX	SEC	HIC
YMC-Triart C18	BioPro IEX QA	YMC-Pack Diol-60	BioPro HIC HT
YMC-Triart Bio C18	BioPro IEX QF	YMC-Pack Diol-120	BioPro HIC BF
Meteoric Core C18 BIO	BioPro IEX SP	YMC-Pack Diol-200	
YMC-Triart Bio C4	BioPro IEX SF	YMC-Pack Diol-300	
		YMC-SEC MAB	



(Monoclonal) Antibodies

- **IEX**
  - BioPro IEX QA
  - BioPro IEX QF
  - BioPro IEX SP
  - BioPro IEX SF
- **SEC**
  - YMC-Pack Diol-200
  - YMC-Pack Diol-300
  - YMC-SEC MAB
- **HIC**
  - BioPro HIC HT
  - BioPro HIC BF
- **RP**
  - YMC-Triart Bio C4

Oligonucleotides / Nucleic Acids

- **IP-RP**
  - YMC-Triart C18
  - YMC-Triart Bio C18
  - Hydrosphere C18
  - YMC-Triart C8
  - YMC-Triart Bio C4
- **IEX**
  - BioPro IEX QA
  - BioPro IEX QF
- **SEC**
  - YMC-Pack Diol-120
  - YMC-Pack Diol-200
  - YMC-Pack Diol-300

Bio QC – Validation kit

Method Validation Kits for BioLC

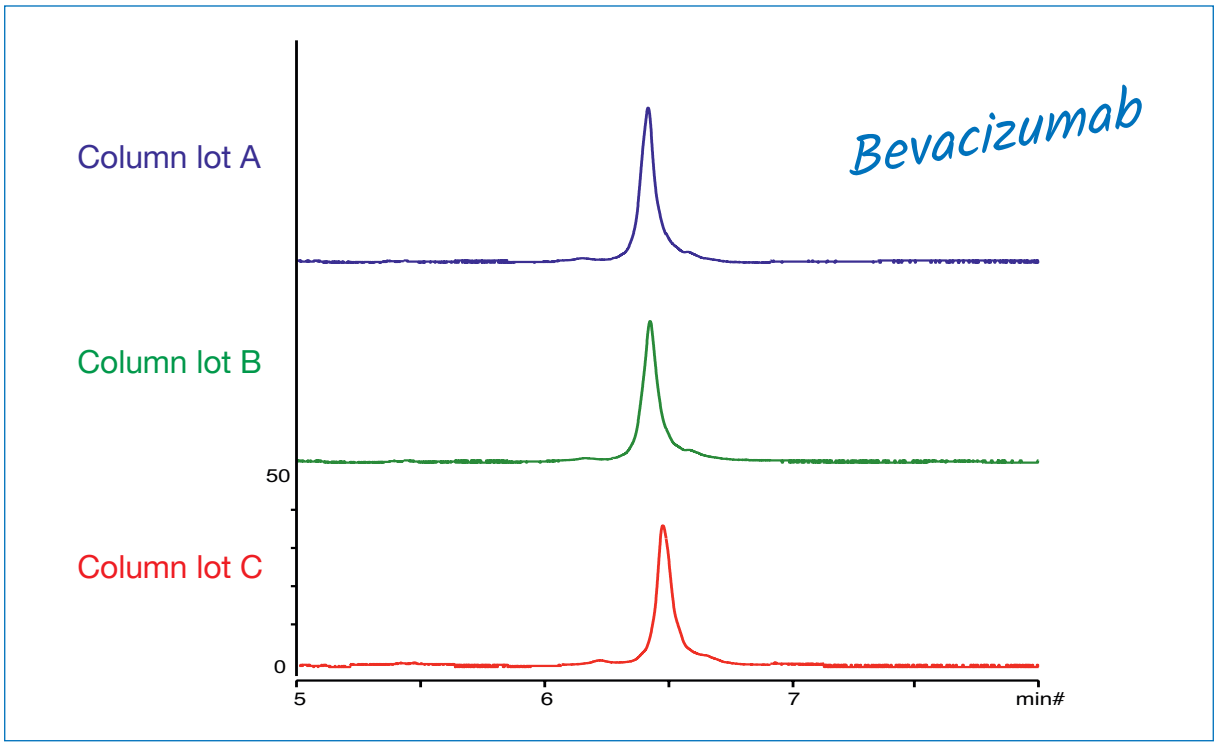
- **for documentation of robustness and reproducibility**
- **three analytical columns from specified lots**

Validation kit:

contains three analytical columns packed with stationary phases from three different batches, in order to solely test the robustness of the particular method.

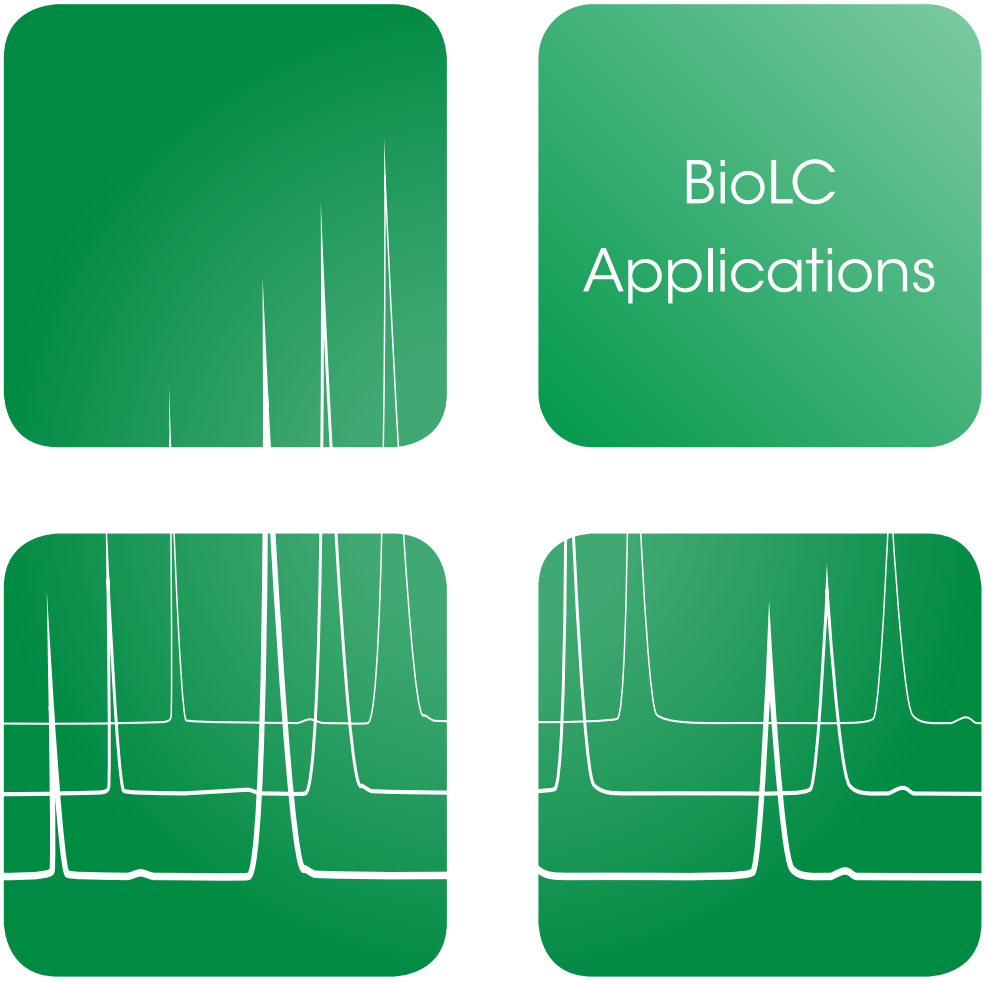
Available dimensions:

Length: 30 or 33, 50, 75, 100, 150, 250, 300 mm
ID: 2.0 or 2.1, 3.0, 4.0, 4.6, 8.0 mm



To order columns for validation, simply order a quantity of three of your desired part number, and include a note requesting three separate lots for validation purposes.

Please call YMC or contact your YMC representative for assistance.

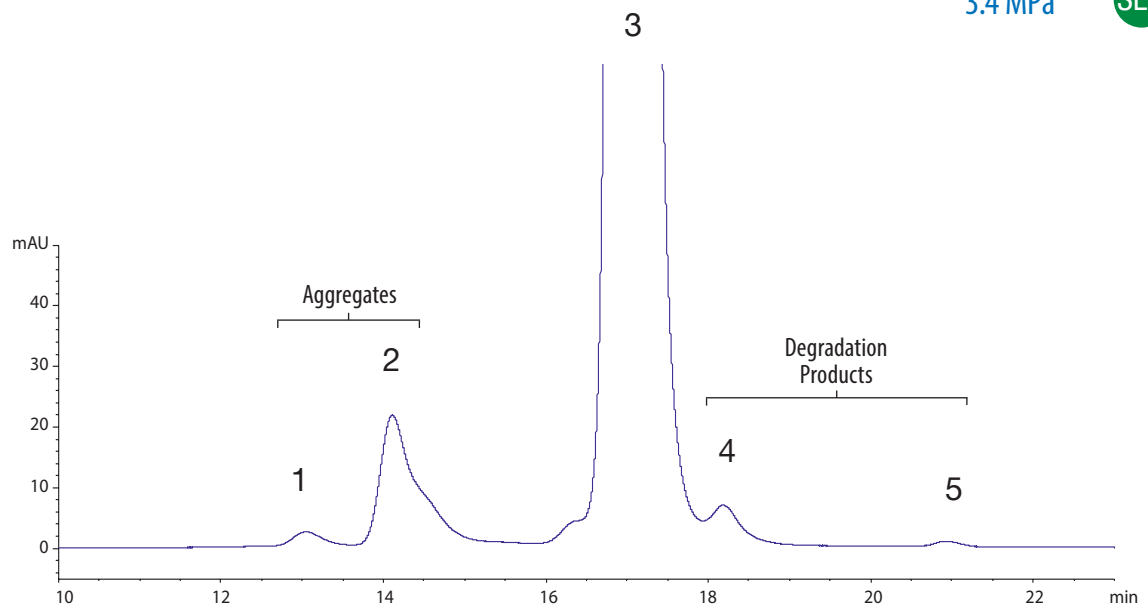

BioLC
Applications

BioLC applications – Antibodies

Bevacizumab and its fragments and aggregates

3.4 MPa

SEC

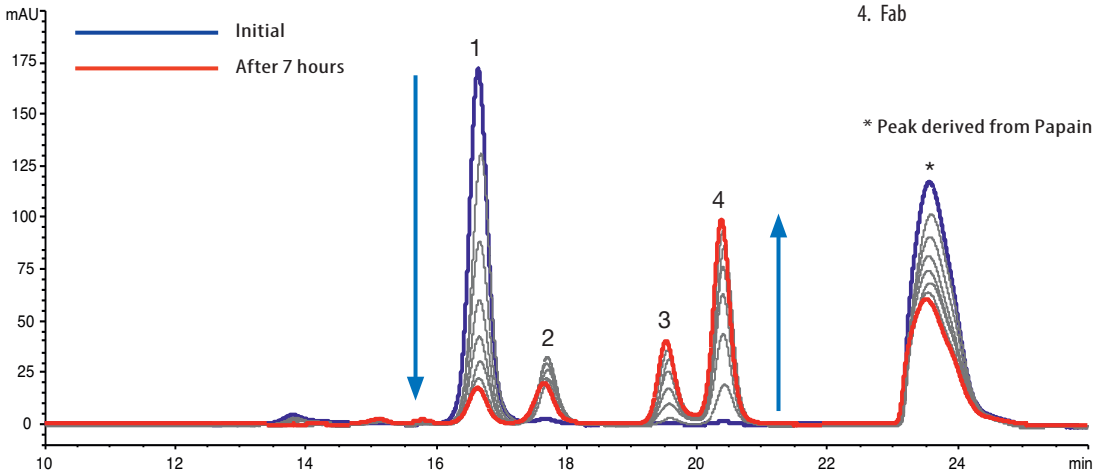


Column:	YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID	Detection:	UV at 280 nm
Part No.:	DLM25S03-3046WT	Cell path:	10 mm
Eluent:	0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl	Injection:	10 µL (5 mg/mL)
Flow rate:	0.165 mL/min	Sample:	Bevacizumab (Avastin®)
Temperature:	25°C		

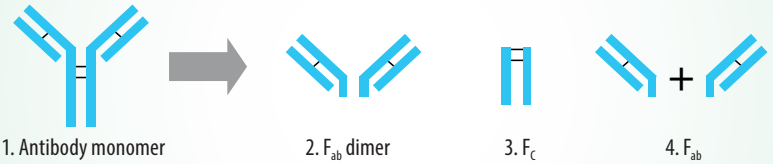
Analysis of digested antibody

SEC

1. Antibody monomer
2. Fab dimer
3. FC
4. Fab



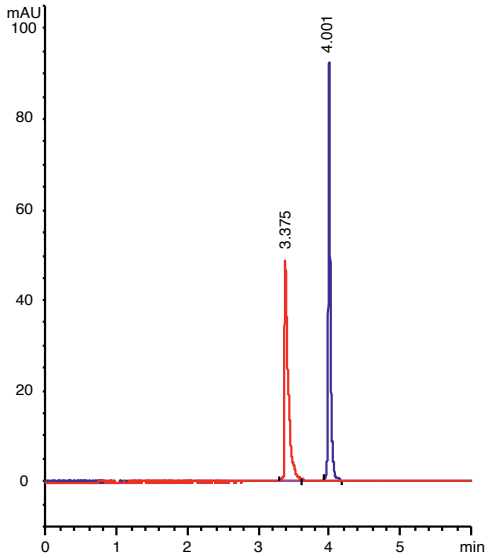
Column:	YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID	Temperature:	25 °C
Part No.:	DLM25S03-3046WT	Detection:	UV at 280 nm
Eluent:	0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl	Injection:	2 µL (3 mg/mL)
Flow rate:	0.165 mL/min	Sample:	Humanised monoclonal IgG1 + Papain



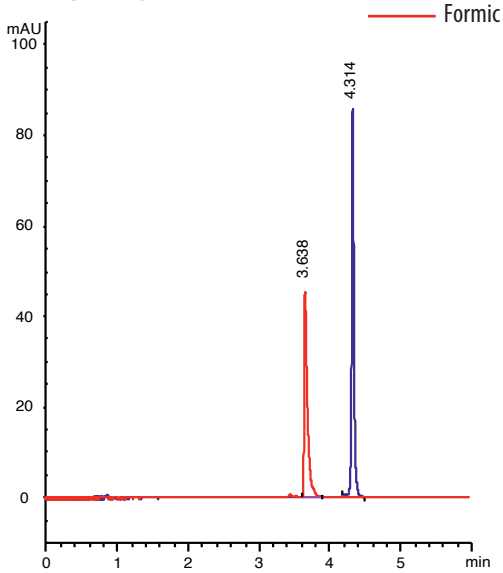
Use of MS compatible conditions for antibody analysis by RP

RP

Adalimumab (Humira®)



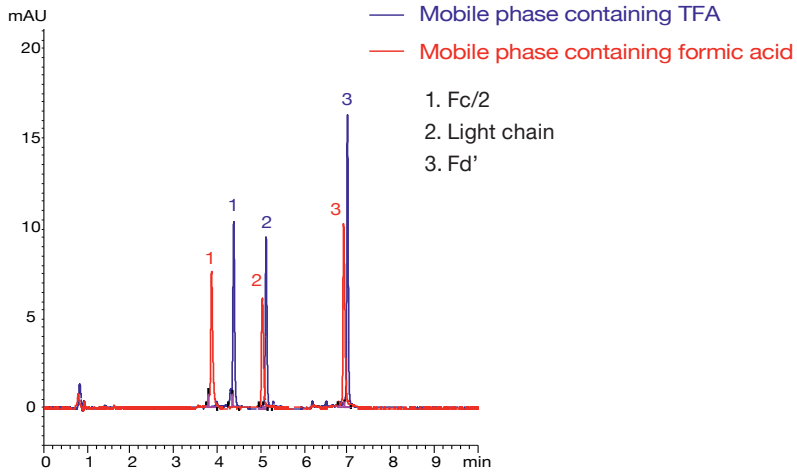
NISTmAb



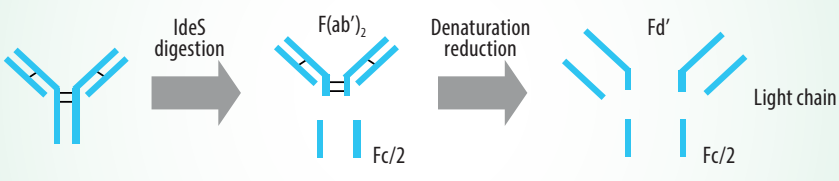
Column:	YMC-Triart Bio C4 (1.9 µm, 30 nm) 150 x 2.1 mm ID	Flow rate:	0.4 mL/min
Part No.:	TB30SP9-15Q1PT	Temperature:	80 °C
Eluent:	A) water/TFA or formic acid (100/0.1)	Detection:	UV at 280 nm (0.13 s, 40 Hz)
	B) acetonitrile/TFA or formic acid (100/0.1)	Injection:	2 µL (0.5 mg/mL)
Gradient:	10–95%B (0–10 min)		

LC/MS compatible analysis of monoclonal antibody fragments

RP

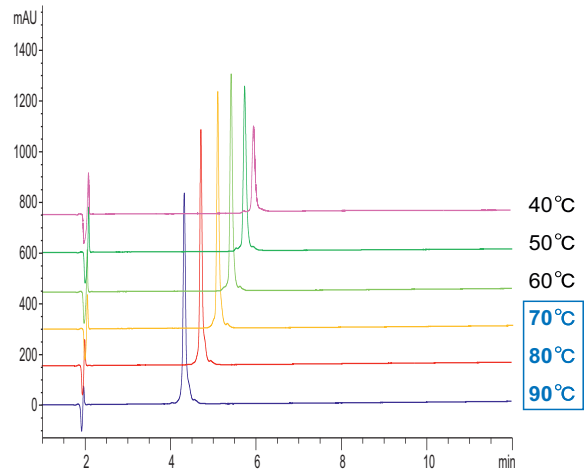


Column:	YMC-Triart Bio C4 (1.9 µm, 30 nm) 150 x 2.1 mm ID	Gradient [formic acid]:	20–45%B (0–10 min), 90%B (10–12.5 min)
Part No.:	TB30SP9-15Q1PT	Flow rate:	0.4 mL/min
Eluent [TFA]:	A) water/TFA (100/0.1)	Temperature:	80°C
	B) acetonitrile/TFA (100/0.1)	Injection:	4 µL (0.25 mg/mL)
Gradient [TFA]:	25–50%B (0–10 min), 90%B (10–12.5 min)	Detection:	UV at 280 nm
Eluent [formic acid]:	A) water/formic acid (100/0.1)	Sample:	mAb Subunit Standard (Waters Corp.)
	B) acetonitrile/formic acid (100/0.1)		



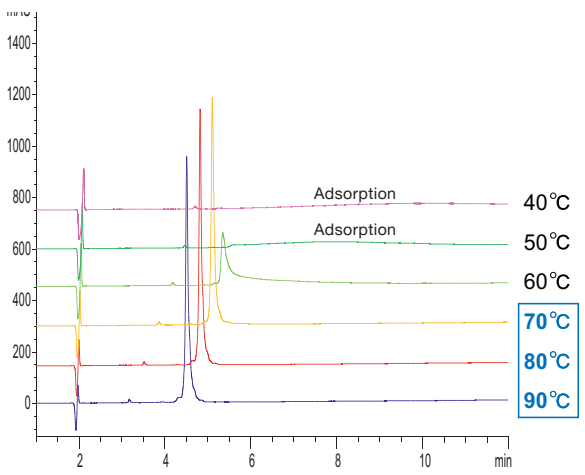
BioLC applications – Antibodies

Adalimumab (Humira®, MW: ca. 148 kDa)



Bevacizumab (Avastin®, MW: ca. 148 kDa)

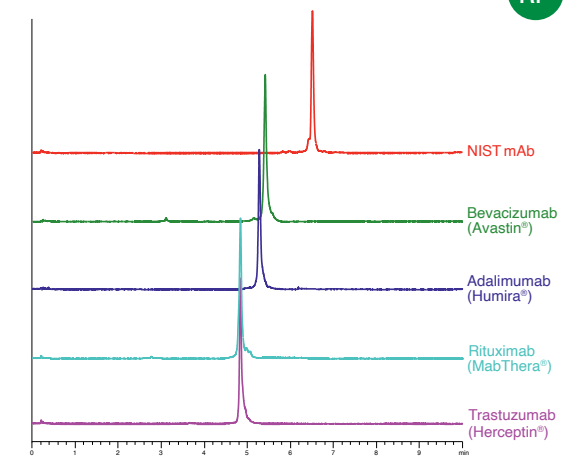
RP



Column:	YMC-Triart Bio C4 (3 µm, 30 nm) 150 x 3.0 mm ID
Part No.:	TB30S03-1503PTH
Eluent:	A) water/TFA (100/0.1) B) acetonitrile/TFA (100/0.1)
Gradient:	30–60%B (0–15 min), 90%B (15–30min)
Flow rate:	0.4 mL/min
Detection:	UV at 220 nm
Injection:	4 µL
Sample:	Adalimumab (0.5 mg/mL) or Bevacizumab (0.5 mg/mL)

Analysis of different monoclonal antibodies

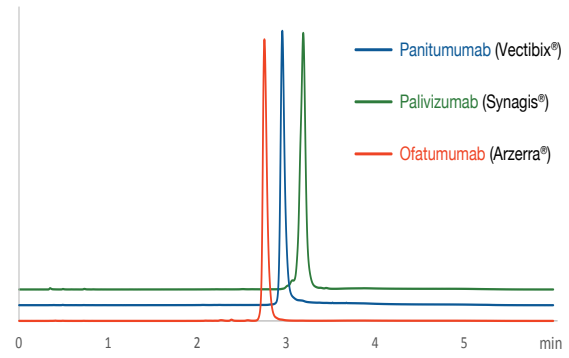
RP



Column:	YMC-Triart Bio C4 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.:	TB30SP9-05Q1PT
Eluent:	A) water/TFA (100/0.1) B) acetonitrile/TFA (100/0.1)
Gradient:	25–45%B (0–10 min)
Flow rate:	0.4 mL/min
Temperature:	80 °C
Detection:	UV at 280 nm (0.13s, 40Hz)
Injection:	2 µL (0.5 mg/mL)

Analysis of challenging monoclonal antibodies

RP



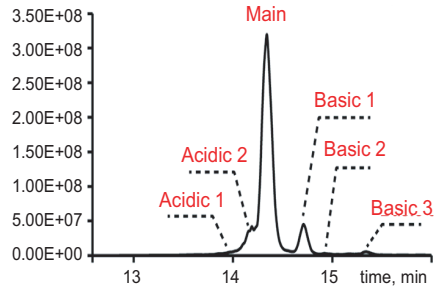
Column:	YMC-Triart Bio C4 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.:	TB30SP9-05Q1PT
	A) water/TFA (100/0.1)
	B) acetonitrile/TFA (100/0.1)
Gradient:	25–50 %B (0–4 min)
Flow rate:	0.4 mL/min
Temperature:	90 °C
Detection:	Fluorescence: ex 280 nm, em 350 nm
Injection:	0.5 µL

By courtesy of University of Geneva,
Institute of Pharmaceutical Sciences of Western Switzerland (ISPSO)

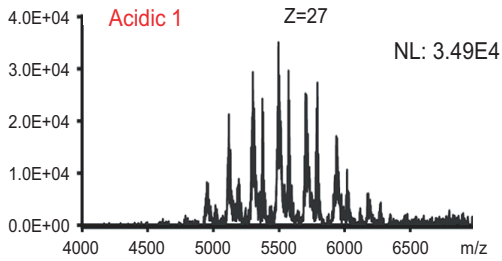
Native online SCX-MS analysis of monoclonal antibodies

IEX

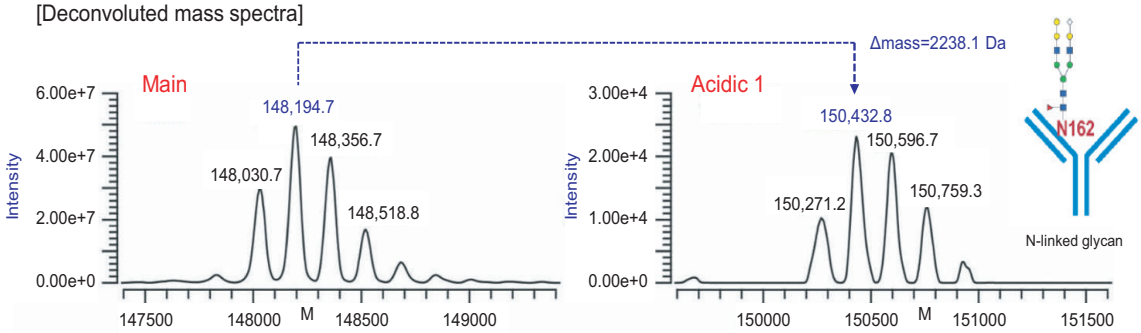
[TIC, native NISTmAb]



[Raw mass spectrum]



[Deconvoluted mass spectra]



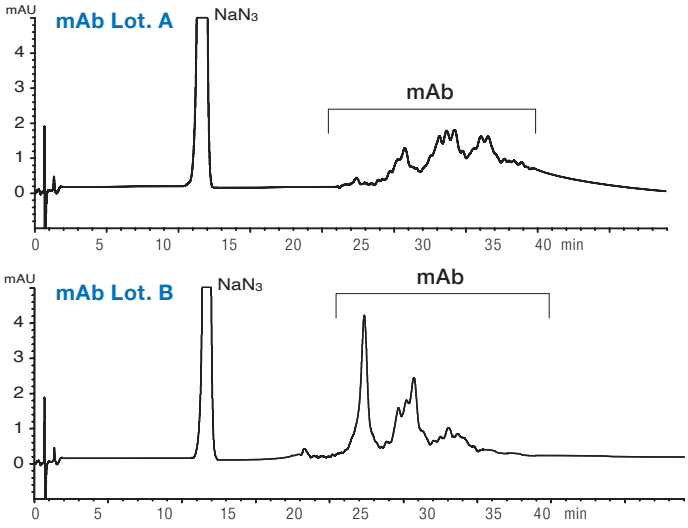
Column:	BioPro IEX SF (5 µm) 100 x 4.6 mm ID	Temperature:	45 °C
Part No.:	SF00S05-1046WP	Detection:	nanospray ionization-mass spectrometry (NSI-MS)
Eluent:	A) 20 mM CH_3COONH_4-CH_3COOH (pH 5.6)	Load:	50 µg
	B) 140 mM CH_3COONH_4-10 mM NH_4HCO_3 (pH 7.4)	System:	LC) ACQUITY UPLC I-Class system (Waters)
Gradient:	0%B (0–2 min), 0–100%B (2–18 min), 100%B (18–22 min)		MS) ExactiveTM Plus EMR mass spectrometer
Flow rate:	0.4 mL/min		(Thermo Fisher Scientific)
	(To enable online simultaneous UV and MS detection, a post-column analytical splitter (~400:1 ratio) was connected)		

By courtesy of S. Wang, Regeneron Pharmaceuticals Inc.

Reference: Y. Yan, A. P. Liu, S. Wang, T. J. Daly und N. Li, Ultrasensitive Characterization of Charge Heterogeneity of Therapeutic Monoclonal Antibodies, Anal. Chem., 2018, 90, 13013-20.

Different production batches of IgG1

IEX

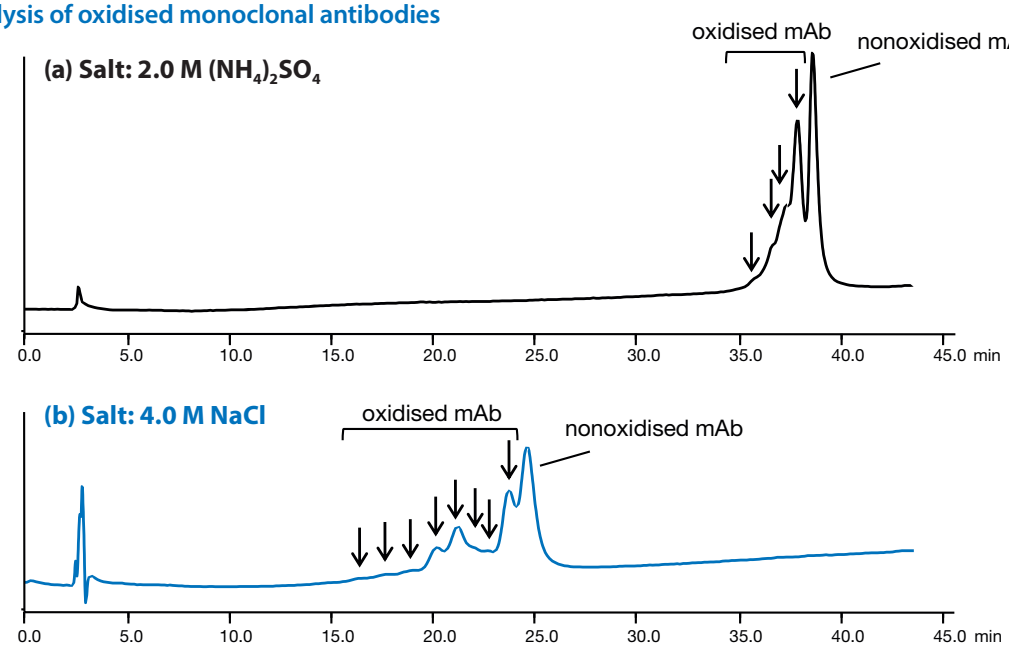


Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID	Temperature:	25 °C
Part No.:	QF00S05-1046WP	Detection:	UV at 220 nm
Eluent:	A) 20 mM Tris-HCl (pH 8.1)	Injection:	14 µL (0.1 mg/mL)
	B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl	Sample:	Mouse monoclonal IgG1 anti-human IgG4
Gradient:	10–25%B (0–60 min)		(Purified by DEAE chromatography, containing NaN_3)
Flow rate:	1.0 mL/min (360 cm/h)		

BioLC applications – Antibodies

Analysis of oxidised monoclonal antibodies

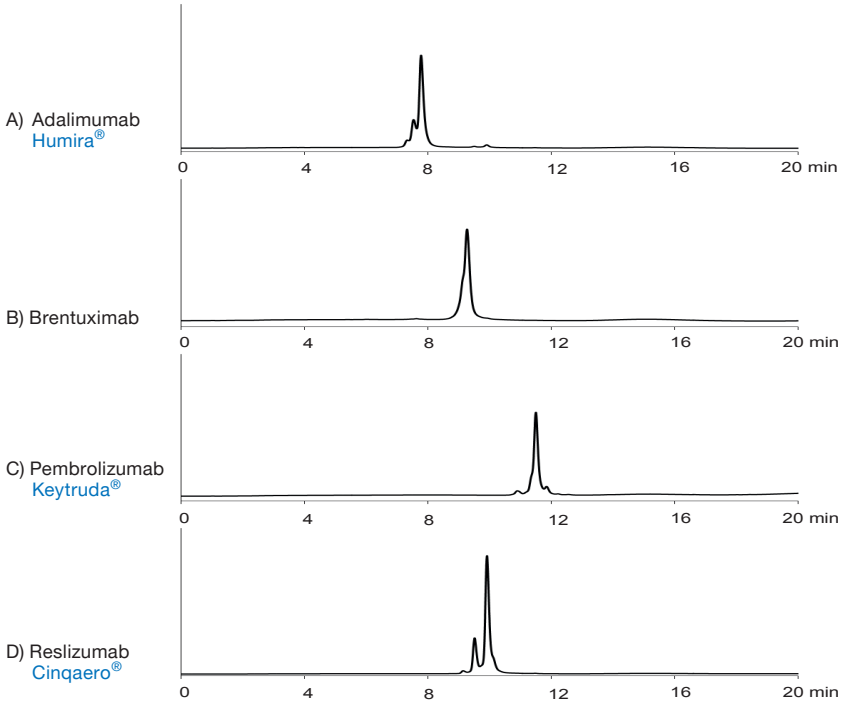
HIC



Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Flow rate:	0.3 mL/min
Part No.:	BHB00S04-1046WT	Temperature:	25 °C
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing salt	Detection:	UV at 280 nm
	B) 100mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Injection:	5 µL (1.0 mg/mL)
Gradient:	40–80%B (0–40 min), 80%B (40–45 min)	Sample:	oxidised NISTmAb

HIC analysis of different monoclonal antibodies using isopropanol as modifier

HIC

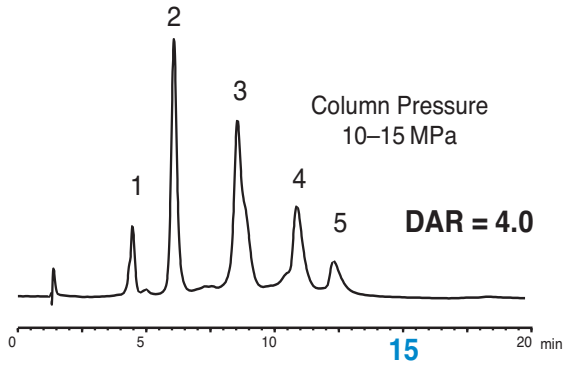


Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Temperature:	20 °C
Part No.:	BHB00S04-1046WT	Detection:	Fluorescence: ex 280 nm, em 360 nm
Eluent:	A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.4) containing 1.5 M $(NH_4)_2SO_4$	Injection:	3 µL (2 mg/mL)
	B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.4) / 2-propanol (85/15)		
Gradient:	0–100%B (0–20 min)		
Flow rate:	1.0 mL/min		

By courtesy of University of Geneva,
Institute of Pharmaceutical Sciences of Western Switzerland (ISPSO)

High throughput DAR determination by shortening analysis time

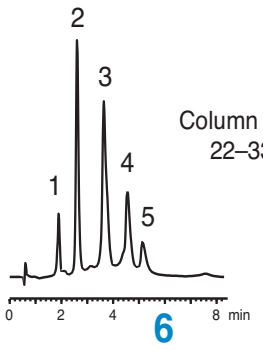
HIC



1. DAR 0
2. DAR 2
3. DAR 4
4. DAR 6
5. DAR 8

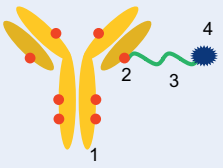
Flow rate
0.5 mL/min

2.5 x faster



Flow rate
1.2 mL/min

Column:	BioPro HIC HT (2.3 µm) 100 x 4.6 mm ID
Part No.:	BHH00SQ3-1046PTH
Eluent:	A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 1.0 M $(NH_4)_2SO_4$
	B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)/2-propanol (85/15)
Gradient:	0–100%B (0–15 min), 100%B (15–20 min)
	0–100%B (0–6.25 min), 100%B (6.25–8.3 min)
Temperature:	25°C
Detection:	UV at 280 nm
Injection:	10 µL
Sample:	Brentuximab vedotin (Adcetris®) (2.5 mg/mL)



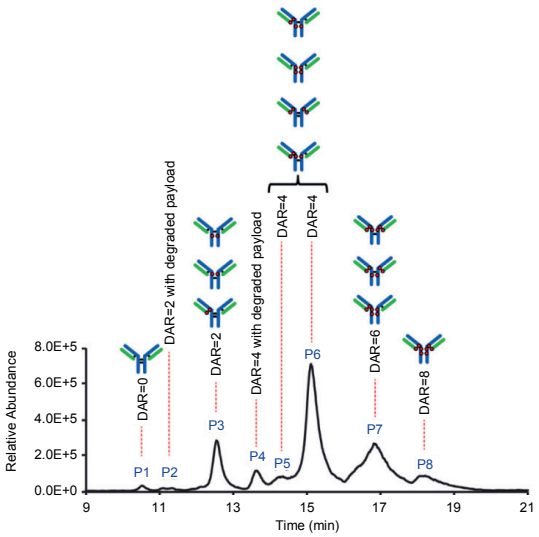
1: Antibody
2: Binding site
3: Linker
4: Cytotoxic drug

BioLC applications – Antibody-Drug-Conjugates

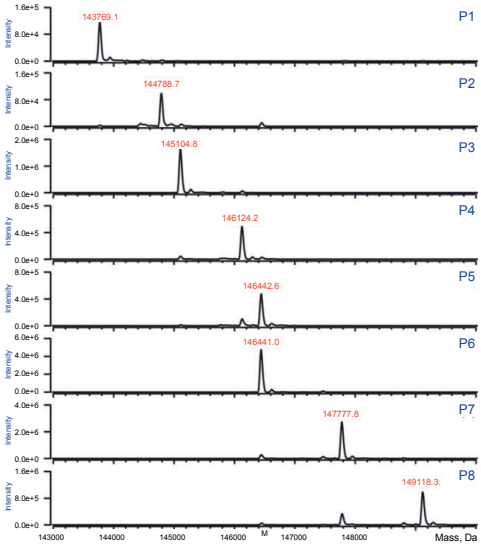
Native online HIC-MS analysis of cys-linked ADCs

HIC

[HIC-MS (BPC: base peak chromatogram)]



[Deconvoluted mass spectra]



Column: BioPro HIC BF (4 µm) 100 x 4.6 mm ID
Part number: BHB00S04-1046WT
Eluent: A) 3 M ammonium acetate in water
B) 2-propanol/water (30/70)
Gradient: 10%B (0–2 min), 10–97%B (2–18 min), 97%B (18–22 min)
Flow rate: 0.3 mL/min
Temperature: ambient
Detection: UV at 280 nm, NSI-MS

Injection: 10 µg
Sample: SigmaMAb ADC-mimic
Setup: Post-column makeup flow:
100% water at 1.5 mL/min (reducing salt conc. 6-fold)
Splitter to reduce the flow rate to 1–5 µL/min

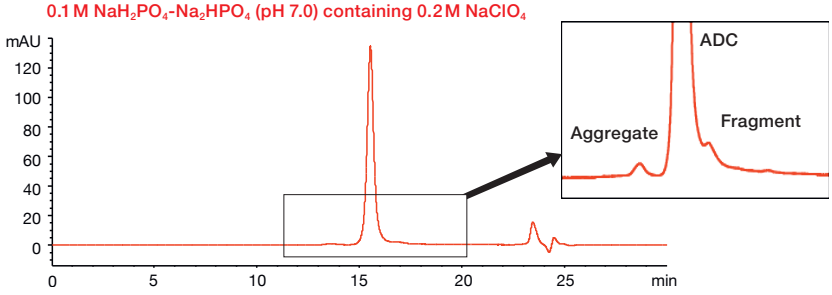
By courtesy of S. Wang, Regeneron Pharmaceuticals Inc.

Reference: Y. Yan, T. Xing, S. Wang, T. J. Daly, N. Li, Online coupling of analytical hydrophobic interaction chromatography with native mass spectrometry for the characterization of monoclonal antibodies and related products, J. Pharm. Biomed. Anal. 186 (2020) 113313.

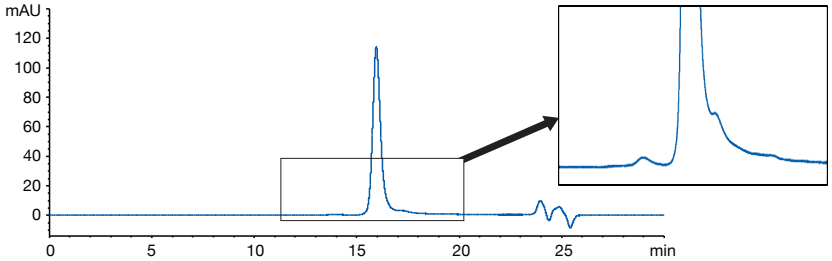
Separation of Brentuximab vedotin from its aggregates and fragments

SEC

0.1 M NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 0.2 M $NaClO_4$



0.1 M NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 0.2 M NaCl/2-propanol (85/15)



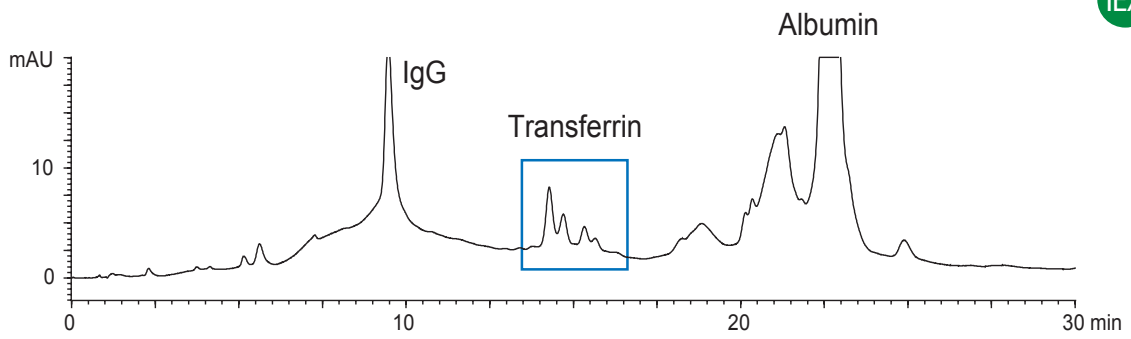
Column: YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID
Part No.: DLM25S03-3046WT
Eluent: 0.1 M phosphate buffer (pH = 7) containing 0.2 M $NaClO_4$
0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl/2-propanol (85/15)
Flow rate: 0.165 mL/min

Temperature: 25 °C
Detection: UV at 280 nm
Injection: 4 µL (2.5 mg/mL)
Sample: Brentuximab vedotin (Adcetris®) for injection

By courtesy of Prof. S. Manabe, Hoshi University, Tokyo/Tohoku University, Sendai Japan.

Separation of proteins in human serum

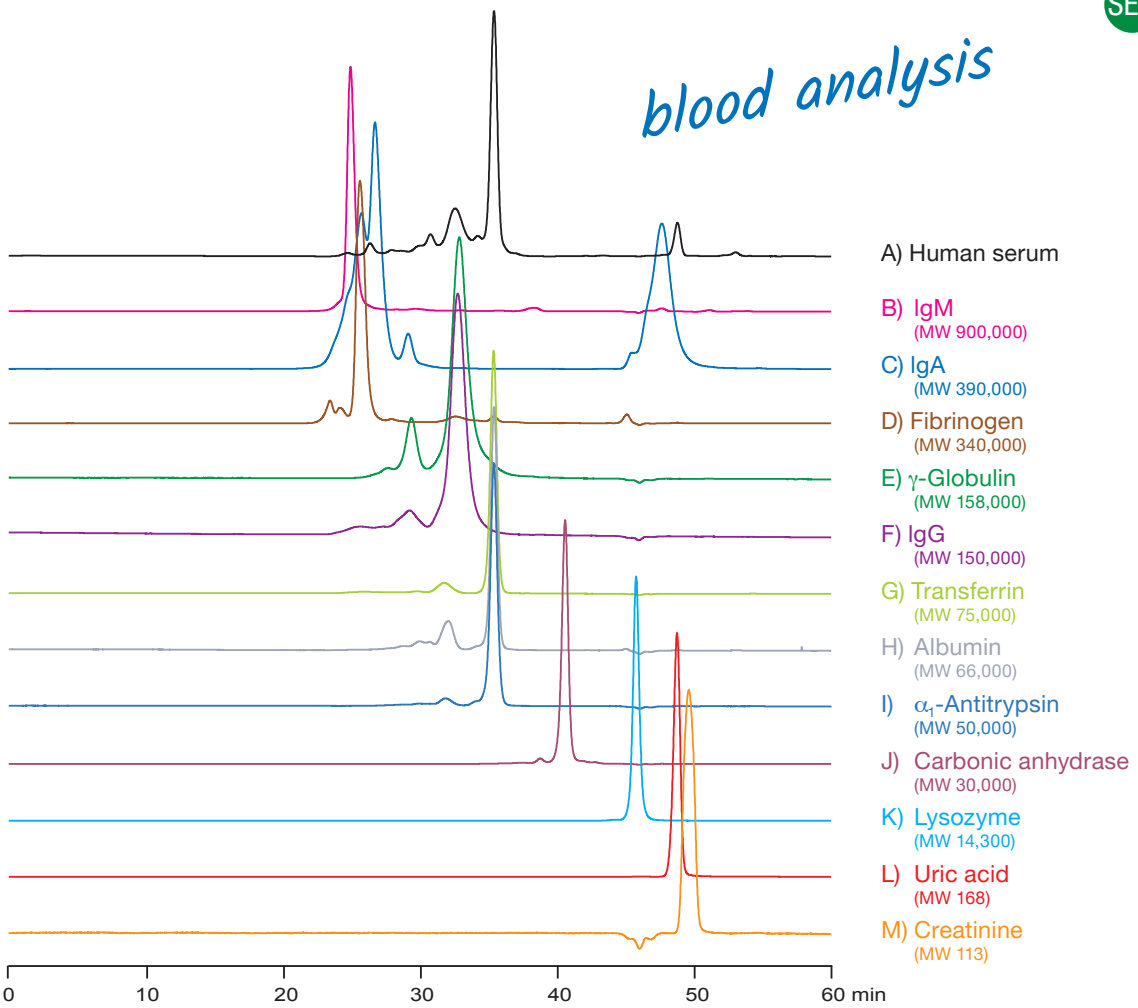
IEX



Column:	BioPro IEX QA (5 µm) 50 x 4.6 mm ID	Flow rate:	0.5 mL/min
Part No.:	QAA0S05-0546WP	Temperature:	25°C
Eluent:	A) 20 mM Tris-HCl (pH 8.6)	Detection:	UV at 280 nm
	B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl	Injection:	20 µL
Gradient:	0–30%B (0–15 min), 30–100%B (15–30 min)	Sample:	Human serum (100 µL/mL)

Plasma constituents

SEC



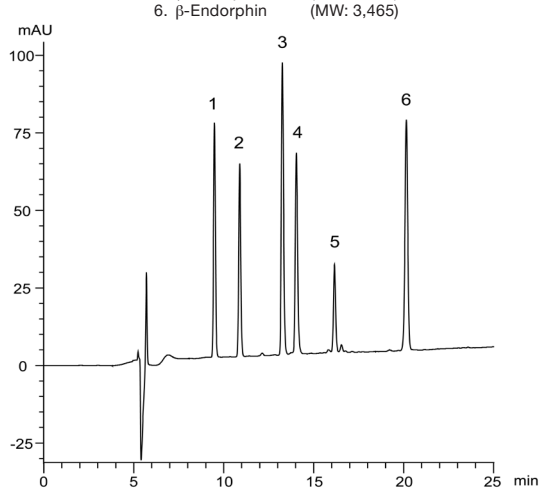
Columns:	YMC-Pack Diol-300 + Diol-200 (5 µm) 300 x 8.0 mm ID x 2	Temperature:	ambient (25 °C)
Part Nos.:	DL30S05-3008WT + DL20S05-3008WT	Detection:	UV at 280 nm
Eluent:	0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl	Injection:	20 µL (L: 1 µL)
Flow rate:	0.5 mL/min	Sample:	A) 100 µL/mL; B-M) 1.0 mg/mL

BioLC applications – Peptides

Peptides covering different MW

RP

1. Oxytocin (MW: 1,007)
2. Met-Enkephalin (MW: 574)
3. Leu-Enkephalin (MW: 556)
4. Neurotensin (MW: 1,673)
5. γ-Endorphin (MW: 1,859)
6. β-Endorphin (MW: 3,465)

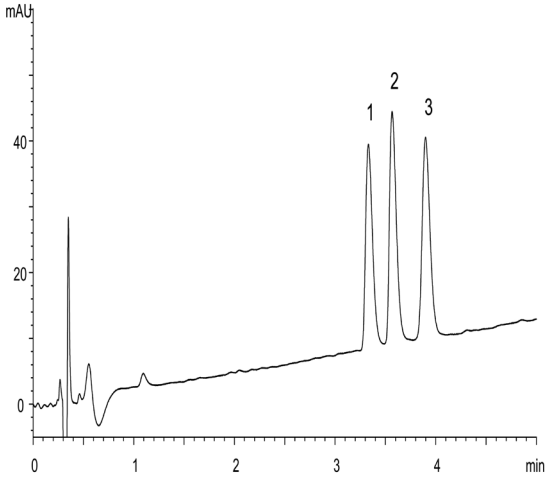


Column:	YMC-Triart C18 (5 µm, 12 nm) 150 x 2.0 mm ID
Part No.:	TA12S05-1502WT
Eluent:	A) water/TFA (100/0.1)
	B) acetonitrile/TFA (100/0.1)
Gradient:	20%–45%B (0–25 min)
Flow rate:	0.2 mL/min
Temperature:	37 °C
Detection:	UV at 220 nm
Injection:	2 µL (0.075 ≈ 0.25 mg/mL)

Antimicrobial peptides

RP

1. α-Defensin-1 (Human) (MW: 3,442)
2. α-Defensin-2 (Human) (MW: 3,371)
3. α-Defensin-3 (Human) (MW: 3,486)

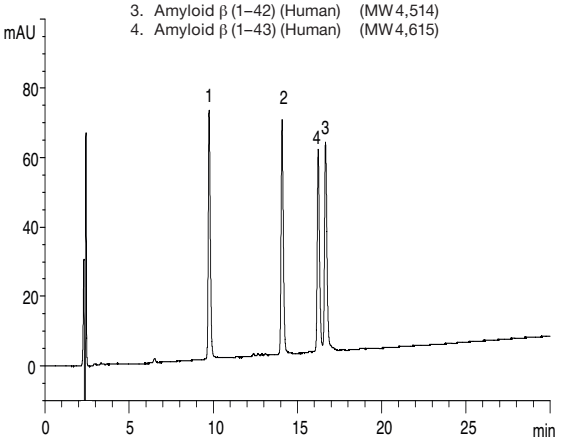


Column:	YMC-Triart C18 (1.9 µm, 12 nm) 50 x 2.0 mm ID
Part No.:	TA12SP9-0502PT
Eluent:	A) water/formic acid (100/0.1)
	B) 2-propanol/acetonitrile/formic acid (50/50/0.08)
Gradient:	10%–25%B (0–10 min)
Flow rate:	0.4 mL/min
Temperature:	70 °C
Detection:	UV at 220 nm
Injection:	1 µL (50 µg/mL)

Amyloid β-peptides

RP

1. Amyloid β (1–38) (Human) (MW 4,132)
2. Amyloid β (1–40) (Human) (MW 4,330)
3. Amyloid β (1–42) (Human) (MW 4,514)
4. Amyloid β (1–43) (Human) (MW 4,615)



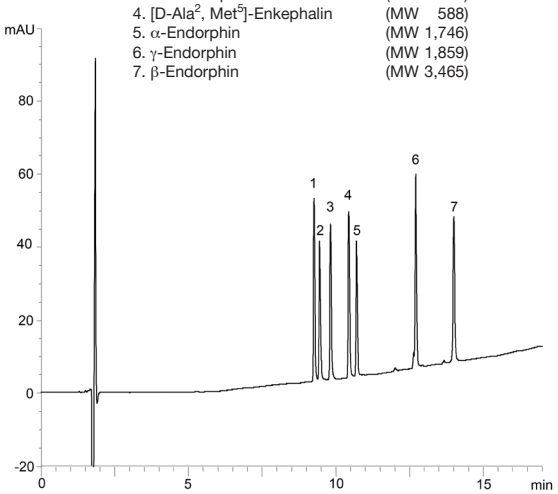
Amyloid β (1–43): Asp-Ala-Glu-Phe-Arg-His-Asp-Ser-Gly-Tyr-Glu-Val-His-His-Gln-Lys-Leu-Val-Phe-Phe-Ala-Glu-Asp-Val-Gly-Ser-Asn-Lys-Gly-Ala-Ile-Ile-Gly-Leu-Met-Val-Gly-Gly-Val-Val-Ile-Ala-Thr

Column:	YMC-Triart Bio C4 (3 µm, 30 nm) 150 x 3.0 mm ID
Part No.:	TB30S03-1503PTH
Eluent:	A) water/TFA (100/0.1)
	B) acetonitrile/TFA (100/0.1)
Gradient:	25–40%B (0–30 min), 90%B (30–40 min)
Flow rate:	0.4 mL/min
Temperature:	70 °C
Detection:	UV at 220 nm
Injection:	4 µL (each 0.1 mg/mL)

Peptides

RP

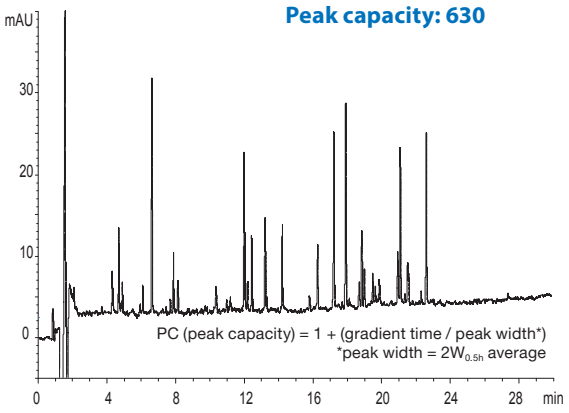
1. BAM-12P (MW 1,425)
2. [D-Ala^2, Met^5]-Enkephalinamide (MW 587)
3. Met-Enkephalin (MW 574)
4. [D-Ala^2, Met^5]-Enkephalin (MW 588)
5. α-Endorphin (MW 1,746)
6. γ-Endorphin (MW 1,859)
7. β-Endorphin (MW 3,465)



Column:	Meteoric Core C18 BIO (2.7 µm, 16 nm) 150 × 2.1 mm ID
Part No.:	CAW16SQ7-15Q1PT
Eluent:	A) water/TFA (100/0.1)
	B) acetonitrile/TFA (100/0.1)
Gradient:	15–55%B (0–15 min), 55%B (15–17 min)
Flow rate:	0.2 mL/min
Temperature:	40 °C
Detection:	UV at 220 nm
Injection:	2 µL (0.02–0.5 mg/mL)
Pressure:	14.9–16.1 MPa (2,160–2,330 psi)

Tryptic digest of Hemoglobin

RP

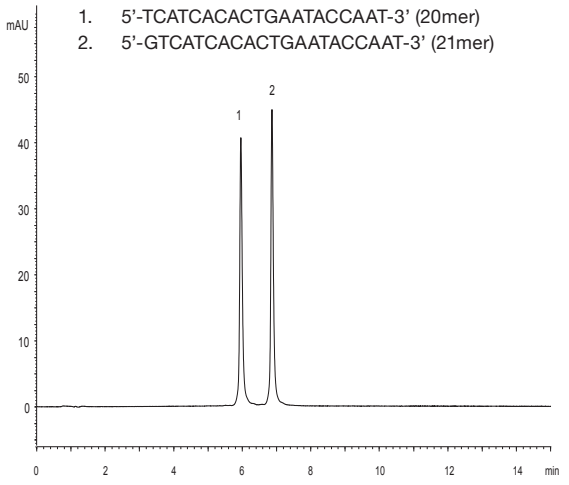


Column:	YMC-Triart C18 (1.9 µm, 12 nm) 200 x 2.0 mm ID (Two coupled 100 x 2.0 mm ID)
Part No.:	TA12SP9-1002PT (2x)
Eluent:	A) water/TFA (100/0.1) B) acetonitrile/TFA (100/0.08)
Gradient:	5–40%B (0–30 min)
Flow rate:	0.4 mL/min
Temperature:	70 °C
Detection:	UV at 220 nm
Injection:	20 µL
Sample:	Triptic digest of Bovine Hemoglobin (2.5 nmol/mL)
Pressure:	58.1–61.6 MPa (8,430–8,930 psi)

Separation of synthetic oligonucleotides (single-strand DNA)

IEX

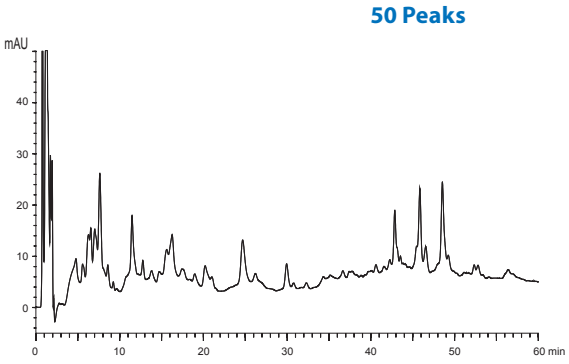
1. 5'-TCATCACACTGAATACCAAT-3' (20mer)
2. 5'-GTCATCACACTGAATACCAAT-3' (21mer)



Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID
Part No.:	QF00S05-1046WP
Eluent:	A) 10 mM NaOH B) 10 mM NaOH containing 1.0 M $NaClO_4$
Gradient:	25–55%B (0–15 min), 100%B (15–20 min)
Flow rate:	1.0 mL/min
Temperature:	25 °C
Detection:	UV at 260 nm
Injection:	4 µL (5 nmol/L)

Peptide mapping of tryptic digests of BSA with highest sensitivity

IEX



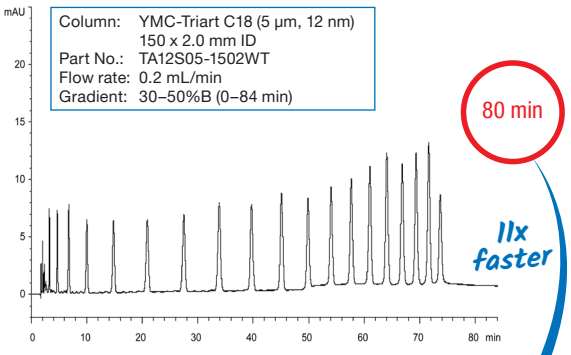
Column:	BioPro IEX QA (5 µm) 50 x 4.6 mm ID
Part No.:	QAA0S05-0546WP
Eluent:	A) 20 mM Tris-HCl (pH 8.6) B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
Gradient:	0–15%B (0-30 min), 15–60%B (30–60 min)
Flow rate:	0.5 mL/min
Temperature:	25°C
Detection:	UV at 220 nm
Injection:	20 µL
Sample:	Tryptic digest of BSA

Oligonucleotides d(T)2-20 method transfer from HPLC to UHPLC

RP

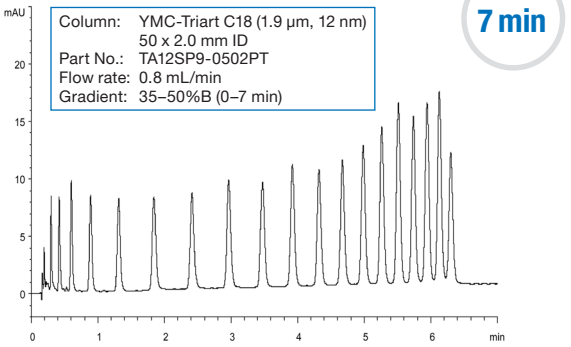
Conventional LC method

Column:	YMC-Triart C18 (5 µm, 12 nm) 150 x 2.0 mm ID
Part No.:	TA12S05-1502WT
Flow rate:	0.2 mL/min
Gradient:	30–50%B (0–84 min)



UHPLC method

Column:	YMC-Triart C18 (1.9 µm, 12 nm) 50 x 2.0 mm ID
Part No.:	TA12SP9-0502PT
Flow rate:	0.8 mL/min
Gradient:	35–50%B (0–7 min)



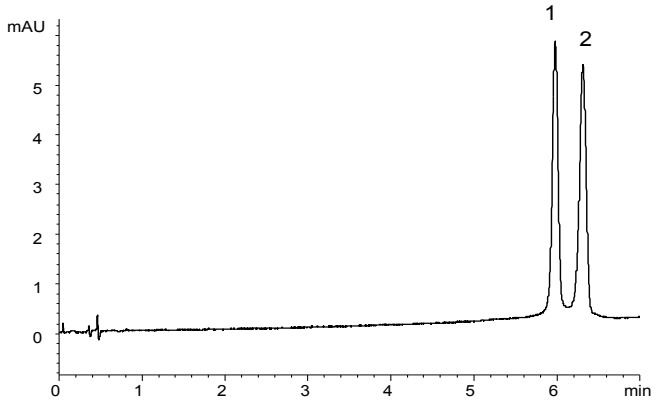
Eluent:	A) 10 mM di-n-butylamine-acetic acid (pH 6.0) B) methanol
Temperature:	37 °C
Detection:	UV at 269 nm
Injection:	1 µL (5 nmol/mL)

BioLC applications – Oligonucleotides

Challenging phosphorothioate oligonucleotides

RP

5'-U^C^A^U^C^A^C^A^C^U^G^A^A^U^A^C^C^A^A^U-3' (RNA 20mer)
5'-G^U^C^A^U^C^A^C^A^C^U^G^A^A^U^A^C^C^A^A^U-3' (RNA 21mer)
^=Phosphorothioated



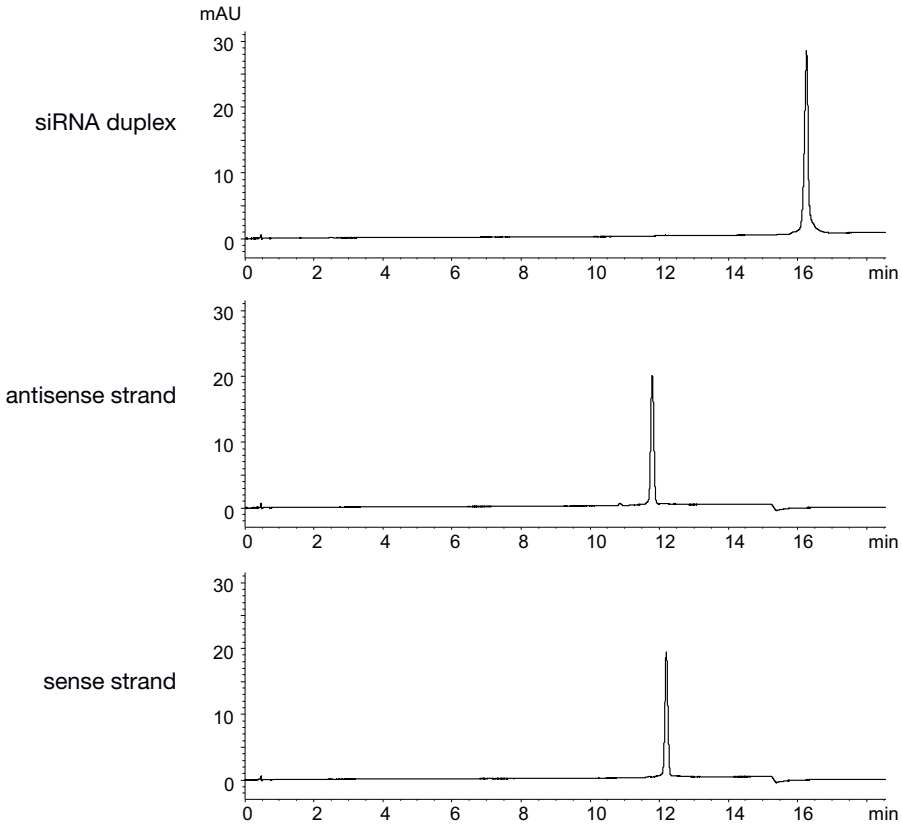
Column: YMC-Accura Triart Bio C18 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.: TA30SP9-05Q1PTC
Eluent: A) 15 mM triethylamine - 400 mM HFIP*
B) methanol
Gradient: 10–20%B (0–10 min)

Flow rate: 0.42 mL/min
Temperature: 65 °C
Detection: UV at 260 nm
Injection: 1 µL (each 1.0 nmol/mL)

*1,1,1,3,3,3-hexafluoro-2-propanol

siRNA under non-denaturing conditions

RP



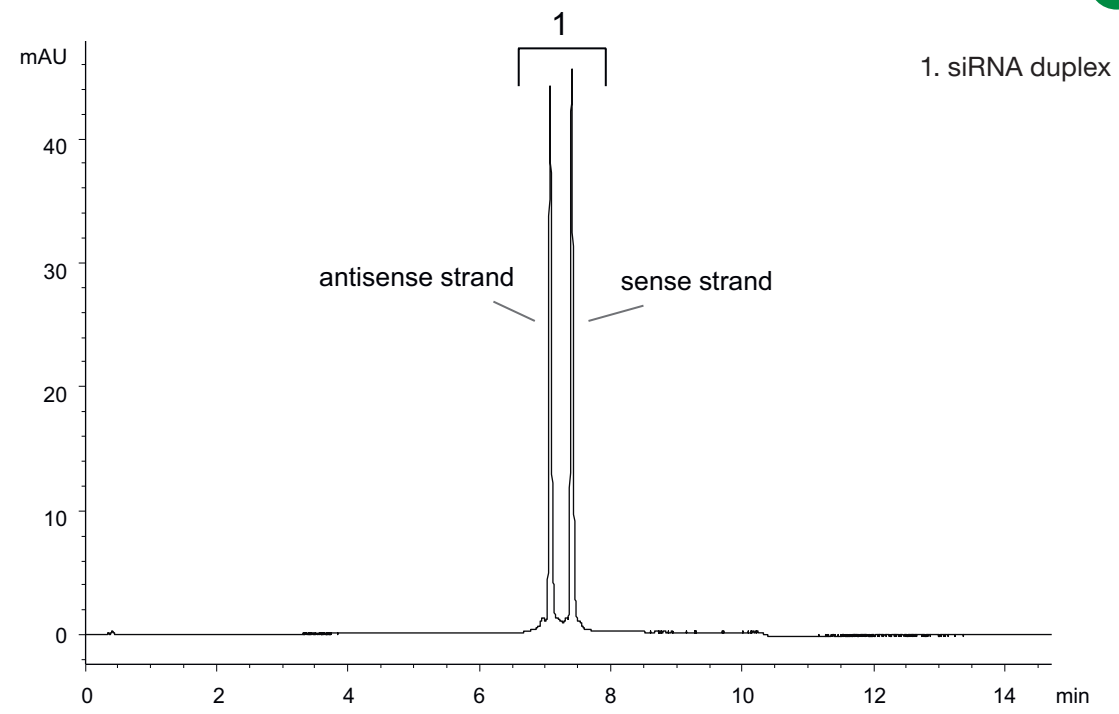
Column: YMC-Accura Triart Bio C18 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.: TA30SP9-05Q1PTC
Eluent: A) 15 mM triethylamine - 400 mM HFIP* (pH 8)
B) methanol
Gradient: 10%–28%B (0–18 min)

Flow rate: 0.42 ml/min
Temperature: 25 °C
Detection: UV at 260 nm
Injection: 1 µl (5 nmol/ml)
Sample: siRNA duplex & single strands

*1,1,1,3,3,3-hexafluoro-2-propanol

siRNA duplex under denaturing conditions

RP

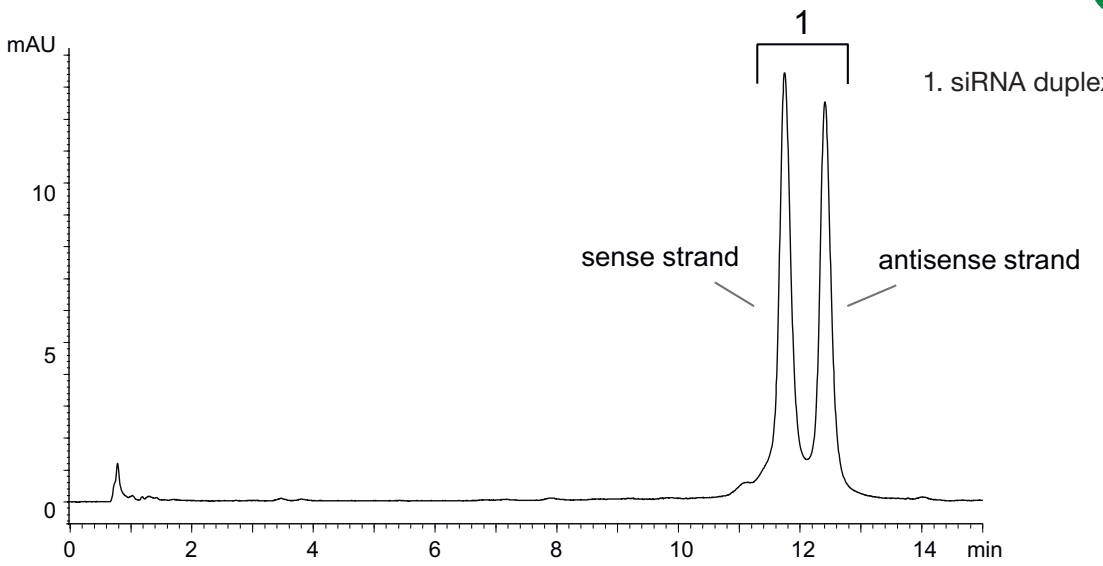


Column:	YMC-Accura Triart Bio C18 (1.9 µm, 30 nm) 50 x 2.1 mm ID	Flow rate:	0.42 ml/min
Part No.:	TA30SP9-05Q1PTC	Temperature:	65 °C
Eluent:	A) 15 mM TEAA* (pH 8)	Detection:	UV at 260 nm
	B) methanol	Injection:	1 µl (5 nmol/ml)
Gradient:	5%–20%B (0–15 min)	Sample:	siRNA duplex

*triethylammonium acetate

siRNA duplex under denaturing conditions

IEX

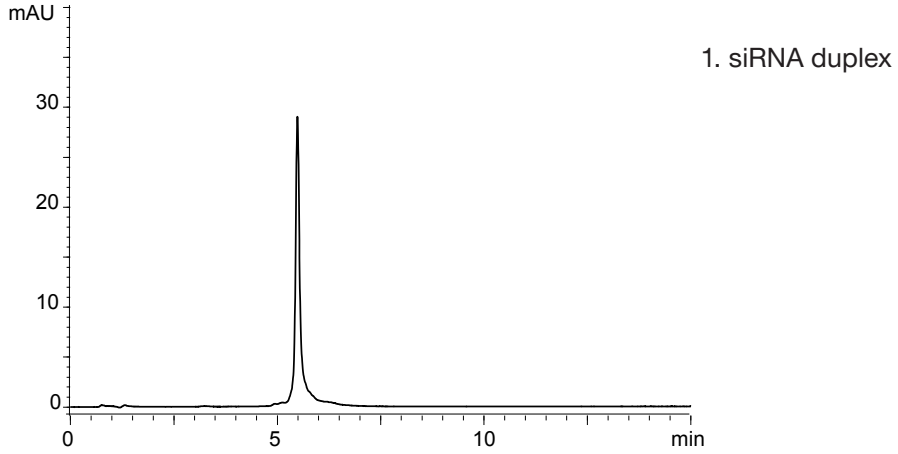


Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID	Flow rate:	1.0 ml/min
Part number:	QF00S05-1046WP	Temperature:	25 °C
Eluent:	A) 10 mM NaOH	Detection:	UV at 260 nm
	B) 10 mM NaOH containing 1 M $NaClO_4$	Injection:	4 µl (5 nmol/ml)
Gradient:	30%–37%B (0–15 min)	Sample:	siRNA duplex

BioLC applications – Oligonucleotides

AEX analysis of siRNA duplex under non-denaturing conditions

IEX

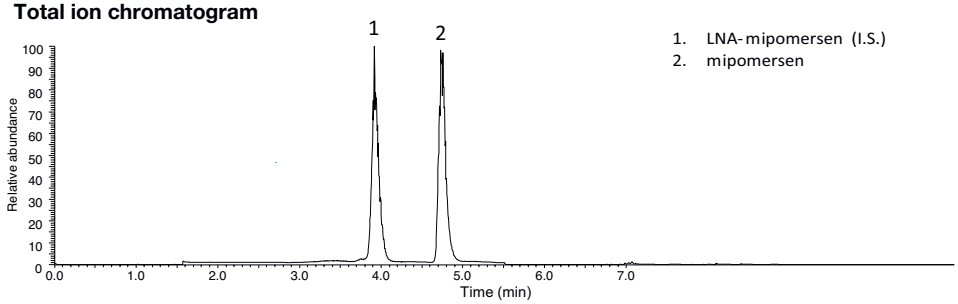


Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID	Flow rate:	1.0 ml/min
Part number:	QF00S05-1046WP	Temperature:	25 °C
Eluent:	A) 20 mM Tris-HCl (pH 8.1)	Detection:	UV at 260 nm
	B) 20 mM Tris-HCl (pH 8.1) containing 1 M $NaClO_4$	Injection:	4 µl (5 nmol/ml)
Gradient:	25%–40%B (0–15 min)	Sample:	siRNA duplex

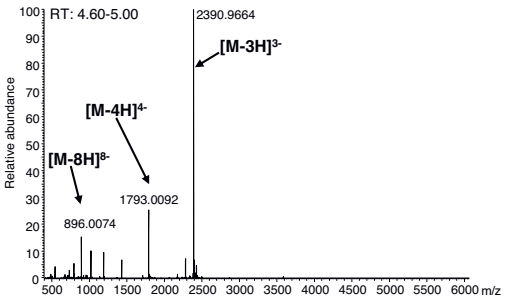
LC-HRMS analysis of the antisense oligonucleotide mipomersen (Kynamro®)

RP

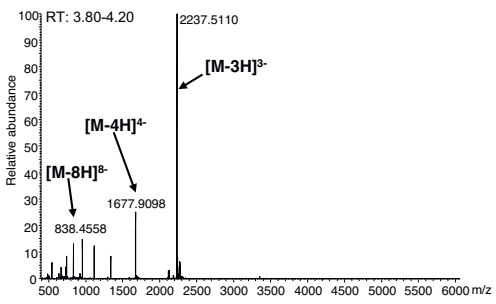
Total ion chromatogram



Mass spectrum of mipomersen



Mass spectrum of LNA-mipomersen



Column:	**YMC-Triart C8 metal-free** (1.9 µm, 12 nm)* 100 x 2.1 mm ID	[Column wash steps]	40–70%B (5.0–5.1 min), 70%B (5.1–7.0 min), 70–10%B (7.0–7.1 min), 10%B (7.1–8.0 min), 10–90%B (8.0–8.1 min), 90%B (8.1–9.0 min), 90–10%B (9.0–9.1 min), 10%B (9.1–10.0 min), 10–90%B (10.0–10.1 min), 90 %B (10.1–11.0 min), 90–10 %B (11.0–11.1 min)
Part No.:	TO12SP9-10Q1PTP	Flow rate:	0.3 mL/min
Eluent:	A) water/triethylamine/HFIP** (100/0.4/2; triethylamine 28.0 mM, HFIP 135.8 mM)	Temperature:	50 °C
	B) methanol/triethylamine/HFIP (100/0.4/2)	Injection:	10 µL (1000 ng/mL)
Gradient:	[Sample separation step] 10–40%B (0–5.0 min)	System :	LC) Vanquish Binary Pump H system HRMS) Orbitrap HRMS Q Exactive Plus

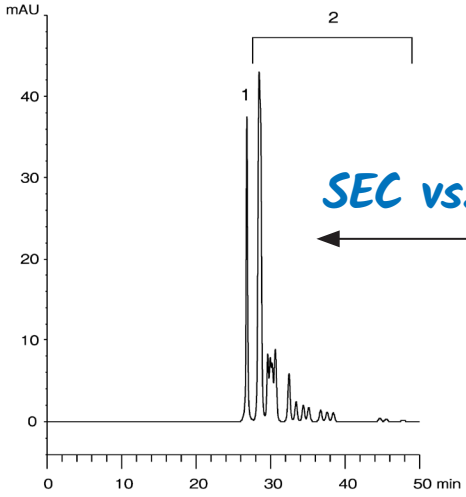
* Prewash the column prior to the first use with water/methanol/phosphoric acid (70/30/0.1) for 1 hour
** 1,1,1,3,3,3-hexafluoro-2-propanol

Reference: Y. Sun et al, Development of a bioanalytical method for an antisense therapeutic using high-resolution mass spectrometry, Bioanalysis, 2020 NOV 26, doi: 10.4155/bio-2020-0225.

Plasmid pBR322 restriction and pBR322 Hae III restriction fragments

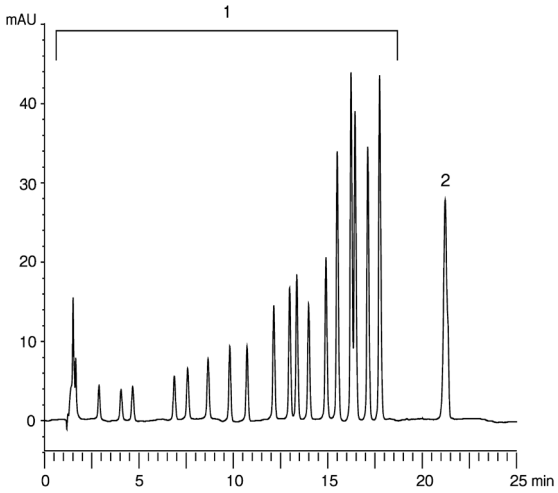
SEC IEX

1. Plasmid pBR322 (4,361 bp)
2. Plasmid pBR322 Hae III digest (8-587 bp)



SEC vs. IEX!

1. Plasmid pBR322 Hae III digest (8-587 bp)
2. Plasmid pBR322 (4,361 bp)



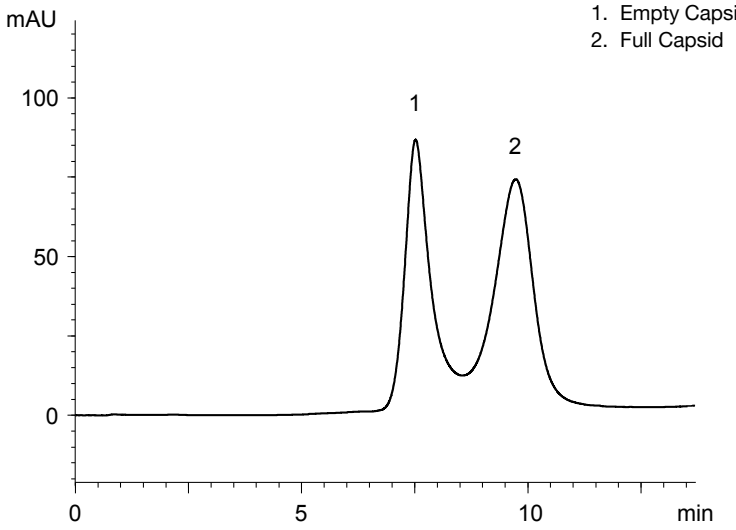
Columns:	YMC-Pack Diol-300 + Diol-200 (5 µm) 500 x 8.0 mm ID
Part Nos.:	DL30S05-5008WT + DL20S05-5008WT
Eluent:	0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate:	0.7 mL/min
Temperature:	ambient (25 °C)
Detection:	UV at 260 nm
Injection:	10 µL

Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID
Part No.:	QF00S05-1046WP
Eluent:	A) 20 mM Tris-HCl (pH 8.1) B) 20 mM Tris-HCl (pH 8.1) containing 1.0 M NaCl
Gradient:	70–85%B (0–20 min), 85%B (20–25 min)
Flow rate:	0.5 mL/min
Temperature:	35 °C
Detection:	UV at 260 nm
Injection:	10 µL

Intact adeno-associated virus

IEX

1. Empty Capsid
2. Full Capsid



Column:	BioPro IEX QF (5 µm) 30 x 4.6 mm ID
Part number:	QF00S05-0346WP
Eluent:	A) 20 mM Bis-trispropane-HCl (pH9.0) B) 20 mM Bis-trispropane-HCl containing 0.5 M $(CH_3)_4NCl$ (pH 9.0)
Gradient:	5%B (0–0.2 min), 20–45%B (0.2–10 min)
Flow rate:	0.5 ml/min
Temperature:	25 °C
Detection:	FLS at Ex. 280 nm, Em. 348 nm
Injection:	2 µL
Sample:	AAV2 (2.59 x 10^{12} vg/mL)

This research was supported by AMED under Grant Number JP18ae0201001.

BioLC applications – AAVs

Denatured adeno-associated viruses

RP

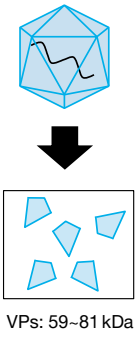
Sample preparation

AAV2
1.46 x 10^{12} vg/mL
AAV5
3.95 x 10^{12} vg/mL

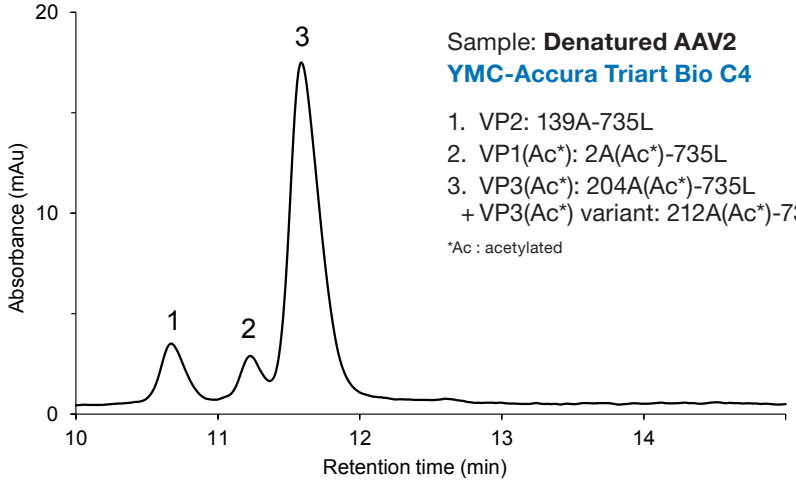
10% acetic acid treatment
RT, 15 min

Centrifuged at 12,000 rpm
5 min

analysis



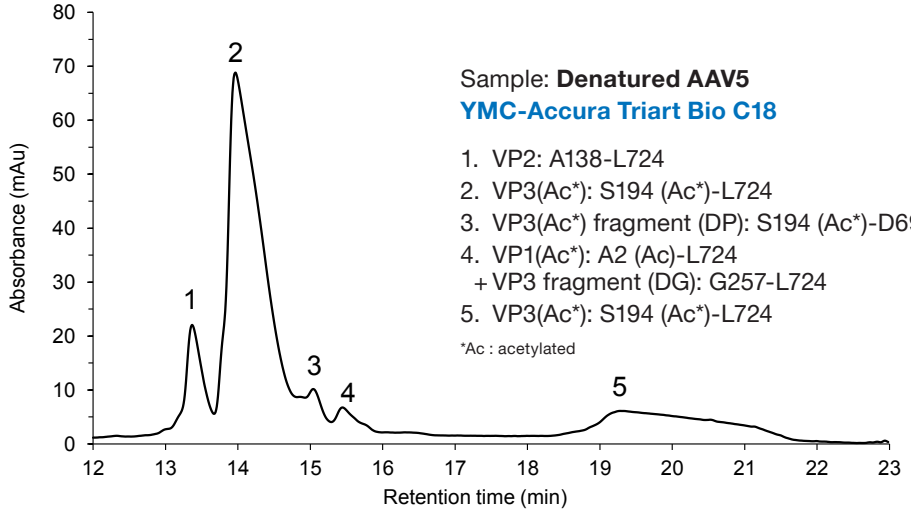
VPs: 59~81 kDa



Sample: **Denatured AAV2**
YMC-Accura Triart Bio C4

1. VP2: 139A-735L
2. VP1(Ac*): 2A(Ac*)-735L
3. VP3(Ac*): 204A(Ac*)-735L
   + VP3(Ac*) variant: 212A(Ac*)-735L

*Ac : acetylated



Sample: **Denatured AAV5**
YMC-Accura Triart Bio C18

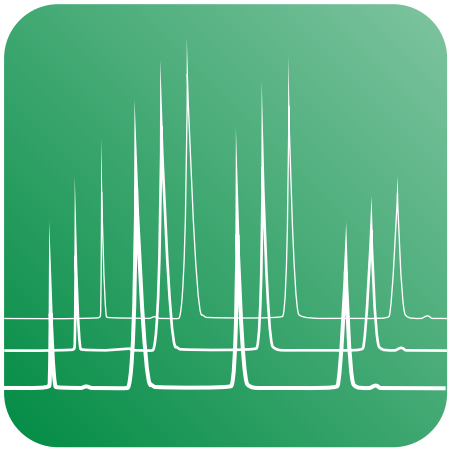
1. VP2: A138-L724
2. VP3(Ac*): S194 (Ac*)-L724
3. VP3(Ac*) fragment (DP): S194 (Ac*)-D695
4. VP1(Ac*): A2 (Ac)-L724
   + VP3 fragment (DG): G257-L724
5. VP3(Ac*): S194 (Ac*)-L724

*Ac : acetylated

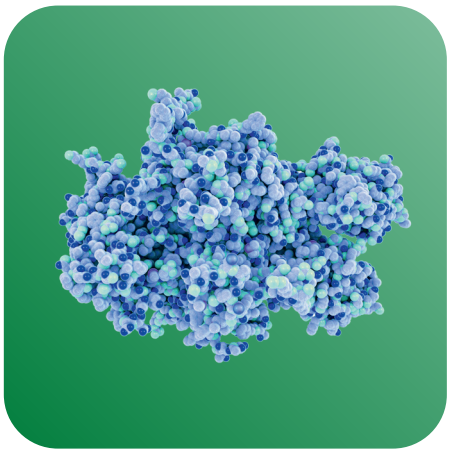
Columns:	YMC-Accura Triart Bio C4 (1.9 µm, 30 nm) 150 x 2.1 mm ID YMC-Accura Triart Bio C18 (1.9 µm, 30 nm) 150 x 2.1 mm ID
Part Nos.:	TB30SP9-15Q1PTC TA30SP9-15Q1PTC
Eluent:	A) water/difluoroaceticacid (100/0.1) B) acetonitrile/difluoroaceticacid(100/0.1)
Gradient:	20–32%B (0–1 min), 32–36%B (1–16 min), 36–80%B (16–20 min)
Flow rate:	0.2 ml/min
Temperature:	80 °C
Detection:	UV at 280 nm ESI-MS (positive ion mode)
Injection:	50 µL

By courtesy of Prof. S. Uchiyama, Osaka University, Japan

This research was supported by AMED under Grant Number JP18ae0201001.



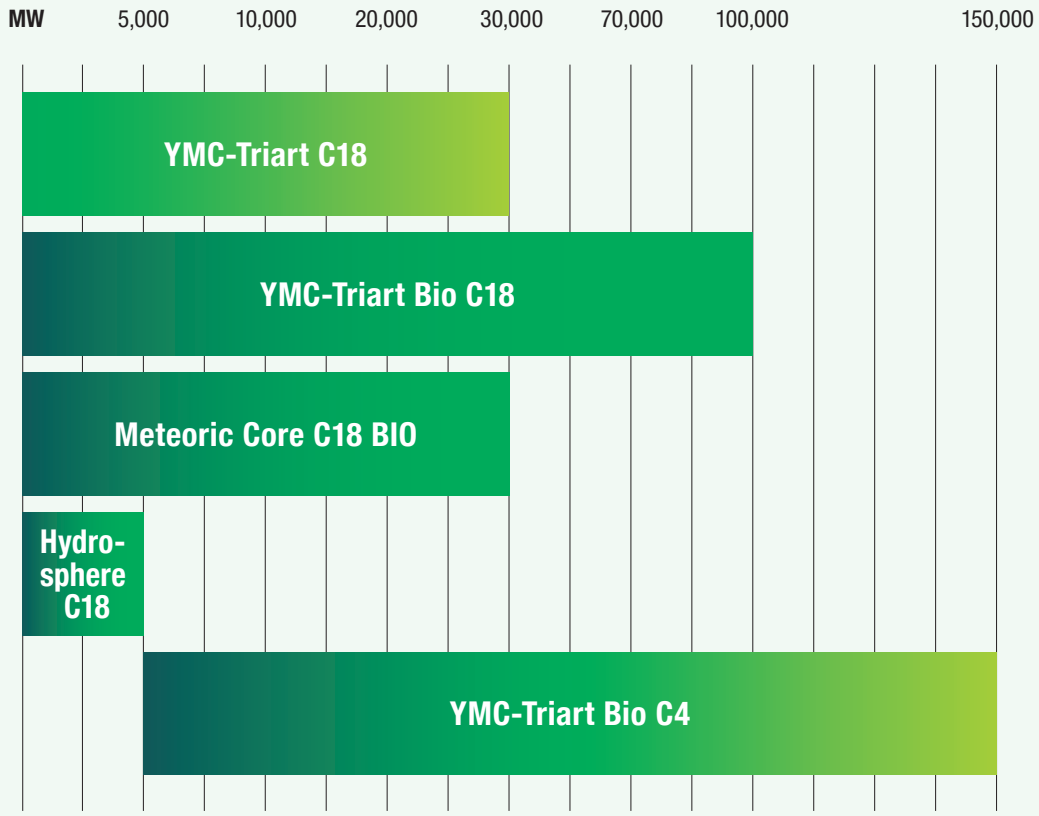
(Bioinert)
RP



RP – Bioseparation Columns

- **Applicable to proteins, antibodies, peptides and oligonucleotides**
- **Selection of C18, C8 and C4 columns**
- **For UHPLC and HPLC**
- **pH- and temperature stable phases**
- **Superior reproducibility**

Column Selection Tool according to molecular weight



most appropriate MW range

extended MW range by elevated temperature

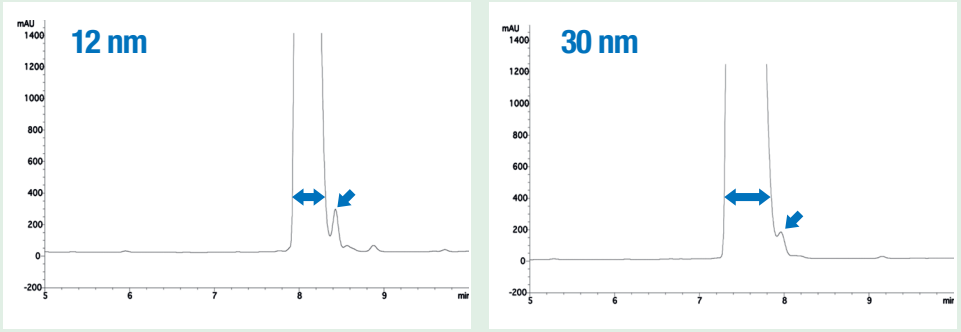
appropriate MW range

Influence of pore size

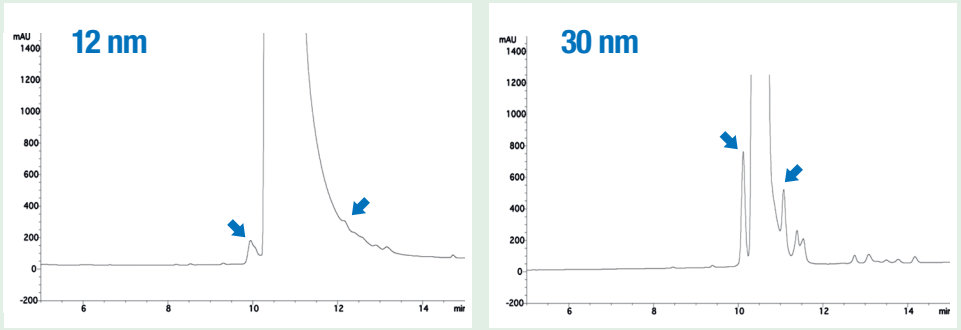
As shown in the table (on the left), the C18 column with 12 nm pore size is suitable for small peptides up to a MW of 5,000 Da. The highest efficiency for large peptides or small proteins can be obtained by using a wide pore C8 phase with 20 nm porosity. Most proteins can be eluted efficiently with a wide pore C4 column with 30 nm porosity.

However, the separation may also be influenced by the hydrophobicity of the peptide/protein and the nature of the column's bonded phase. Therefore, for initial method development, it can be useful, in the first instance, to follow the arrow shown in the ***Column Selection Tool*** for method optimisation.

Angiotensin II
(MW 1,046)



BSA
(MW 67,000)



For smaller peptides a small pore size is more successful. Larger molecules are separated much better with larger pore sizes!

RP – UHPLC / HPLC Selectivities

Selectivities for proteins/peptides and antibodies

	YMC-Triart Bio C4	YMC-Triart C18	YMC-Triart Bio C18	Meteoric Core C18 BIO
Base particle	organic/inorganic hybrid silica			core-shell type silica
Modification	C4 (USP L26)	C18 (USP L1)	C18 (USP L1)	C18 (USP L1)
Particle Size / µm	1.9, 3, 5	1.9, 3, 5	1.9, 3, 5	2.7
Pore Size / nm	30	12	30	16
pH range	1.0–10.0	1.0–12.0	1.0–12.0	1.5–10.0
Temperature range	pH < 7: 90 °C pH > 7: 50 °C	pH < 7: 90 °C pH > 7: 50 °C	pH < 9: 90 °C pH > 9: 50 °C	pH < 7: 70 °C pH > 7: 50 °C

Selectivities for oligonucleotides

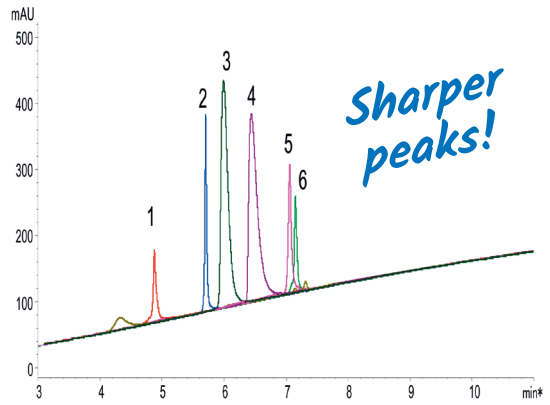
	YMC-Triart C18	YMC-Triart Bio C18	YMC-Triart C8	Hydrosphere C18
Base particle	organic/inorganic hybrid silica			silica
Modification	C18 (USP L1)	C18 (USP L1)	C8 (USP L7)	C18 (USP L1)
Particle Size / µm	1.9, 3, 5	1.9, 3, 5	1.9, 3, 5	2, 3, 5
Pore Size / nm	12	30	12	12
pH range	1.0 – 12.0	1.0–12.0	1.0–12.0	2.0–8.0
Temperature range	pH < 7: 90 °C pH > 7: 50 °C	pH < 9: 90 °C pH > 9: 50 °C	pH < 7: 90 °C pH > 7: 50 °C	50 °C

Bioinert YMC-Triart columns are available for improved sensitivity, peak shape and recovery of coordinating compounds such as nucleotides, oligonucleotides or phosphorylated proteins/peptides.

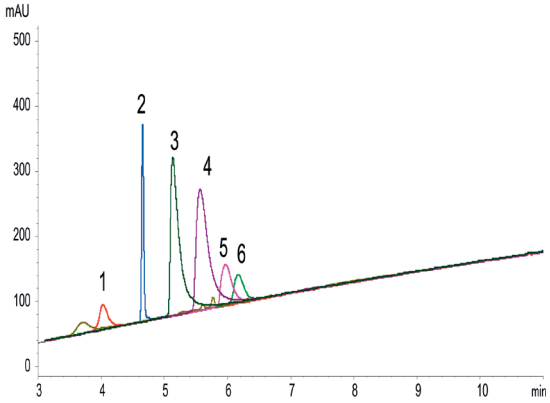
Better performance using YMC-Triart Bio C4

High sensitivity and sharp peaks under LC/MS compatible conditions

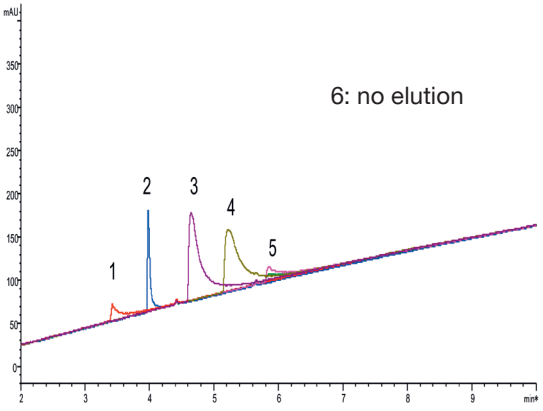
YMC-Triart Bio C4 (3 µm, 30 nm)



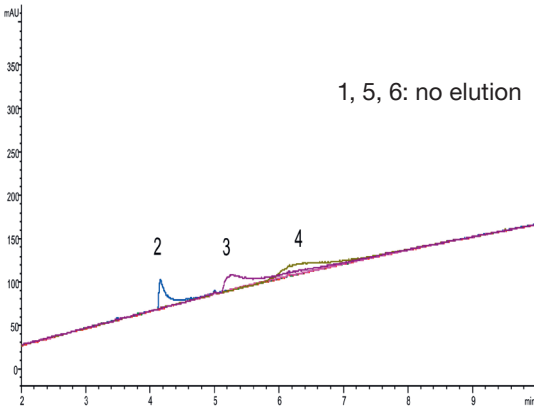
XBridge Protein BEH C4 (3.5 µm, 30 nm)



AdvanceBio RP-mAb C4 (3.5 µm, 45 nm)



Aeris widepore C4 (3.6 µm, 20 nm)



Column:	150 x 3.0 mm ID
Part No.:	TB30S03-1503PTH
Eluent:	A) water/formic acid (100/0.1) B) acetonitrile/formic acid (100/0.1)
Gradient:	10–95%B (0–15 min)
Flow rate:	0.4 mL/min (for 3.0 mm ID) 1.0 mL/min (for 4.6 mm ID)
Temperature:	40 °C

Detection:	UV at 220 nm
Sample:	1. Cytochrome c (Horse heart) 2. Insulin (Bovine pancreas) 3. Transferrin (Human) 4. BSA 5. β-Lactoglobulin (Bovine) 6. α-Chymotrypsinogen A (Bovine pancreas)

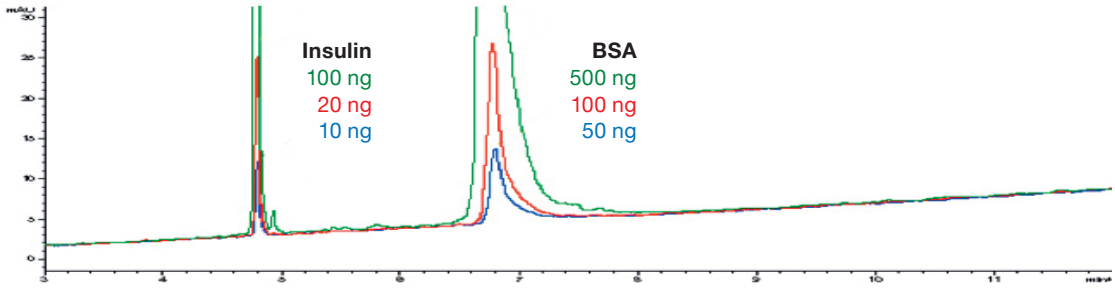
YMC-Triart Bio C4 shows better peak shape and recovery with a mobile phase containing formic acid, which is commonly used for LC/MS analysis. Therefore, YMC-Triart Bio C4 is ideal for high sensitivity analysis of proteins.

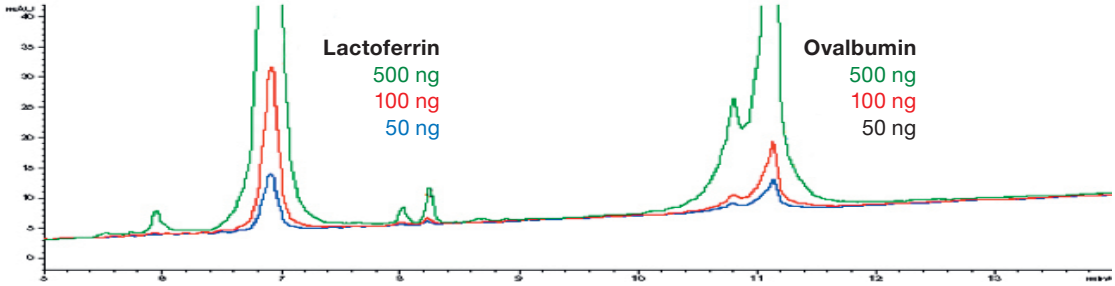
RP – YMC-Triart Bio C4: No column adsorption

No sample adsorption by YMC-Triart Bio C4 columns

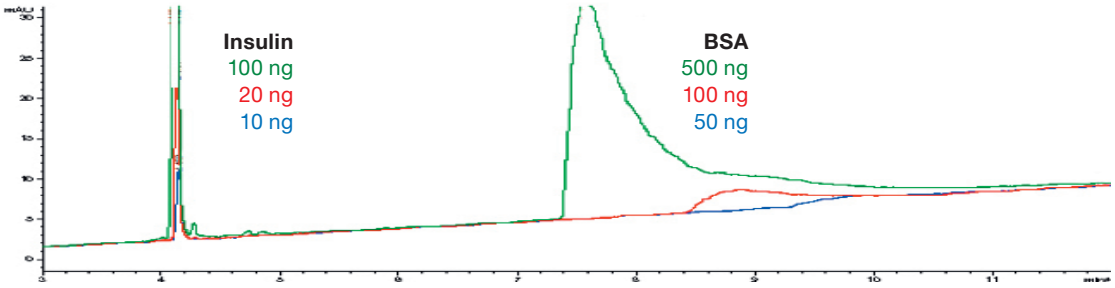
Ideal for Microanalysis

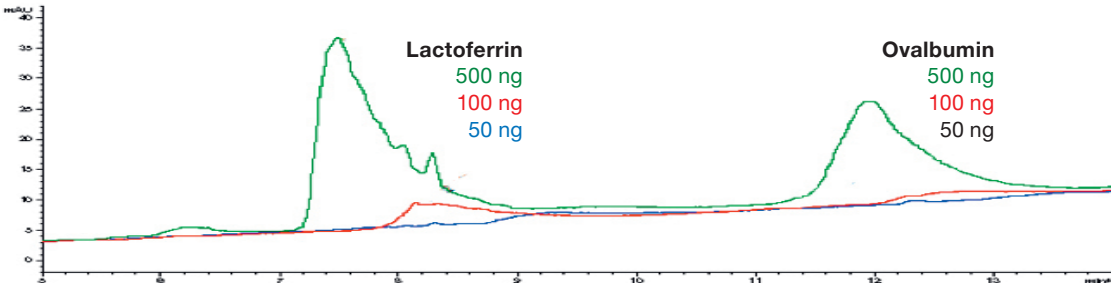
YMC-Triart Bio C4 (1.9 µm, 30 nm)





Aeris widepore C4 (3.6 µm, 20 nm)



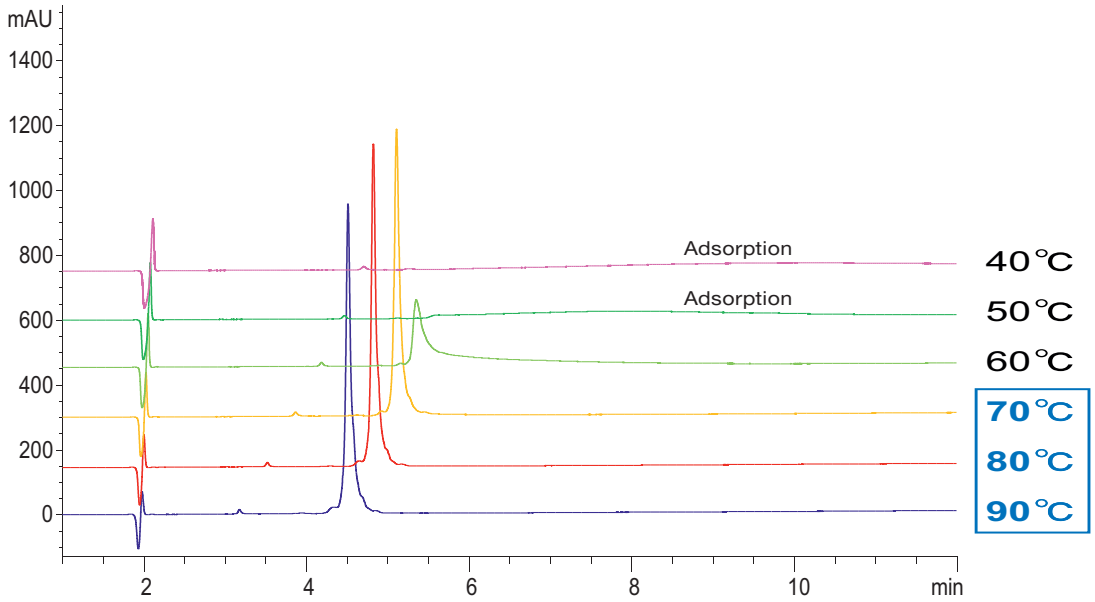


Column:	150 x 2.1 mm ID
Part No.:	TB30SP9-15Q1PT
Eluent:	A) water/TFA (100/0.05)
	B) acetonitrile/TFA (100/0.05)
Gradient:	25-60%B (0-15 min), 90%B (15-20 min), 25%B (20-35 min)
Flow rate:	0.2 mL/min
Temperature:	40 °C
Detection:	UV at 220 nm

No sample adsorption was observed on YMC-Triart Bio C4 even at a low sample loading. This makes YMC-Triart Bio C4 ideal for microanalysis of proteins.

High temperature tolerance allows antibody analysis

Bevacizumab (Avastin®, MW: ca. 148 kDa)



Column:	YMC-Triart Bio C4 (3 µm, 30 nm) 150 x 3.0 mm ID
Part No.:	TB30S03-1503PTH
Eluent:	A) water/TFA (100/0.1) B) acetonitrile/TFA (100/0.1)
Gradient:	30–60%B (0–15 min), 90%B (15–30min)
Flow rate:	0.4 mL/min
Detection:	UV at 220 nm
Injection:	4 µL
Sample:	Bevacizumab (0.5 mg/mL)

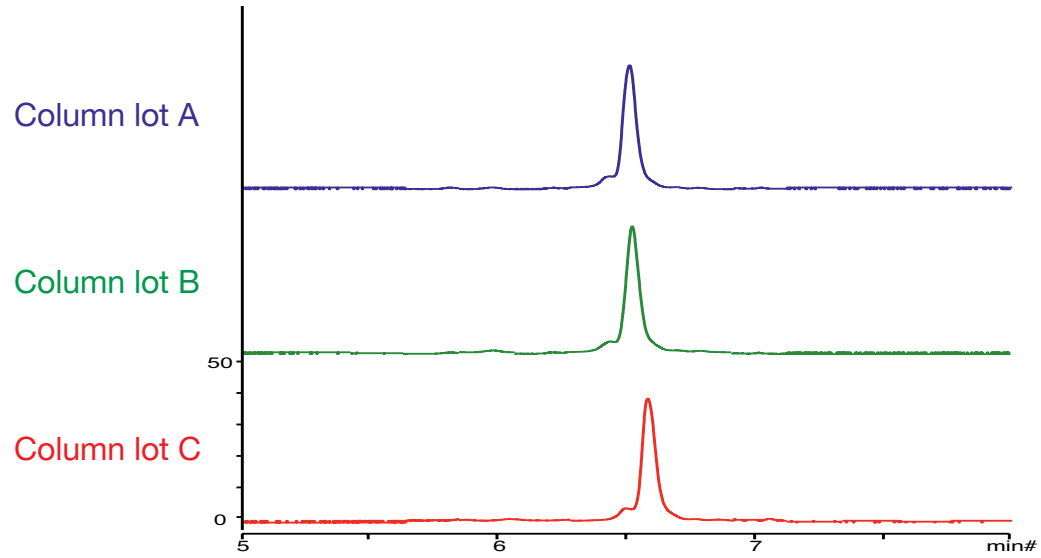
"The possibility to use temperatures up to 90 °C with YMC-Triart Bio C4 simplifies the development of analytical methods. Furthermore, a good peak shape can be obtained without the addition of TFA, which means that I have fewer problems when using it for MS."

Lars M. H. Reinders, Institute for Energy and Environmental Technology e. V. (IUTA, DE)

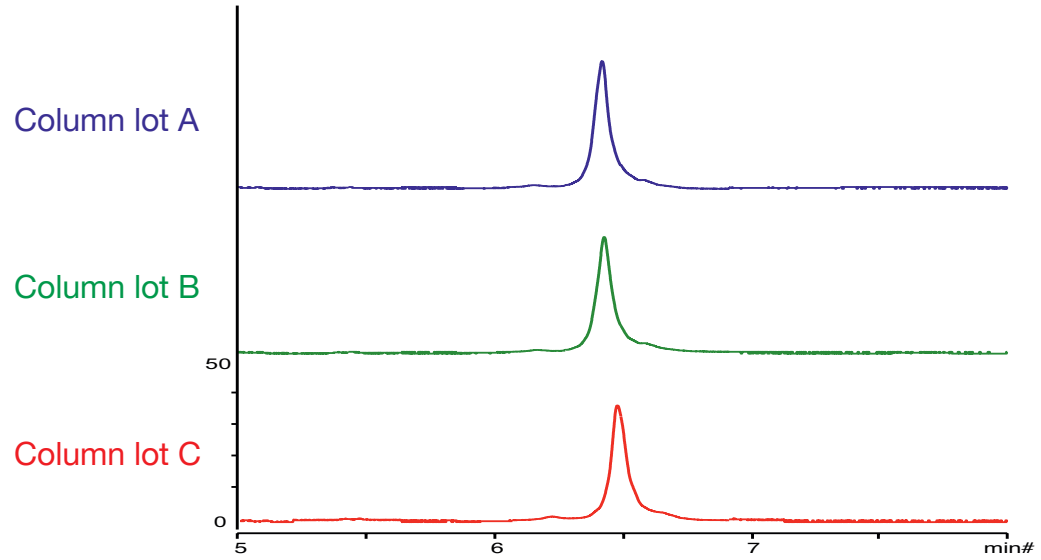
RP – YMC-Triart Bio C4: Reproducibility

Excellent Batch-to-batch reproducibility for antibody analysis

NISTmAb, 8671



Bevacizumab (Avastin®)



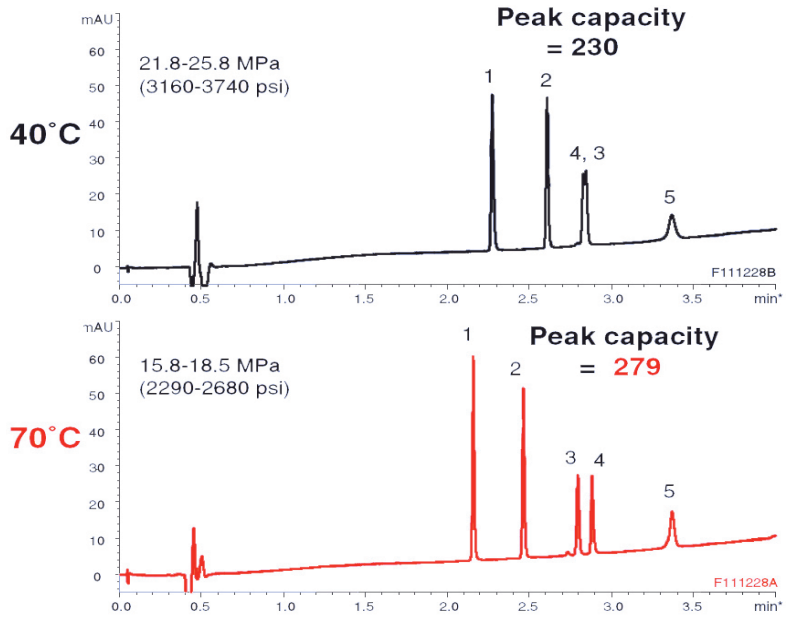
Column:	YMC-Triart Bio C4 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.:	TB30SP9-05Q1PT
Eluent:	A) water/TFA (100/0.1), B) acetonitrile/TFA (100/0.1)
Gradient:	25–45%B (0–10 min)
Flow rate:	0.4 mL/min
Temperature:	80 °C
Detection:	UV at 280 nm
Injection:	2 µL (0.5 mg/mL)

YMC-Triart Bio C4 shows excellent lot-to-lot reproducibility for antibodies. Not only is retention time highly reproducible, but also the resolution of minor impurity peaks. This makes YMC-Triart Bio C4 ideal for quality control of biopharmaceuticals.

More temperature flexiblity using YMC-Triart

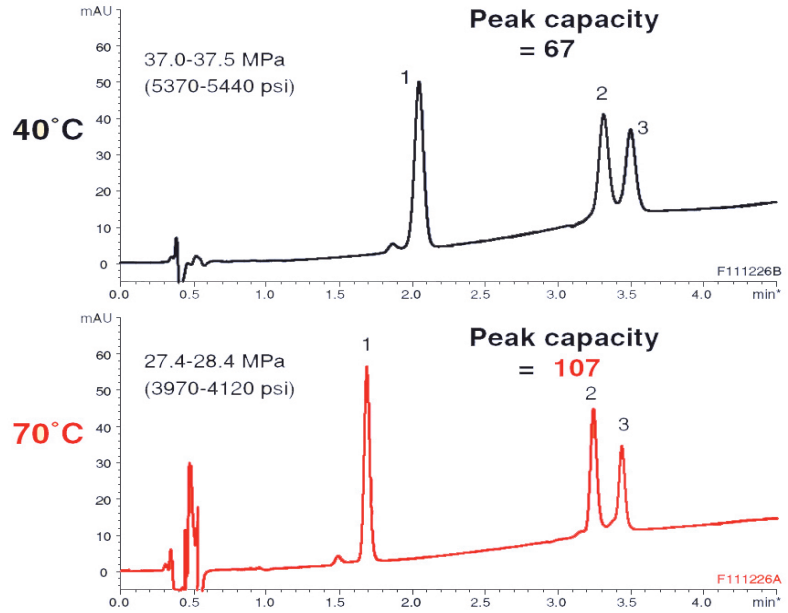
Highly efficient RP-HPLC separation of proteins

Mixture A (MW 500–18,400)



Analytes	MW	Peak width ½h (min)	
		40 °C	70 °C
Mixture A			
1. Oxytocin	1,007	0.017	0.014
2. Leu-Enkephalin	556	0.015	0.015
3. β-Endorphin	3,465	–	0.016
4. Insulin	5,733	–	0.015
5. β-Lactoglobulin A	18,400	0.043	0.030
Mixture B			
1. Lysozyme	14,300	0.069	0.044
2. α-Chymotrypsinogen	25,700	0.080	0.049
3. β-Lactoglobulin A	18,400	0.080	0.048

Mixture B (MW 14,300–25,700)



High temperatures only possible with YMC-Triart

Column: YMC-Triart C18 (1.9 µm, 12 nm) 50 x 2.0 mm ID
Part-No.: TA12SP9-0502WT
Eluent: A) water/TFA (100/0.1)
B) acetonitrile/TFA (100/0.1) - mixture A
B) acetonitrile/2-propanol/TFA (50/50/0.1) - mixture B
Gradient: 10–80%B (0–5 min) - mixture A
30–60%B (0–5 min) - mixture B

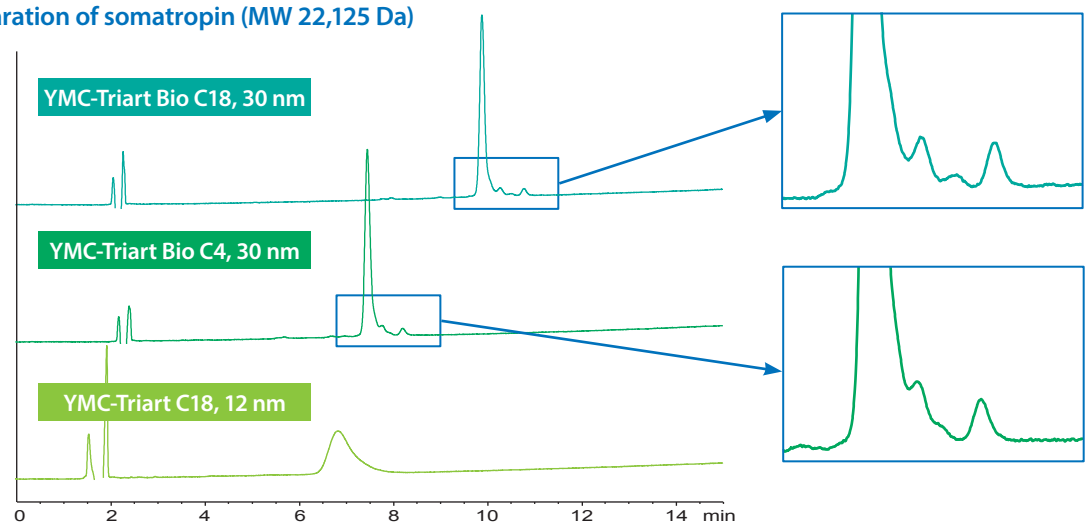
Flow rate: 0.4 mL/min
Detection: UV at 220 nm
Injection: 1 µL (50 µg/mL) - condition A
1 µl (250 µg/mL) - condition B
System: Agilent 1200SL

PC (peak capacity) = 1 + (gradient time / peak width*)
*peak width = $2W_{0.5h}$ average

RP – YMC-Triart Bio C18: Great peak shapes

Ideal solutions for any kind of biomolecule

Separation of somatropin (MW 22,125 Da)

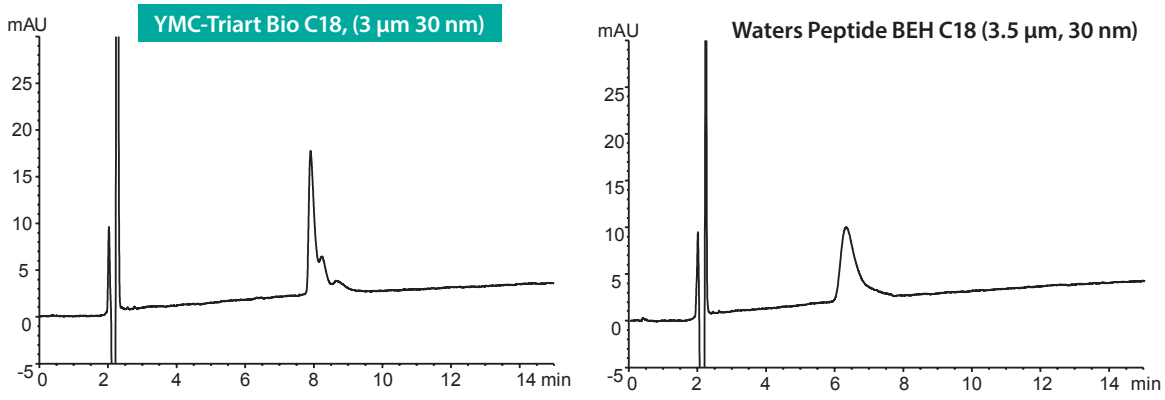


Columns:	150 x 3.0 mm ID (3 µm)	Gradient:	50–70%B (0–15 min)
Part Nos.:	TA30S03-1503PTH TB30S03-1503PTH TA12S03-1503PTH	Flow rate:	0.425 mL/min
		Temperature:	40 °C
		Detection:	UV at 220 nm
Eluent:	A) water/TFA (100/0.1) B) acetonitrile/TFA (100/0.08)	Injection:	4 µL
		Sample:	Somatropin (0.1 mg/mL)

In this example of somatropin, a peptide of 22,125 Da, good peak shape can be obtained with the widepore columns YMC-Triart Bio C18 and YMC-Triart Bio C4. Excellent separation was achieved using YMC-Triart Bio C18 with longer alkyl chains in its bonded phase.

Ideal for MS conditions

Good peak shape with mobile phase containing formic acid



Column:	150 x 3.0 mm ID; 150 x 4.6 mm ID	Flow rate:	0.425 mL/min for 3.0 mm ID; 1.0 mL/min for 4.6 mm ID
Part No.:	TA30S03-1503PTH	Temperature:	40 °C
Eluent:	A) water/formic acid (100/0.1) B) acetonitrile/formic acid (100/0.08)	Detection:	UV at 220 nm
		Sample:	Somatropin (0.1 mg/mL)
Gradient:	45–65%B (0–15 min)		

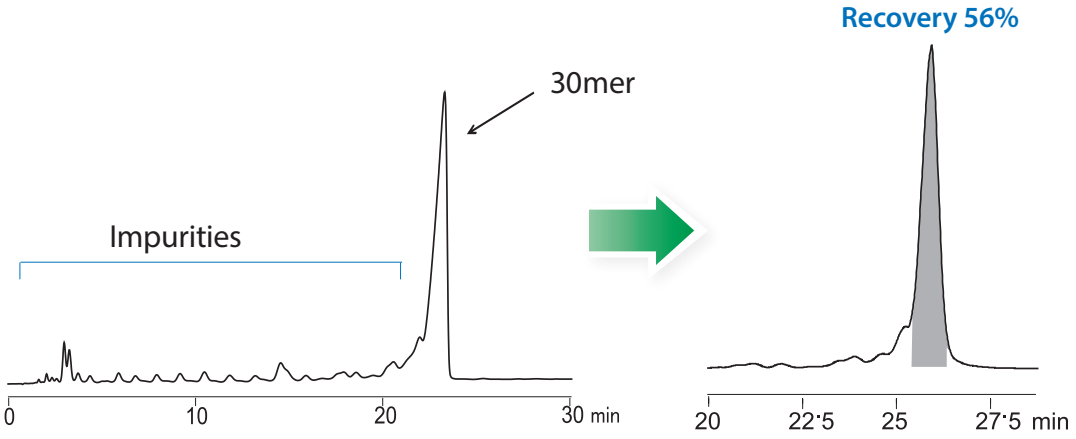
YMC-Triart Bio C18 is suitable for highly sensitive analysis and structural analysis of proteins using LC/MS since good peak shapes in mobile phase containing formic acid can be achieved.

Easy purification of oligonucleotides with YMC-Actus semiprep columns

Purification of synthetic 30mer oligonucleotide

Analysis 1.0 mL/min, 5 µL injection
Hydrosphere C18
50 x 4.6 mm ID, 5 µm

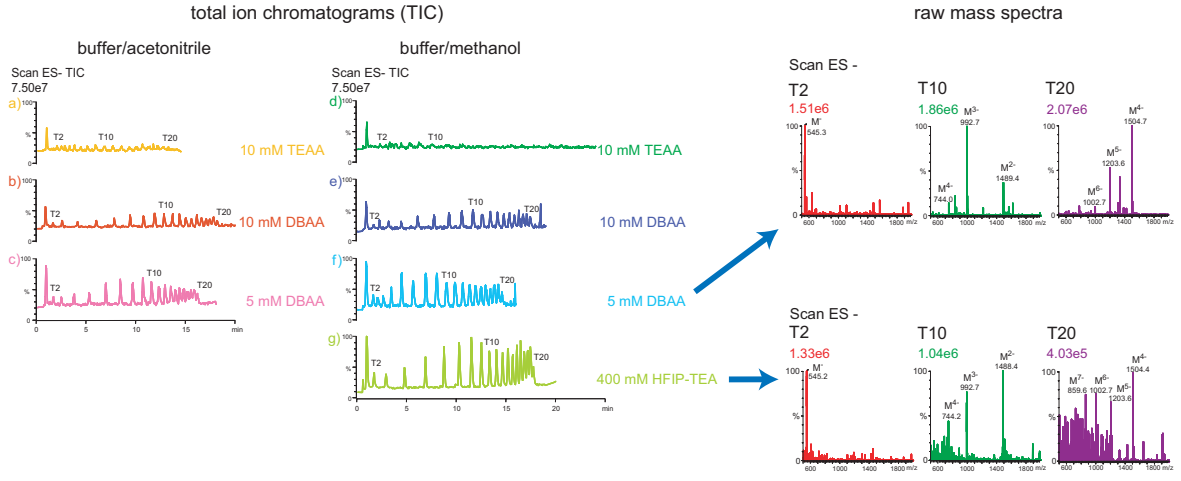
Purification 19 mL/min, 100 µL injection
YMC-Actus Hydrosphere C18
50 x 20 mm ID, 5 µm



Part Nos.:	HS12S05-0546WT HS12S05-0520WX
Eluent:	A) 10 mM DBA-acetic acid (pH 6.0) / methanol (60/40) B) 10 mM DBA-acetic acid (pH 6.0) / methanol (20/80)
Gradient:	10%–35%B (0–30 min.)
Temperature:	ambient
Detection:	UV at 269 nm
Sample:	synthetic oligonucleotide (100 µM)

purity>99%

Influences of mobile phase conditions on intensity of ESI-MS

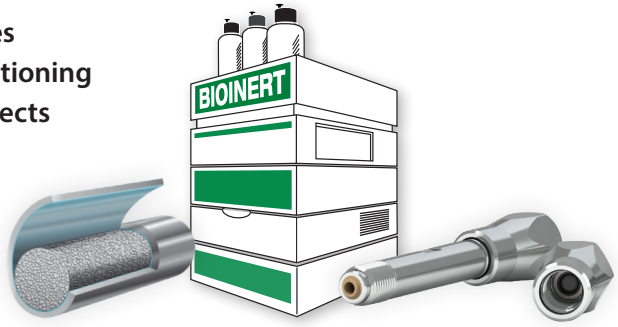


Column:	Hydrosphere C18 (3 µm, 12 nm) 50 x 2.0 mm	
Part No.:	HS12S03-0502WT	
Flow rate:	0.2 mL/min	
Temperature:	35°C	
Detection:	ESI negative mode	
Injection:	5 µL (25 pmol/component)	
a)	Eluent:	A) 10 mM TEAA buffer (pH 6.0) B) eluent A/acetonitrile (80/20)
	Gradient:	50–65%B (0–20 min)
b/c)	Eluent:	A) 10/5 mM DBAA buffer (pH 6.0) B) eluent A/acetonitrile (50/50)
	Gradient:	30–75%B (0–20 min)
d)	Eluent:	A) 10 mM TEAA buffer (pH 6.0) B) eluent A/methanol (50/50)
	Gradient:	44–50%B (0–25 min)
e/f)	Eluent:	A) 10/5 mM DBAA buffer (pH 6.0) B) eluent A/methanol (20/80)
	Gradient:	42–70%B (0–20 min)
g)	Eluent:	A) 400 mM HFIP-TEA buffer (pH 7.0) B) methanol
	Gradient:	7–35%B (0–20 min)
Sample:	Oligodeoxythymidylic acid [d(pT)$_{2\text{-}20}$]	

RP – YMC-Triart: Bioinert hardware

Bioinert columns for bioseparations and coordinating compounds

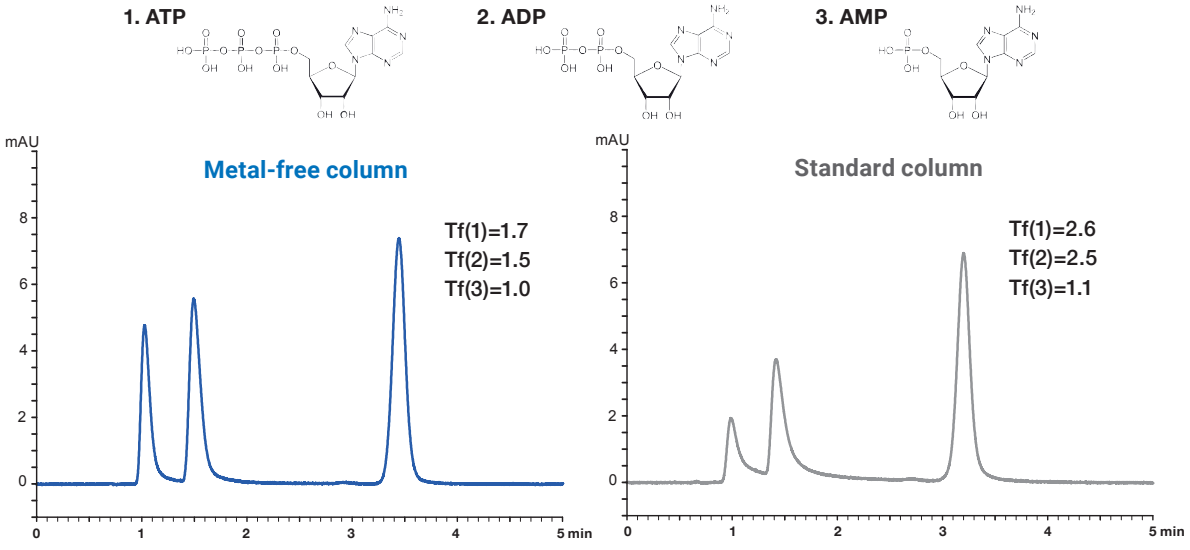
- Exceptional peak shapes with high sensitivities
- Excellent recoveries without column preconditioning
- Superior reproducibility and no carry-over effects
- Ideal for highly sensitive LC/MS analyses
- Different bioinert hardware options



Specification

	YMC-Accura Triart	YMC-Triart PEEK-lined
YMC-Triart modifications	C18, C18 ExRS, Bio C18, C8, Bio C4, Phenyl, PFP, Diol-HILIC	
Particle Size	1.9, 3 and 5 µm	
Column hardware	Bioinert coated stainless steel	PEEK-lined stainless steel
Frit hardware	Bioinert coated stainless steel	PEEK
Hardware properties	Less hydrophobic	More hydrophobic
Pressure limit	1.9 µm: 100 MPa (15,000 psi) 3/5 µm: 45 MPa (6,525 psi)	
Column connection	No special connections required	Selected universal connectors such as MarvelXACT™

Improved sensitivity for coordination compounds



Column: YMC-Triart C18 (3 µm, 12 nm) 50 x 2.1 mm ID
Part Nos.: TA12S03-05Q1PTP (metal-free) or
TA12S03-05Q1PTH (standard hardware)
Eluent: 5 mM $HCOONH_4$
Flow rate: 0.21 mL/min

Temperature: 25 °C
Detection: UV at 265 nm
Injection: 1 µL (10 mg/mL)
System: bioinert/"metal-free" HPLC system

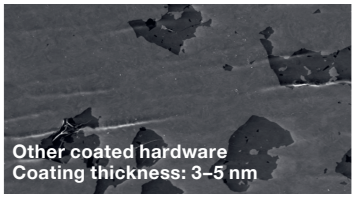
Metal coordinating compounds, which have a phosphate group in their structure, tend to show poor peak shape due to interactions with metals, such as the stainless steel in column bodies and frits. By using a bioinert column hardware, better peak shapes can be expected.
Nucleotides with phosphate groups also show better peak shapes when compared to the regular column hardware.
The applied YMC-Triart PEEK-lined as well as the YMC-Accura Triart column hardware are ideal for highly sensitive analyses using LC/MS.

YMC-Accura Triart: durable bioinert coating

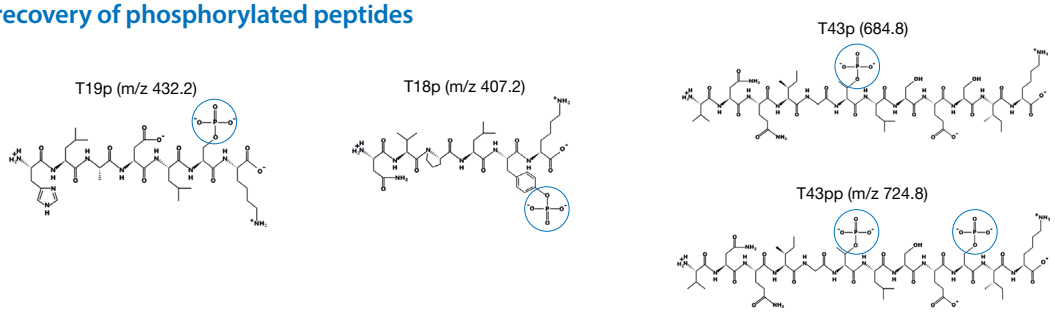
The robust bioinert coating used on YMC-Accura hardware is 130 to 320-fold thicker making it more durable than other similar hardware concepts. A long-term inertness against sensitive substances is ensured. In order to demonstrate its robustness, a YMC-Accura column was packed multiple times. Even though this is quite a challenge for the column surface, the coating remains unaffected (SEM* picture: top area is bare steel for comparison).

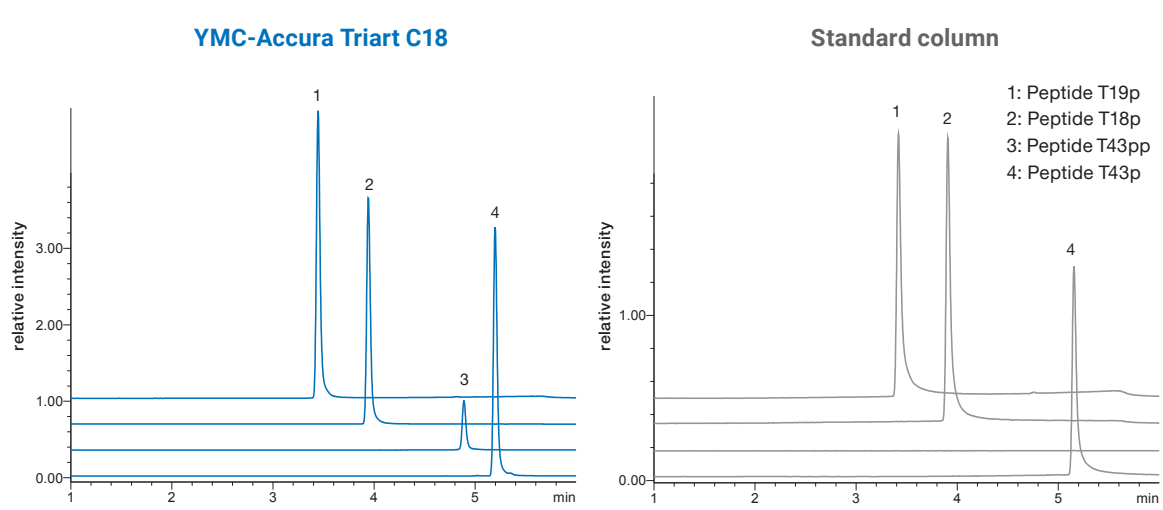
*Scanning Electron Microscope

Other coated columns can lose their inertness over time. This will again lead to adsorption of sensitive compounds on the uncovered metallic surfaces. Peak tailing, loss of recovery and sample carry-over are typical results of the delamination of the coating. After only unpacking a coated competitor column most of the coating is already delaminated (dark spots: remaining coating).



Full recovery of phosphorylated peptides





Columns:	YMC-Accura Triart C18 (1.9 µm, 30 nm) 100 x 2.1 mm ID (bioinert hardware) YMC-Triart C18 (1.9 µm, 30 nm) 100 x 2.1 mm ID (standard hardware)	Gradient:	0.7%–25%B (0–5 min), 25%B (5–6.6 min), 0.7%B (6.6–8 min)
Part Nos.:	TA12SP9-10Q1PTC TA12SP9-10Q1PT	Flow rate:	0.6 ml/min
Eluent:	A) water + 0.1% formic acid B) acetonitrile + 0.1% formic acid	Temperature:	60 °C
		Detection:	ESI-MS
		Injection:	2 µl (10 pmol/µl)
		Sample:	Massprep phosphopeptide enolase standard (Waters)
		System:	Shimadzu Nexera XS inert Shimadzu LCMS-2020

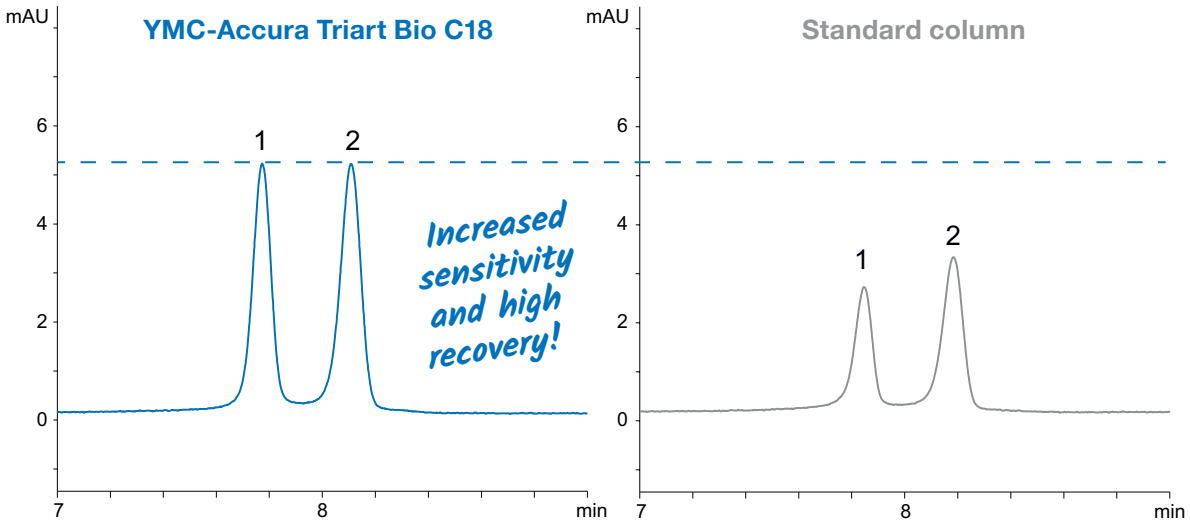
By courtesy of Shimadzu Europa.

The use of a bioinert coated YMC-Accura Triart C18 column led to higher intensities and peak areas of four phosphopeptides, compared to the stainless steel column. The high recovery rate of the YMC-Accura Triart C18 column also enabled the detection of the challenging phosphopeptide T43pp, which contains two phosphate residues. In contrast, detection of peptide T43pp was unsuccessful with the standard column, even after ten injections no signal was observed.

RP – YMC-Triart: Bioinert hardware

Significantly higher sensitivity and recovery

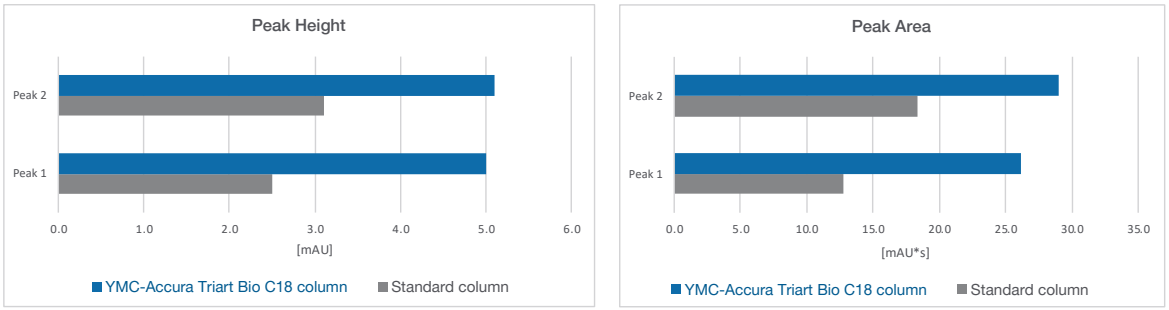
Ideal choice for challenging analytes such as phosphorothioate oligonucleotides



Column: YMC-Accura Triart Bio C18 (1.9 µm, 30 nm) 50 x 2.1 mm ID
Part No.: TA30SP9-05Q1PTC
Eluent: A) 15 mM triethylamine - 400 mM HFIP*
B) methanol
Gradient: 8–18%B (0–10 min)
Flow rate: 0.42 mL/min
Temperature: 65 °C
Detection: UV at 260 nm
Injection: 1 µL
Sample: All PS RNA 20mer (1) (5'-U^C^A^U^C^A^C^A^C^U^G^A^A^U^A^C^C^A^A^U-3')
All PS RNA 21mer (2) (5'-G^U^C^A^U^C^A^C^A^C^U^G^A^A^U^A^C^C^A^A^U-3')
^=Phosphorothioate

*1,1,1,3,3,3-hexafluoro-2-propanol

High sensitivity and recovery

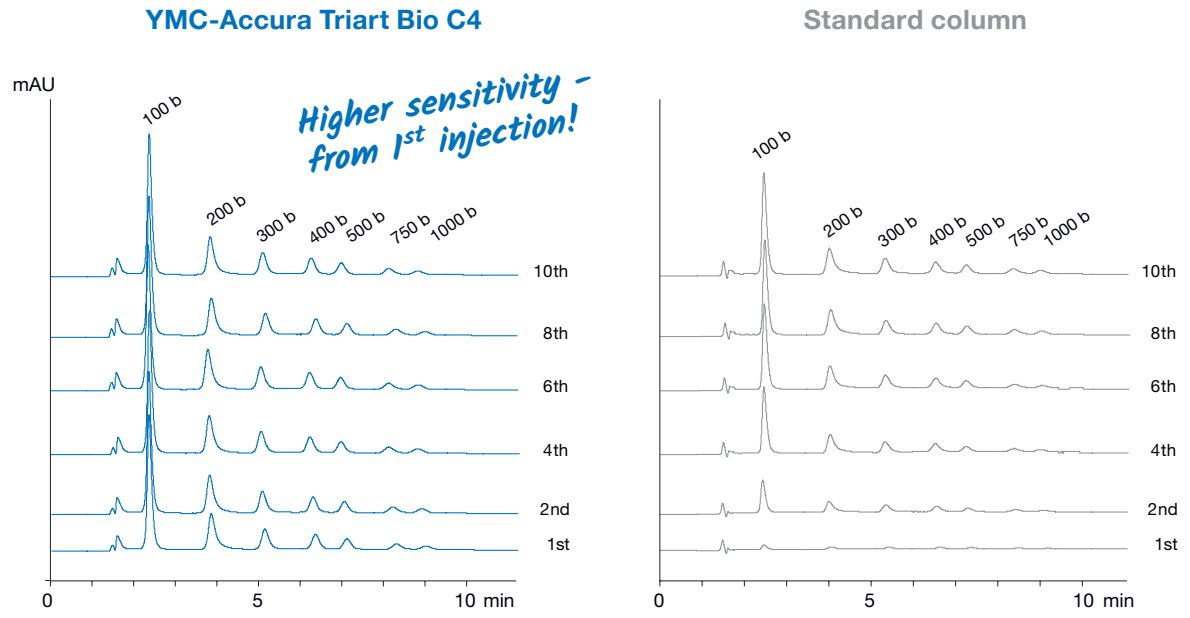


Doubled peak height and area!

The YMC-Accura Triart Bio C18 column provides double peak heights and peak areas for the oligonucleotides compared to those for regular stainless-steel columns. YMC-Accura Triart columns enhance the sensitivity significantly and help to save precious samples without any loss.

Reliable results from the first injection

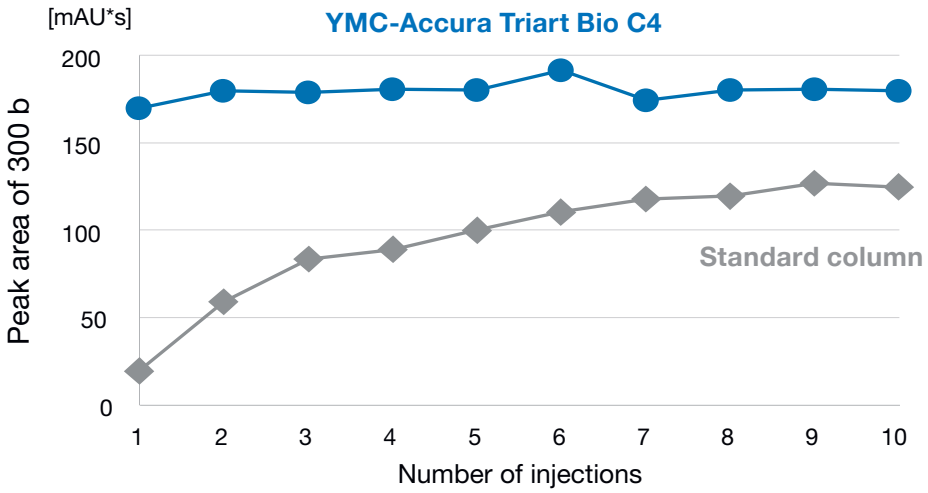
No preconditioning required for reliable results



Column:	YMC-Accura Triart Bio C4 (1.9 µm, 30 nm) 100 x 2.1 mm ID	Flow rate:	0.2 mL/min
Part No.:	TA30SP9-10Q1PTC	Temperature:	80°C
Eluent:	A) 50 mM TEAA* (pH 7.0)/acetonitrile (95/5)	Detection:	UV at 254 nm
	B) 50 mM TEAA (pH 7.0)/acetonitrile (50/50)	Injection:	1 µL (0.25 mg/mL)
Gradient:	9–14%B (0–10 min), 80%B (10–15 min)	Sample:	100–1,000 bases (Century™-Plus RNA Markers)

* Triethylammonium acetate

Constantly higher peak areas and therefore recoveries

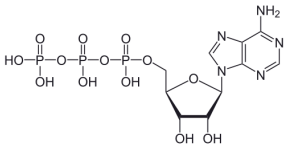


The YMC-Accura Triart Bio C4 column shows stable peak areas from the first injection, while the standard stainless-steel column provides only 10% of the peak area (for the 300 base marker) with the first injection. Even after the tenth injection, the peak areas of the stainless-steel column are considerably less than those of the YMC-Accura Triart column.

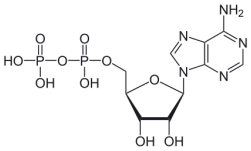
RP – Expert Tips: (Oligo)nucleotides

Influence of system and column hardware on the analysis of nucleotides

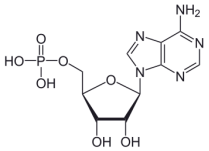
1 ATP



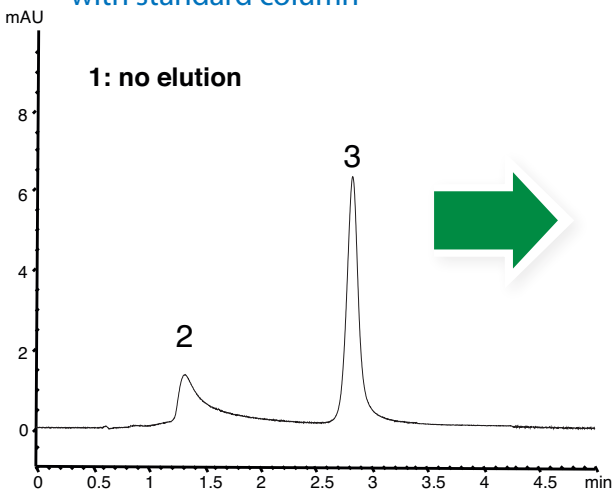
2 ADP



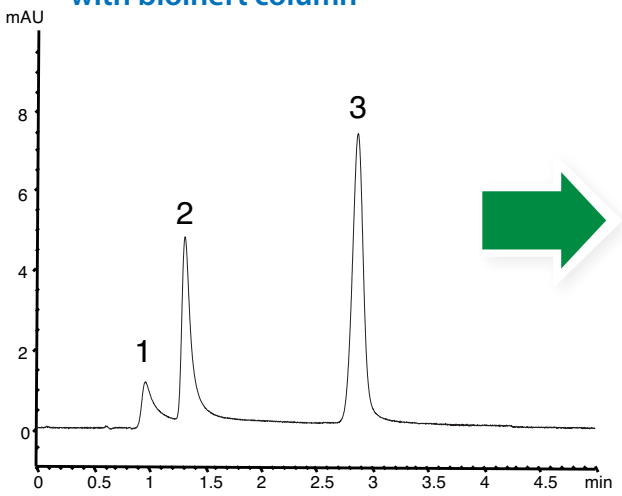
3 AMP

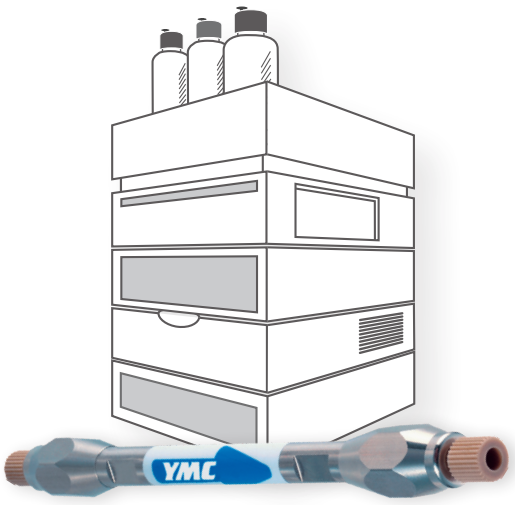


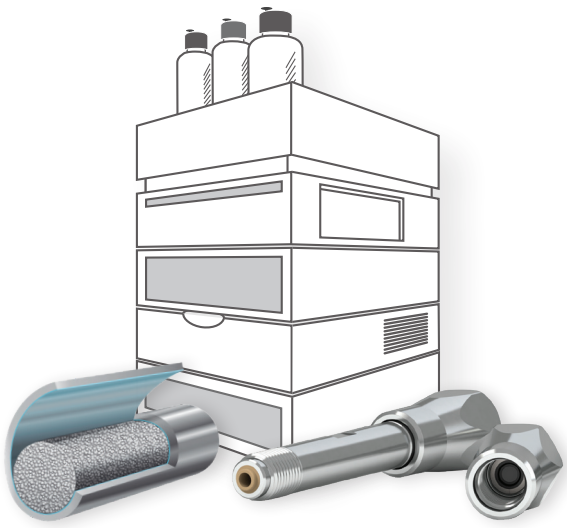
Ordinary HPLC system with standard column



Ordinary HPLC system with bioinert column





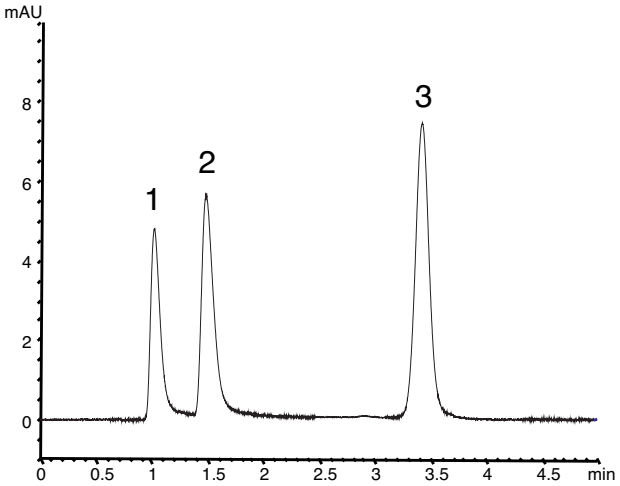


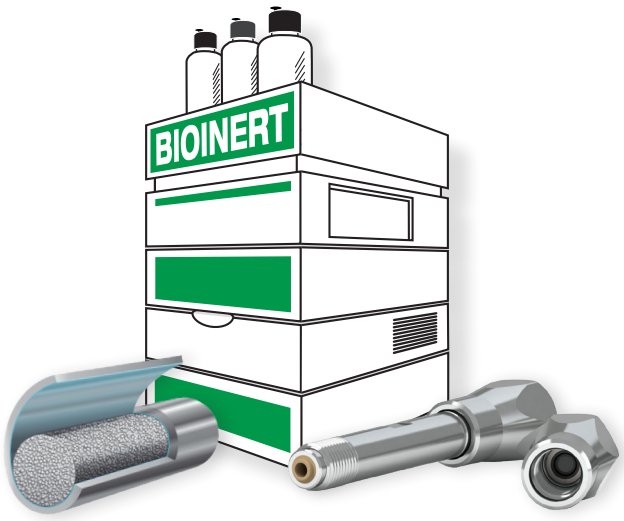
Column:	YMC-Triart C18 (3 µm, 12 nm) 50 x 2.1 mm ID
Part Nos:	TA12S03-05Q1PT (standard hardware)
	TA12S03-05Q1PTP (**bioinert** hardware)
Eluent:	5 mM $HCOONH_4$
Flow rate:	0.21 mL/min
Temperature:	25°C
Detection:	UV at 265 nm
Injection:	1 µL (10 µg/mL)

*Bioinert HPLC system: PEEK sample loop, PEEK injector port, and PEEK tubing are used.

ATP peak is detected, and peak shape of ADP is improved as a result of using a bioinert column.

Bioinert HPLC system* with bioinert column





"Metal-free YMC columns significantly reduce non-specific adsorption phenomena"

"YMC-Triart C18 metal-free columns significantly reduce non-specific adsorption phenomena during peptides analysis. We use these columns in our laboratory for a specific application. We obtain very good chromatographic resolution and excellent robustness, which is very appreciable during routine analysis."

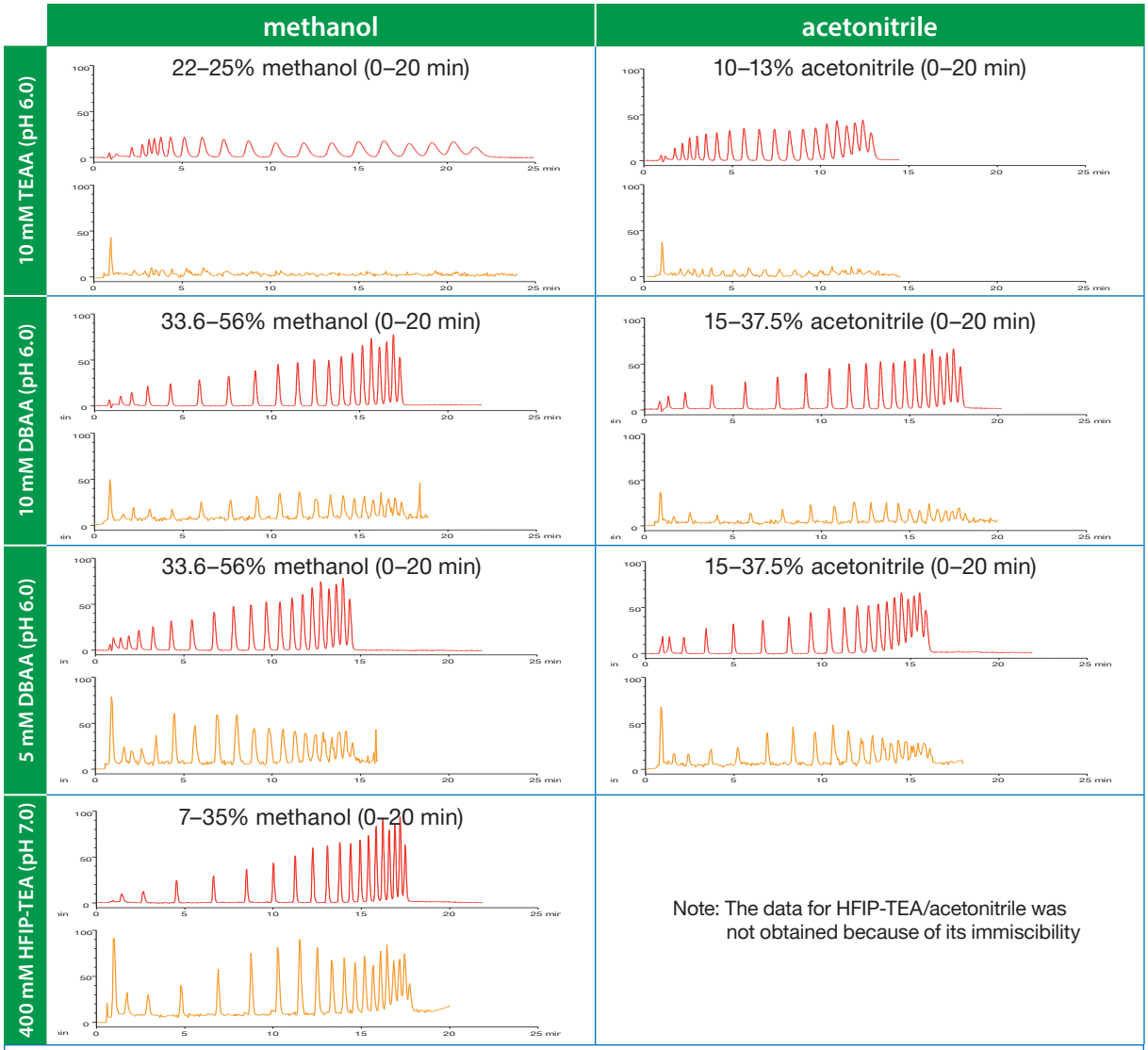
Cynthia Mongongu, LADF,
Laboratoire AntiDopage Français,
Université Paris-Saclay (FR)

Peak shape is greatly improved as a result of using a bioinert HPLC system.

RP – Expert Tips: Oligonucleotides

Effect of composition and salt concentration of ion-pairing mobile phase on the separation and signal intensity

Comparison of separation and ESI-MS signal intensity using different ion-pairing buffers and organic solvents



Column:	Hydrosphere C18 (3.0 µm, 12 nm) 50 x 2.0 mm ID
Part No.:	HS12S03-0502WT
Flow rate:	0.2 mL/min
Temperature:	35 °C
Detection:	UV at 269 nm (red), ESI negative mode (orange)
Injection:	5 µl (25 pmol/mL)
Sample:	Oligo(deoxythymidylic acid) [dp (dT) 2-20]

eluent abbreviations

TEAA:	triethylammonium acetate
DBAA:	di-n-butylammonium acetate
HFIP:	1,1,1,3,3,3-hexafluoro-2-propanol
TEA:	triethylamine

The mobile phase composition has different effects on the separation and signal intensity in electrospray ionisation mass spectrometry (ESI-MS) of oligonucleotides. Using different gradient conditions, acceptable retention and resolution can be achieved (upper UV chromatograms; red trace) for each separation by optimising the gradient slope of the organic solvent regardless of the type of mobile phase. The ESI-MS intensity is significantly influenced by the type and concentration of ion-pairing buffer as shown in the lower MS chromatograms (orange trace). HFIP-TEA buffer/methanol systems provide the maximum MS intensity. Enhanced retention and MS intensity are obtained using 10 mM DBAA buffer compared to 10 mM TEAA buffer, and the lower DBAA concentration results in approximately 1.5–3 times increase in the intensity without any change in the concentration of organic solvent.

1.9 µm UHPLC columns (max. pressure 100 MPa)

Phase	Column ID [mm]	Column length [mm]					Guard cartridges* with 5 mm length
		30	50	75	100	150	(pack of 3)
YMC-Triart C18	1.0	—	TA12SP9-0501WT	—	TA12SP9-1001WT	TA12SP9-1501WT	TA12SP9-E5Q1CC**
	2.0	TA12SP9-0302PT	TA12SP9-0502PT	TA12SP9-L502PT	TA12SP9-1002PT	TA12SP9-1502PT	TA12SP9-E5Q1CC**
	2.1	TA12SP9-03Q1PT	TA12SP9-05Q1PT	TA12SP9-L5Q1PT	TA12SP9-10Q1PT	TA12SP9-15Q1PT	TA12SP9-E5Q1CC**
	3.0	—	TA12SP9-0503PT	TA12SP9-L503PT	TA12SP9-1003PT	TA12SP9-1503PT	TA12SP9-E503CC
YMC-Triart Bio C18	2.0	TA30SP9-0302PT	TA30SP9-0502PT	TA30SP9-L502PT	TA30SP9-1002PT	TA30SP9-1502PT	TA30SP9-E5Q1CC**
	2.1	TA30SP9-03Q1PT	TA30SP9-05Q1PT	TA30SP9-L5Q1PT	TA30SP9-10Q1PT	TA30SP9-15Q1PT	TA30SP9-E5Q1CC**
	3.0	—	TA30SP9-0503PT	TA30SP9-L503PT	TA30SP9-1003PT	TA30SP9-1503PT	TA30SP9-E503CC
YMC-Triart C8	2.0	TO12SP9-0302PT	TO12SP9-0502PT	TO12SP9-L502PT	TO12SP9-1002PT	TO12SP9-1502PT	TO12SP9-E5Q1CC**
	2.1	TO12SP9-03Q1PT	TO12SP9-05Q1PT	TO12SP9-L5Q1PT	TO12SP9-10Q1PT	TO12SP9-15Q1PT	TO12SP9-E5Q1CC**
	3.0	—	TO12SP9-0503PT	TO12SP9-L503PT	TO12SP9-1003PT	TO12SP9-1503PT	TO12SP9-E503CC
YMC-Triart Bio C4	2.0	TB30SP9-0302PT	TB30SP9-0502PT	TB30SP9-L502PT	TB30SP9-1002PT	TB30SP9-1502PT	TB30SP9-E5Q1CC**
	2.1	TB30SP9-03Q1PT	TB30SP9-05Q1PT	TB30SP9-L5Q1PT	TB30SP9-10Q1PT	TB30SP9-15Q1PT	TB30SP9-E5Q1CC**
	3.0	—	TB30SP9-0503PT	TB30SP9-L503PT	TB30SP9-1003PT	TB30SP9-1503PT	TB30SP9-E503CC

*Guard cartridge holder required, part no. XPCHUHP
**Guard cartridge: 2.1 mm ID

1.9 µm bioinert coated UHPLC columns (max. pressure 100 MPa)

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Accura Triart C18	2.1	TA12SP9-05Q1PTC	TA12SP9-100Q1PTC	TA12SP9-15Q1PTC
YMC-Accura Triart Bio C18	2.1	TA30SP9-05Q1PTC	TA30SP9-10Q1PTC	TA30SP9-15Q1PTC
YMC-Accura Triart C8	2.1	TO30SP9-05Q1PTC	TO12SP9-10Q1PTC	TO12SP9-15Q1PTC
YMC-Accura Triart Bio C4	2.1	TB30SP9-05Q1PTC	TB30SP9-10Q1PTC	TB30SP9-15Q1PTC

1.9 µm PEEK-lined UHPLC columns (max. pressure 100 MPa)

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18 metal-free	2.1	TA12SP9-05Q1PTP	TA12SP9-10Q1PTP	TA12SP9-15Q1PTP
YMC-Triart Bio C18 metal-free	2.1	TA30SP9-05Q1PTP	TA30SP9-10Q1PTP	TA30SP9-15Q1PTP
YMC-Triart C8 metal-free	2.1	TO12SP9-05Q1PTP	TO12SP9-10Q1PTP	TO12SP9-15Q1PTP
YMC-Triart Bio C4 metal-free	2.1	TB30SP9-05Q1PTP	TB30SP9-10Q1PTP	TB30SP9-15Q1PTP

Special column connectors required.

RP – Ordering information

3 µm HPLC columns (max. pressure 25–45 MPa)

Phase	Column ID [mm]	Column length [mm]						Guard cartridges* with 10 mm length
		30/33	50	75	100	150	250	(pack of 5)
YMC-Triart C18	2.1	TA12S03-H3Q1PTH	TA12S03-05Q1PTH	TA12S03-L5Q1PTH	TA12S03-10Q1PTH	TA12S03-15Q1PTH	–	TA12S03-01Q1GC
	3.0	–	TA12S03-0503PTH	TA12S03-L503PTH	TA12S03-1003PTH	TA12S03-1503PTH	–	TA12S03-0103GC
	4.6	TA12S03-H346PTH	TA12S03-0546PTH	TA12S03-L546PTH	TA12S03-1046PTH	TA12S03-1546PTH	TA12S03-2546PTH	TA12S03-0104GC
YMC-Triart Bio C18	2.1	TA30S03-H3Q1PTH	TA30S03-05Q1PTH	TA30S03-L5Q1PTH	TA30S03-10Q1PTH	TA30S03-15Q1PTH	–	TA30S03-01Q1GC
	3.0	–	TA30S03-0503PTH	TA30S03-L503PTH	TA30S03-1003PTH	TA30S03-1503PTH	–	TA30S03-0103GC
	4.6	TA30S03-H346PTH	TA30S03-0546PTH	TA30S03-L546PTH	TA30S03-1046PTH	TA30S03-1546PTH	TA30S03-2546PTH	TA30S03-0104GC
YMC-Triart C8	2.1	TO12S03-H3Q1PTH	TO12S03-05Q1PTH	TO12S03-L5Q1PTH	TO12S03-10Q1PTH	TO12S03-15Q1PTH	–	TO12S03-01Q1GC
	3.0	–	TO12S03-0503PTH	TO12S03-L503PTH	TO12S03-1003PTH	TO12S03-1503PTH	–	TO12S03-0103GC
	4.6	TO12S03-H346PTH	TO12S03-0546PTH	TO12S03-L546PTH	TO12S03-1046PTH	TO12S03-1546PTH	TO12S03-2546PTH	TO12S03-0104GC
YMC-Triart Bio C4	2.1	TB30S03-03Q1PTH	TB30S03-05Q1PTH	TB30S03-L5Q1PTH	TB30S03-10Q1PTH	TB30S03-15Q1PTH	–	TB30S03-01Q1GC
	3.0	–	TB30S03-0503PTH	TB30S03-L503PTH	TB30S03-1003PTH	TB30S03-1503PTH	–	TB30S03-0103GC
	4.6	TB30S03-0346PTH	TB30S03-0546PTH	TB30S03-L546PTH	TB30S03-1046PTH	TB30S03-1546PTH	TB30S03-2546PTH	TB30S03-0104GC
Hydrosphere C18	2.1	HS12S03-03Q1WT	HS12S03-05Q1WT	HS12S03-L5Q1WT	HS12S03-10Q1WT	HS12S03-15Q1WT	HS12S03-25Q1WT	HS12S03-01Q1GC
	3.0	HS12S03-0303WT	HS12S03-0503WT	HS12S03-L503WT	HS12S03-1003WT	HS12S03-1503WT	HS12S03-2503WT	HS12S03-0103GC
	4.6	HS12S03-0346WT	HS12S03-0546WT	HS12S03-L546WT	HS12S03-1046WT	HS12S03-1546WT	HS12S03-2546WT	HS12S03-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm bioinert coated HPLC columns (max. pressure 45 MPa)

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Accura Triart C18	2.1	TA12S03-05Q1PTC	TA12S03-10Q1PTC	TA12S03-15Q1PTC
	4.6	TA12S03-0546PTC	TA12S03-1046PTC	TA12S03-1546PTC
YMC-Accura Triart Bio C18	2.1	TA30S03-05Q1PTC	TA30S03-10Q1PTC	TA30S03-15Q1PTC
	4.6	TA30S03-0546PTC	TA30S03-1046PTC	TA30S03-1546PTC
YMC-Accura Triart C8	2.1	TO12S03-05Q1PTC	TO12S03-10Q1PTC	TO12S03-15Q1PTC
	4.6	TO12S03-0546PTC	TO12S03-1046PTC	TO12S03-1546PTC
YMC-Accura Triart Bio C4	2.1	TB30S03-05Q1PTC	TB30S03-10Q1PTC	TB30S03-15Q1PTC
	4.6	TB30S03-0546PTC	TB30S03-1046PTC	TB30S03-1546PTC

3 µm PEEK-lined HPLC columns (max. pressure 45 MPa)

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18 metal-free	2.1	TA12S03-05Q1PTP	TA12S03-10Q1PTP	TA12S03-15Q1PTP
	4.6	TA12S03-0546PTP	TA12S03-1046PTP	TA12S03-1546PTP
YMC-Triart Bio C18 metal-free	2.1	TA30S03-05Q1PTP	TA30S03-10Q1PTP	TA30S03-15Q1PTP
	4.6	TA30S03-0546PTP	TA30S03-1046PTP	TA30S03-1546PTP
YMC-Triart C8 metal-free	2.1	TO12S03-05Q1PTP	TO12S03-10Q1PTP	TO12S03-15Q1PTP
	4.6	TO12S03-0546PTP	TO12S03-1046PTP	TO12S03-1546PTP
YMC-Triart Bio C4 metal-free	2.1	TB30S03-05Q1PTP	TB30S03-10Q1PTP	TB30S03-15Q1PTP
	4.6	TB30S03-0546PTP	TB30S03-1046PTP	TB30S03-1546PTP

Special column connectors required.

2.7 µm Core-Shell columns (max. pressure 60 MPa)

Phase	Column ID [mm]	Column length [mm]					Precolumn filter 0.5 µm
		30	50	75	100	150	(pack of 5)
Meteoric Core C18 BIO	2.1	CAW16SQ7-03Q1PT	CAW16SQ7-05Q1PT	CAW16SQ7-L5Q1PT	CAW16SQ7-10Q1PT	CAW16SQ7-15Q1PT	XRPRCS35
	3.0	CAW16SQ7-0303PT	CAW16SQ7-0503PT	CAW16SQ7-L503PT	CAW16SQ7-1003PT	CAW16SQ7-1503PT	
	4.6	CAW16SQ7-0346PT	CAW16SQ7-0546PT	CAW16SQ7-L546PT	CAW16SQ7-1046PT	CAW16SQ7-1546PT	

*Holder required, part no. XRPRCS03

5 µm HPLC columns (max. pressure 25–45 MPa)

Phase	Column ID [mm]	Column length [mm]						Guard cartridges* with 10 mm length
		30/33	50	75	100	150	250	(pack of 5)
YMC-Triart C18	2.1	TA12S05-H3Q1PTH	TA12S05-05Q1PTH	TA12S05-L5Q1PTH	TA12S05-10Q1PTH	TA12S05-15Q1PTH	–	TA12S05-01Q1GC
	3.0	–	TA12S05-0503PTH	TA12S05-L503PTH	TA12S05-1003PTH	TA12S05-1503PTH	–	TA12S05-0103GC
	4.6	TA12S05-H346PTH	TA12S05-0546PTH	TA12S05-L546PTH	TA12S05-1046PTH	TA12S05-1546PTH	TA12S05-2546PTH	TA12S05-0104GC
YMC-Triart Bio C18	2.1	TA30S05-H3Q1PTH	TA30S05-05Q1PTH	TA30S05-L5Q1PTH	TA30S05-10Q1PTH	TA30S05-15Q1PTH	–	TA30S05-01Q1GC
	3.0	–	TA30S05-0503PTH	TA30S05-L503PTH	TA30S05-1003PTH	TA30S05-1503PTH	–	TA30S05-0103GC
	4.6	TA30S05-H346PTH	TA30S05-0546PTH	TA30S05-L546PTH	TA30S05-1046PTH	TA30S05-1546PTH	TA30S05-2546PTH	TA30S05-0104GC
YMC-Triart C8	2.1	TO12S05-H3Q1PTH	TO12S05-05Q1PTH	TO12S05-L5Q1PTH	TO12S05-10Q1PTH	TO12S05-15Q1PTH	–	TO12S05-01Q1GC
	3.0	–	TO12S05-0503PTH	TO12S05-L503PTH	TO12S05-1003PTH	TO12S05-1503PTH	–	TO12S05-0103GC
	4.6	TO12S05-H346PTH	TO12S05-0546PTH	TO12S05-L546PTH	TO12S05-1046PTH	TO12S05-1546PTH	TO12S05-2546PTH	TO12S05-0104GC
YMC-Triart Bio C4	2.1	TB30S05-H3Q1PTH	TB30S05-05Q1PTH	TB30S05-L5Q1PTH	TB30S05-10Q1PTH	TB30S05-15Q1PTH	–	TB30S05-01Q1GC
	3.0	–	TB30S05-0503PTH	TB30S05-L503PTH	TB30S05-1003PTH	TB30S05-1503PTH	–	TB30S05-0103GC
	4.6	TB30S05-H346PTH	TB30S05-0546PTH	TB30S05-L546PTH	TB30S05-1046PTH	TB30S05-1546PTH	TB30S05-2546PTH	TB30S05-0104GC
Hydrosphere C18	2.1	HS12S05-03Q1WT	HS12S05-05Q1WT	HS12S05-L5Q1WT	HS12S05-10Q1WT	HS12S05-15Q1WT	HS12S05-25Q1WT	HS12S05-01Q1GC
	3.0	HS12S05-0303WT	HS12S05-0503WT	HS12S05-L503WT	HS12S05-1003WT	HS12S05-1503WT	HS12S05-2503WT	HS12S05-0103GC
	4.6	HS12S05-0346WT	HS12S05-0546WT	HS12S05-L546WT	HS12S05-1046WT	HS12S05-1546WT	HS12S05-2546WT	HS12S05-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

5 µm bioinert coated HPLC columns (max. pressure 45 MPa)

Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Accura Triart C18	2.1	TA12S05-05Q1PTC	TA12S05-10Q1PTC	TA12S05-15Q1PTC
	4.6	TA12S05-0546PTC	TA12S05-1046PTC	TA12S05-1546PTC
YMC-Accura Triart Bio C18	2.1	TA30S05-05Q1PTC	TA30S05-10Q1PTC	TA30S05-15Q1PTC
	4.6	TA30S05-0546PTC	TA30S05-1046PTC	TA30S05-1546PTC
YMC-Accura Triart C8	2.1	TO12S05-05Q1PTC	TO12S05-10Q1PTC	TO12S05-15Q1PTC
	4.6	TO12S05-0546PTC	TO12S05-1046PTC	TO12S05-1546PTC
YMC-Accura Triart Bio C4	2.1	TB30S05-05Q1PTC	TB30S05-10Q1PTC	TB30S05-15Q1PTC
	4.6	TB30S05-0546PTC	TB30S05-1046PTC	TB30S05-1546PTC

RP – Ordering information

5 µm PEEK-lined HPLC columns (max. pressure 45 MPa)

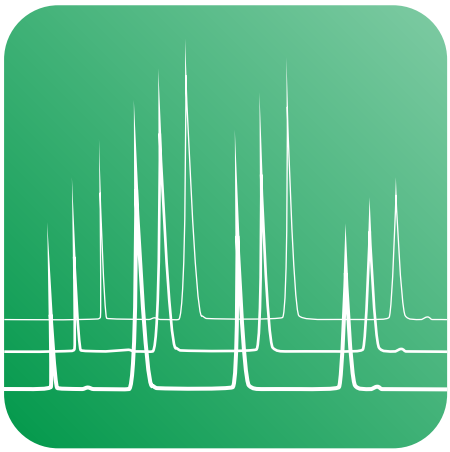
Phase	Column ID [mm]	Column length [mm]		
		50	100	150
YMC-Triart C18 metal-free	2.1	TA12S05-05Q1PTP	TA12S05-10Q1PTP	TA12S05-15Q1PTP
	4.6	TA12S05-0546PTP	TA12S05-1046PTP	TA12S05-1546PTP
YMC-Triart Bio C18 metal-free	2.1	TA30S05-05Q1PTP	TA30S05-10Q1PTP	TA30S05-15Q1PTP
	4.6	TA30S05-0546PTP	TA30S05-1046PTP	TA30S05-1546PTP
YMC-Triart C8 metal-free	2.1	TO12S05-05Q1PTP	TO12S05-10Q1PTP	TO12S05-15Q1PTP
	4.6	TO12S05-0546PTP	TO12S05-1046PTP	TO12S05-1546PTP
YMC-Triart Bio C4 metal-free	2.1	TB30S05-05Q1PTP	TB30S05-10Q1PTP	TB30S05-15Q1PTP
	4.6	TB30S05-0546PTP	TB30S05-1046PTP	TB30S05-1546PTP

Special column connectors required.

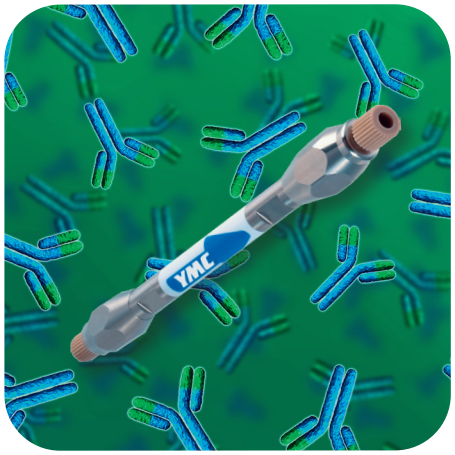
5 µm YMC-Actus high-throughput (semi)preparative columns (max. pressure 20–30 MPa)

Phase	Column ID [mm]	Column length [mm]					Guard cartridges* with 10 mm length
		50	75	100	150	250	(pack of 5)
YMC-Triart C18	20	TA12S05-0520WX	TA12S05-L520WX	TA12S05-1020WX	TA12S05-1520WX	TA12S05-2520WX	TA12S05-0120CCN
	30	TA12S05-0530WX	TA12S05-L530WX	TA12S05-1030WX	TA12S05-1530WX	TA12S05-2530WX	TA12S05-0130CCN
	50	TA12S05-0553DX	–	TA12S05-1053DX	TA12S05-1553DX	TA12S05-2553DX	TA12S05-0553DXG**
YMC-Triart Bio C18	20	TA30S05-0520WX	TA30S05-L520WX	TA30S05-1020WX	TA30S05-1520WX	TA30S05-2520WX	TA30S05-0120CCN
	30	TA30S05-0530WX	TA30S05-L530WX	TA30S05-1030WX	TA30S05-1530WX	TA30S05-2530WX	TA30S05-0130CCN
	50	TA30S05-0553DX	–	TA30S05-1053DX	TA30S05-1553DX	TA30S05-2553DX	TA30S05-0553DXG**
YMC-Triart C8	20	TO12S05-0520WX	TO12S05-L520WX	TO12S05-1020WX	TO12S05-1520WX	TO12S05-2520WX	TO12S05-0120CCN
	30	TO12S05-0530WX	TO12S05-L530WX	TO12S05-1030WX	TO12S05-1530WX	TO12S05-2530WX	TO12S05-0130CCN
	50	TO12S05-0553DX	–	TO12S05-1053DX	TO12S05-1553DX	TO12S05-2553DX	TO12S05-0553DXG**
YMC-Triart Bio C4	20	TB30S05-0520WX	TB30S05-L520WX	TB30S05-1020WX	TB30S05-1520WX	TB30S05-2520WX	TB30S05-0120CCN
	30	TB30S05-0530WX	TB30S05-L530WX	TB30S05-1030WX	TB30S05-1530WX	TB30S05-2530WX	TB30S05-0130CCN
	50	TB30S05-0553DX	–	TB30S05-1053DX	TB30S05-1553DX	TB30S05-2553DX	TB30S05-0553DXG**
Hydrosphere C18	20	HS12S05-0520WX	HS12S05-L520WX	HS12S05-1020WX	HS12S05-1520WX	HS12S05-2520WX	HS12S05-0120CCN
	30	HS12S05-0530WX	HS12S05-L530WX	HS12S05-1030WX	HS12S05-1530WX	HS12S05-2530WX	HS12S05-0130CCN
	50	HS12S05-0553DX	–	HS12S05-1053DX	HS12S05-1553DX	HS12S05-2553DX	–

*Guard cartridge holder required, part no. XPGHF2P20ID (20 mm ID)
XPGHF2P30ID (30 mm ID)
no holder required for 50 mm



SEC



SEC – UHPLC / HPLC Selectivities

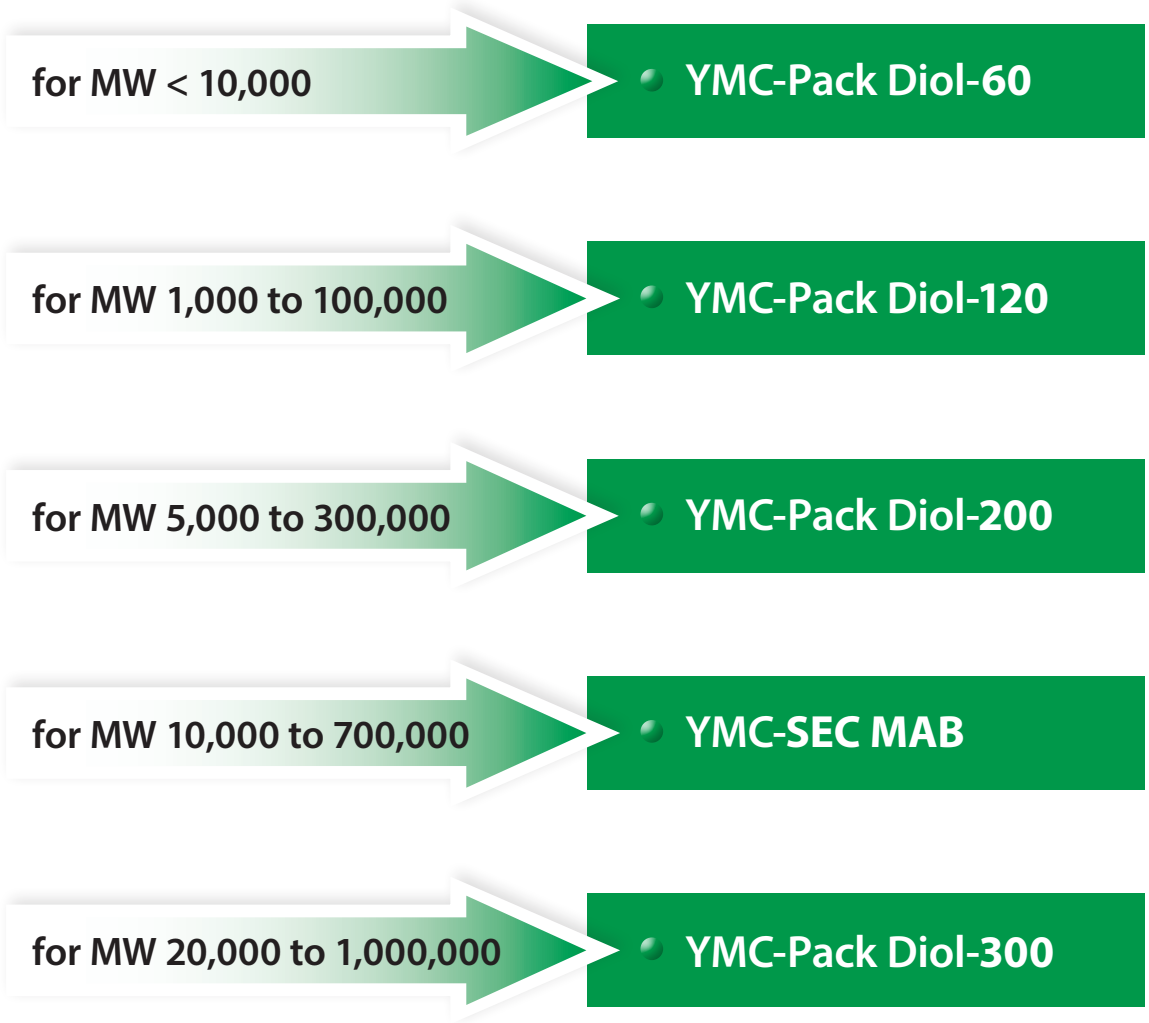
- **Applicable to proteins, antibodies, their fragments and peptides**
- **Also applicable to carbohydrates and nucleic acid components**
- **Excellent reproducibility with minimal secondary interactions**
- **2 µm for UHPLC**
- **Cost effective**

	YMC-Pack Diol-60	YMC-Pack Diol-120	YMC-Pack Diol-200	YMC-Pack Diol-300	YMC-SEC MAB
	For peptides and small proteins	For intermediate proteins	For large proteins	For very large proteins	For antibodies, fragments and aggregates
Base particle	Silica				
Particle Size / µm	3, 5	3, 5	2, 3, 5	2, 3, 5	3
Pore Size / nm	6	12	20	30	25
Modification	Dihydroxypropyl				
Temperature range	40 °C				
Pressure limit	2 µm: 45 MPa (6,525 psi); 3/5 µm: 20 MPa (3,000 psi)				14 MPa (2,030 psi)

"The YMC-Pack Diol SEC column has been successfully used for subsequent method validation."

Rubén Pedrosa Segon, Head of Quality Control Pharmaceutical Department, OFICE S.L. (ES)

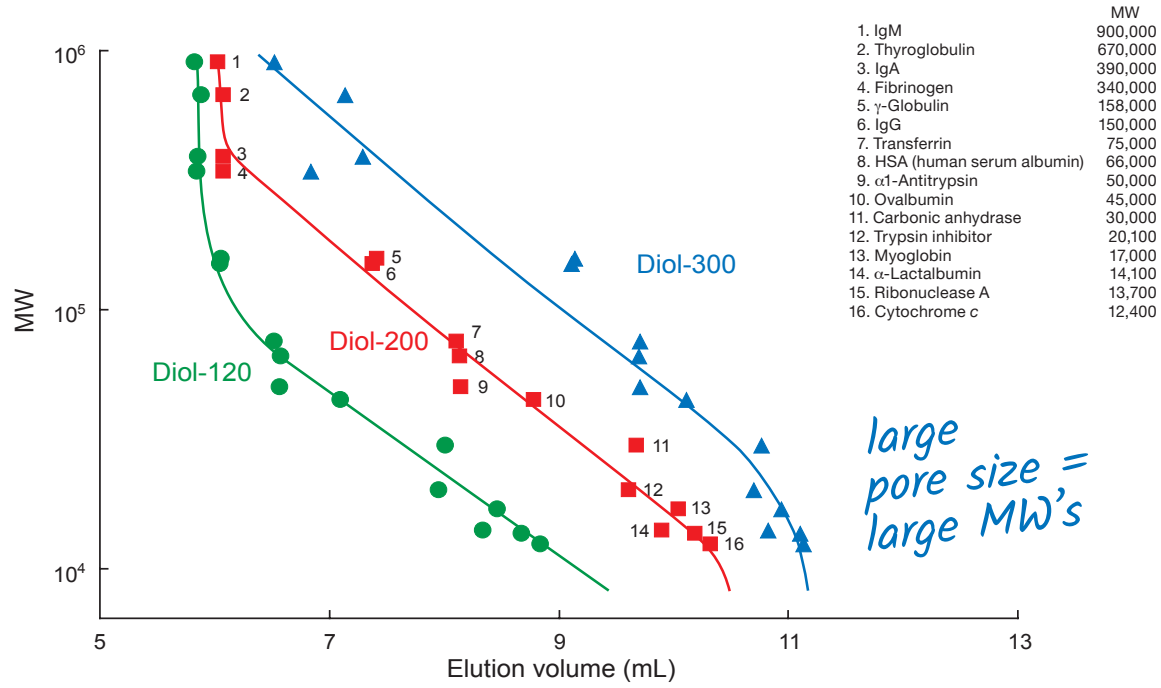
Column Selection Tool



SEC – YMC-Pack Diol: Phase selection for proteins

Phases for different MW ranges

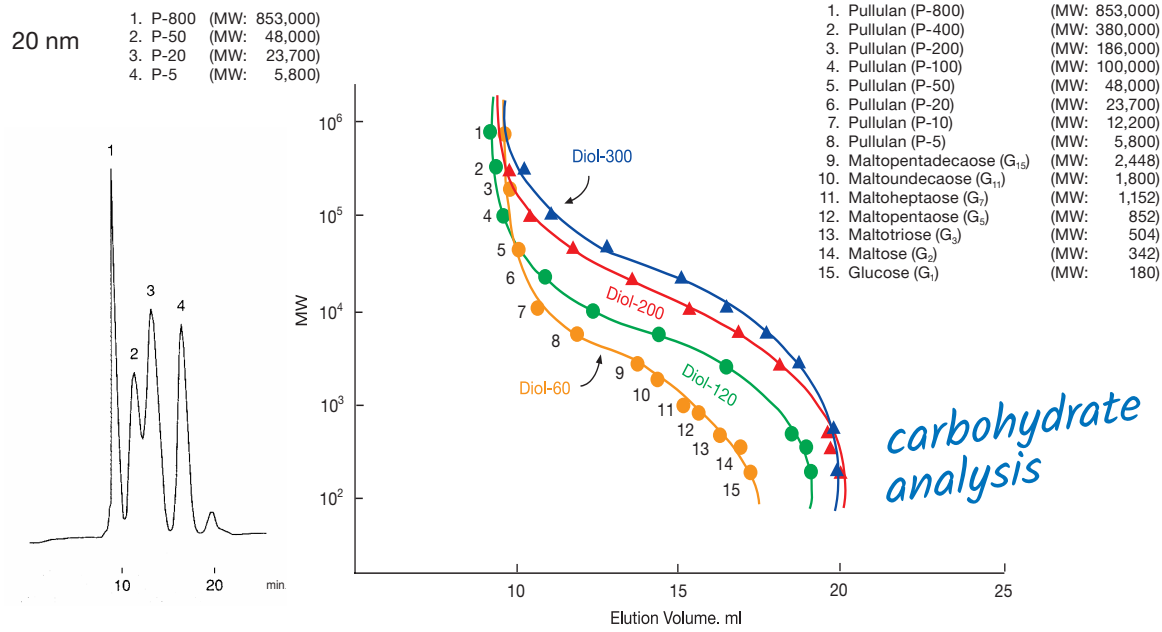
For separation of proteins with molecular weights from 10,000 to several 100,000 Da



	MW
1. IgM	900,000
2. Thyroglobulin	670,000
3. IgA	390,000
4. Fibrinogen	340,000
5. γ-Globulin	158,000
6. IgG	150,000
7. Transferrin	75,000
8. HSA (human serum albumin)	66,000
9. α1-Antitrypsin	50,000
10. Ovalbumin	45,000
11. Carbonic anhydrase	30,000
12. Trypsin inhibitor	20,100
13. Myoglobin	17,000
14. α-Lactalbumin	14,100
15. Ribonuclease A	13,700
16. Cytochrome *c*	12,400

Column:	YMC-Pack Diol, 300 x 8.0 mm ID	Flow rate:	0.5 mL/min
Part Nos.:	DL12S05-3008WT, DL20S05-3008WT, DL30S05-3008WT	Temperature:	25 °C
Eluent:	0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl	Detection:	UV at 280 nm

For molecular weight determination of oligosaccharides and polysaccharides



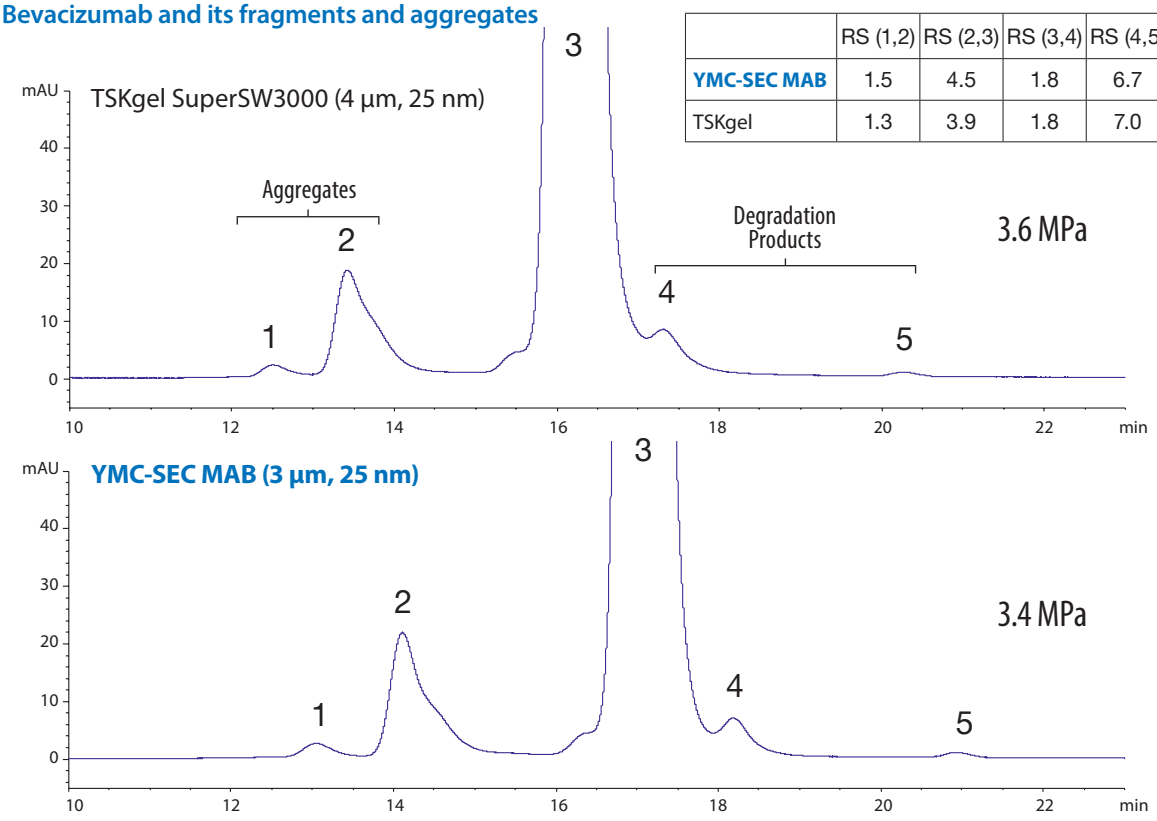
	MW
1. Pullulan (P-800)	853,000
2. Pullulan (P-400)	380,000
3. Pullulan (P-200)	186,000
4. Pullulan (P-100)	100,000
5. Pullulan (P-50)	48,000
6. Pullulan (P-20)	23,700
7. Pullulan (P-10)	12,200
8. Pullulan (P-5)	5,800
9. Maltopentadecaose (G_{15})	2,448
10. Maltoundecaose (G_{11})	1,800
11. Maltoheptaose (G_7)	1,152
12. Maltopentaose (G_5)	852
13. Maltotriose (G_3)	504
14. Maltose (G_2)	342
15. Glucose (G_1)	180

Column:	YMC-Pack Diol (20 nm) 500 x 8.0 mm ID	Flow rate:	1.0 mL/min
Part No.:	DL20S05-5008WT	Temperature:	ambient
Eluent:	water	Detection:	RI

Ideal choice for monoclonal antibodies

Bevacizumab and its fragments and aggregates

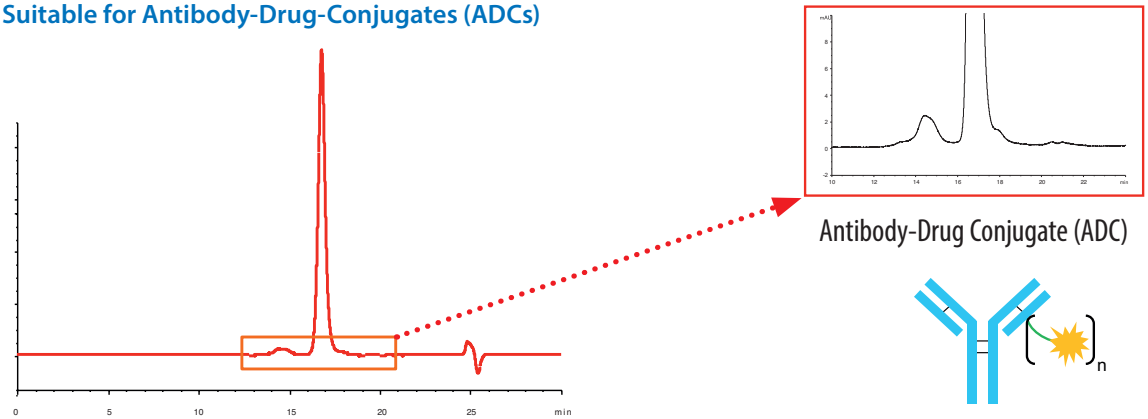
	RS (1,2)	RS (2,3)	RS (3,4)	RS (4,5)
YMC-SEC MAB	1.5	4.5	1.8	6.7
TSKgel	1.3	3.9	1.8	7.0



Column: 300 x 4.6 mm ID
Part No.: DLM25S03-3046WT
Eluent: 0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl
Flow rate: 0.165 mL/min
Temperature: 25 °C

Detection: UV at 280 nm
Cell path: 10 mm
Injection: 10 µL (5 mg/mL)
Sample: Bevacizumab (Avastin®)

Suitable for Antibody-Drug-Conjugates (ADCs)



Column: YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID
Part No.: DLM25S03-3046WT
Eluent: 0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl / 2-propanol (85 / 15)
Flow rate: 0.165 mL/min

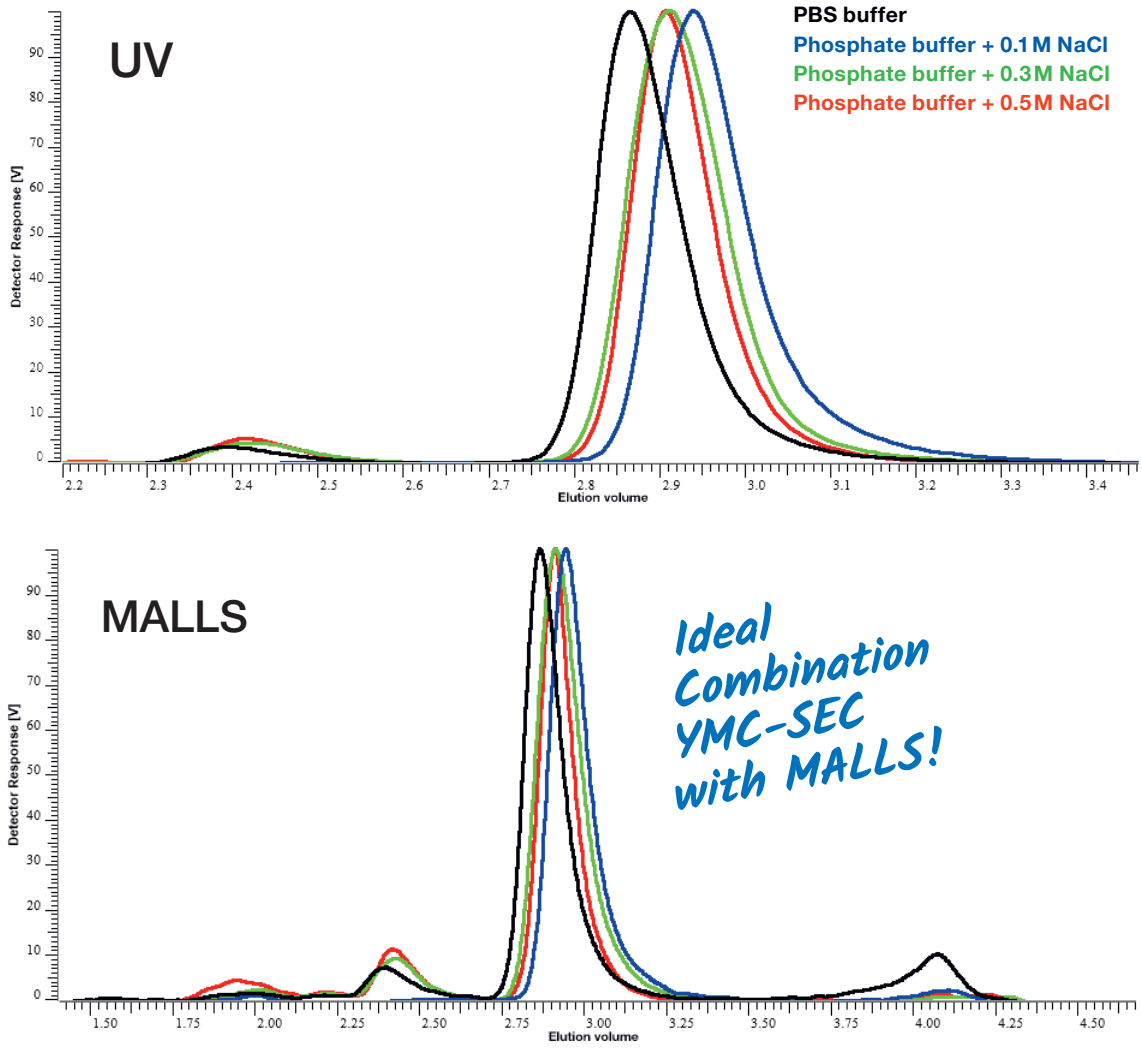
Temperature: 25 °C
Detection: UV at 280 nm
Injection: 4 µL (2.5 mg/mL)
Sample: SigmaMAb Antibody Drug Conjugate Mimic

YMC-SEC MAB is also suitable for the analysis of Antibody-Drug Conjugates (ADCs). The addition of an organic solvent to the mobile phase can improve the results obtained for ADC analysis.

SEC – YMC-SEC MAB: MALLS

YMC-SEC columns ideally combined with light scattering detection

Detection of higher molar mass species by MALLS



Column:	YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID
Part No.:	DLM25S03-3046WT
Eluent:	Phosphate buffer pH 6.6 containing 0.3 M NaCl
Flow rate:	0.33 mL/min
Temperature:	25 °C
Detection:	MALLS at 90° angle (PSS SLD7100), UV at 280 nm
Injection volume:	10 µL
Sample:	Bevacizumab (Avastin®) dosage form (10 mg/mL, diluted to 1 mg/mL)
System:	PSS-SECcurity GPC systems, 1260 Infinity II
Software:	WinGPC Unichrom

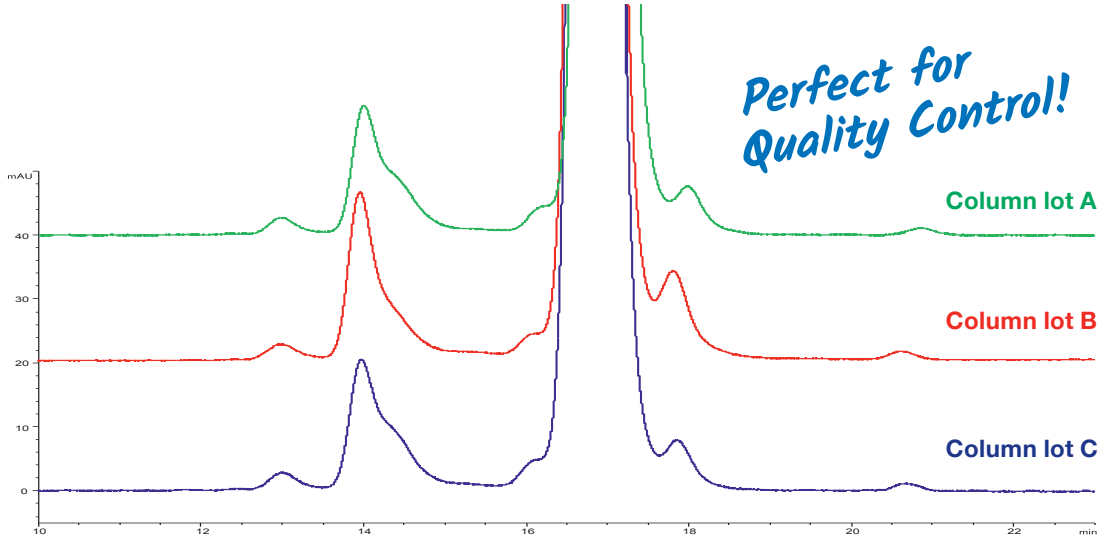
Courtesy of Thorsten Hofe, PSS Polymer Standards Service GmbH, Mainz, Germany.

Four different buffers, a phosphate buffered saline (PBS) pH 7.4 and phosphate buffers pH 6.6 with varying concentrations of NaCl, were used to develop a suitable MALLS detection method for mAbs.

A defined minimum ionic strength is necessary to achieve a robust method with good resolution. The phosphate buffer with 0.3 M NaCl appeared to be the most suitable eluent.

Compared to UV detection, the MALLS signal shows 2 higher molar mass species, aggregates of Bevacizumab, at about 2.0 mL and 2.3 mL elution volume.

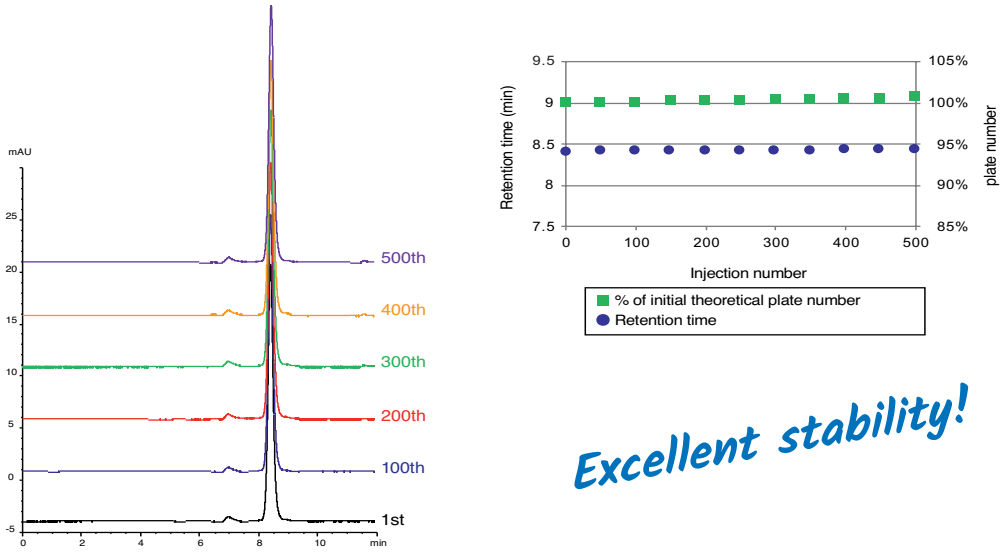
Excellent lot-to-lot reproducibility



Column:	YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID	Temperature:	25 °C
Part No.:	DLM25S03-3046WT	Detection:	UV at 280 nm
Eluent:	0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl	Injection:	10 µL (5 mg/mL)
Flow rate:	0.165 mL/min	Sample:	humanised monoclonal antibody

YMC-SEC MAB provides excellent reproducibility for the separation of monomer and aggregates as well as for monomer and fragments, making it very effective for quality control of antibody drugs.

High column stability



Column:	YMC-SEC MAB (3 µm, 25 nm) 300 x 4.6 mm ID	Temperature:	25 °C
Part No.:	DLM25S03-3046WT	Detection:	UV at 280 nm
Eluent:	0.1 M phosphate buffer (pH = 7) containing 0.2 M NaCl	Injection:	2 µL (1 mg/mL)
Flow rate:	0.33 mL/min	Sample:	humanised monoclonal antibody

Excellent stability is provided for monoclonal antibody analysis without any changes in theoretical plate number or elution time even after more than 500 injections.

SEC – YMC-Pack Diol: Resolution & throughput

Benefits of using smaller particles

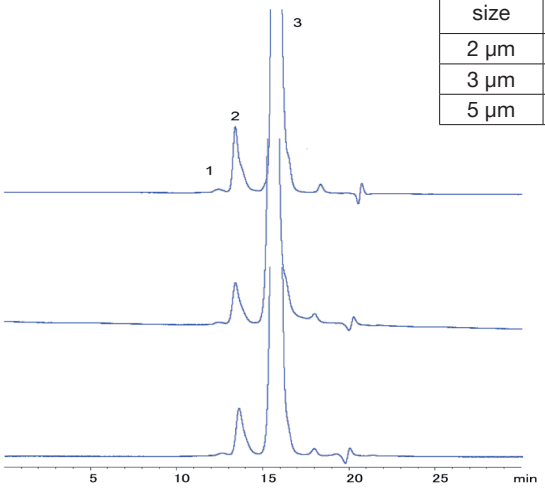
Higher resolution for analysis of monoclonal antibodies

Particle size	Rs (1,2)	Rs (2,3)	N (3)
2 µm	1.17	4.15	16,200
3 µm	1.03	3.18	10,400
5 µm	0.88	2.67	8,500

(A) YMC-Pack Diol-300 (2 µm)
300 x 4.6 mm ID

(B) YMC-Pack Diol-300 (3 µm)
300 x 4.6 mm ID

(C) YMC-Pack Diol-300 (5 µm)
300 x 4.6 mm ID



Columns: YMC-Pack Diol-300, 300 x 4.6 mm ID
Part Nos.: (A) DL30S02-3046PTH
(B) DL30S03-3046WT
(C) DL30S05-3046WT
Eluent: 0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.2 mL/min
Temperature: ambient
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody (IgG1)

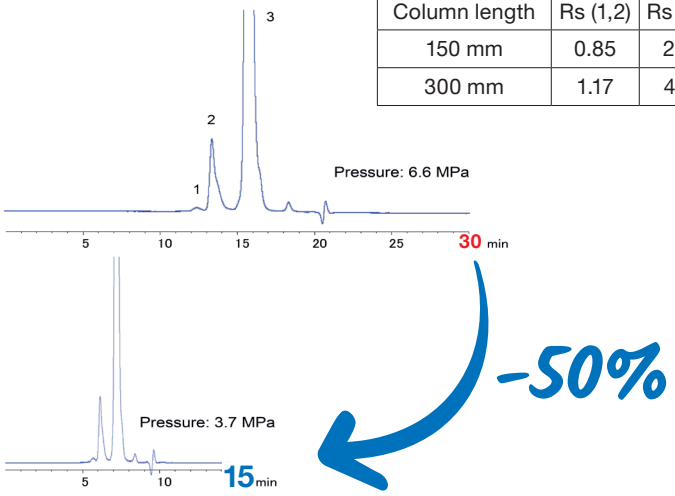
All three particle sizes show identical separation patterns for monoclonal antibody analysis. This allows easy method transfer between HPLC and UHPLC.
A method developed using conventional HPLC can be directly transferred to UHPLC using a 2 µm YMC-Pack Diol column. YMC-Pack Diol UHPLC columns greatly improve the resolution between aggregates and the monomer peak. In addition, a shoulder peak which can be observed after the monomer peak can be partially separated using the 2 µm column.

High throughput analysis of monoclonal antibodies

Column length	Rs (1,2)	Rs (2,3)	N (3)
150 mm	0.85	2.75	8,700
300 mm	1.17	4.15	16,200

YMC-Pack Diol-300 (2 µm)
300 x 4.6 mm ID

YMC-Pack Diol-300 (2 µm)
150 x 4.6 mm ID



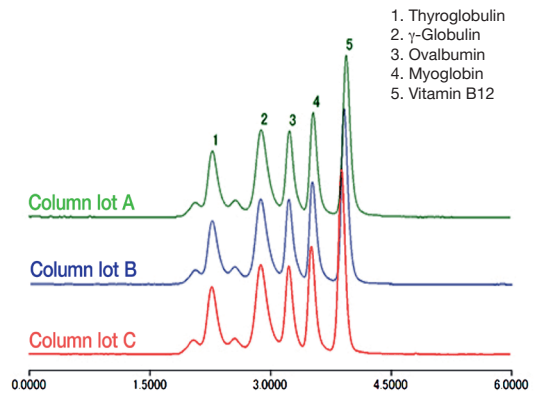
Columns: YMC-Pack Diol-300, 150 or 300 x 4.6 mm ID
Part Nos.: DL30S02-3046PTH / DL30S02-1546PTH
Eluent: 0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.2 mL/min
Temperature: ambient
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody (IgG1)

By using a 150 mm length column, 50 % shorter run times can be achieved with the good resolution as for a 300 mm length column (compare upper and lower chromatograms). This allows an increase in throughput to be achieved.

Reproducibility and stability data

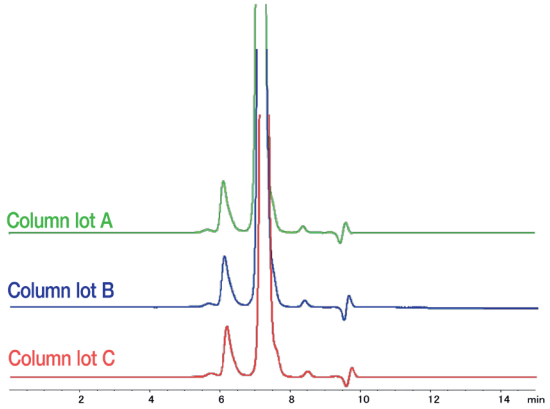
Excellent batch-to–batch reproducibility

Standard proteins



Column: YMC-Pack Diol-300 (2 µm) 150 x 4.6 mm ID
Part No.: DL30S02-1546PTH
Eluent: 0.1 M NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.5 mL/min
Temperature: ambient
Detection: UV at 280 nm
Sample: Standard proteins

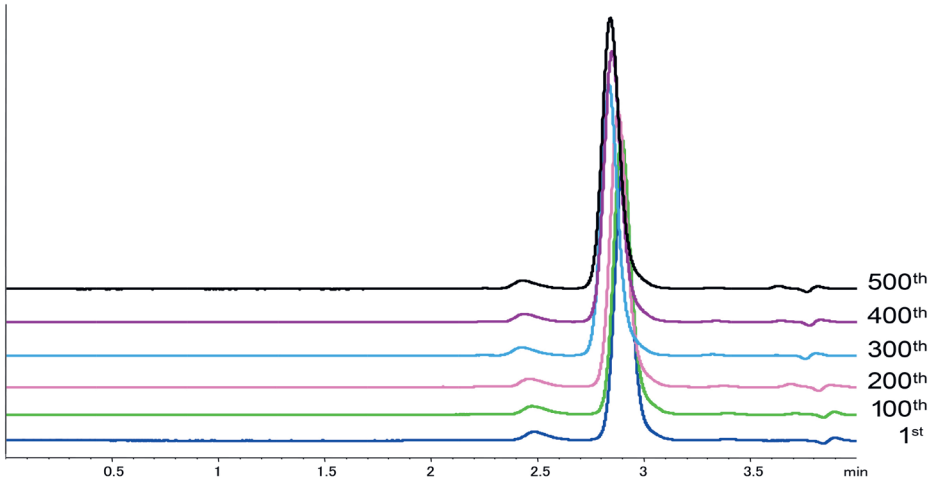
Humanised monoclonal antibody



Column: YMC-Pack Diol-300 (2 µm) 150 x 4.6 mm ID
Part No.: DL30S02-1546PTH
Eluent: 0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.2 mL/min
Temperature: 25 °C
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody

YMC-Pack Diol UHPLC columns have excellent batch-to-batch reproducibility. This makes YMC-Pack Diol 2 µm columns the ideal choice for the quality control of bio-based drugs including monoclonal antibodies.

Long-term stability



Column: YMC-Pack Diol-300 (2 µm) 150 x 4.6 mm ID
Part No.: DL30S02-1546PTH
Eluent: 0.1 M KH_2PO_4-K_2HPO_4 (pH 7.0) containing 0.2 M NaCl
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 280 nm
Sample: Humanised monoclonal antibody

YMC-Pack Diol UHPLC columns maintain their performance for more than 500 injections of sample during monoclonal antibody analysis. This ensures reproducible and reliable quality control of bio-based drugs including monoclonal antibodies.

SEC – Ordering information

2 µm UHPLC columns (max. pressure 45 MPa)

Phase	Column ID [mm]	Column length [mm]		Guard cartridges* with 10 mm length
		150	300	(pack of 5)
YMC-Pack Diol-200	4.6	DL20S02-1546PTH	DL20S02-3046PTH	DL20S02-0104GC
YMC-Pack Diol-300	4.6	DL30S02-1546PTH	DL30S02-3046PTH	DL30S02-0104GC

*Guard cartridge holder required, part no. XPGCH-Q1

3 µm HPLC columns (max. pressure 14–20 MPa)

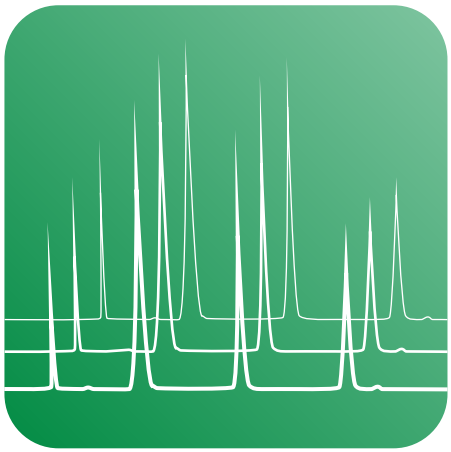
Phase	Column ID [mm]	Column length [mm]			Guard cartridges* with 10/30 mm length
		150	250	300	(pack of 5)
YMC-SEC MAB	4.6	–	–	DLM25S03-3046WT	DLM25S03-0104GC
	6.0	–	–	–	–
	8.0	–	–	DLM25S03-3008WT	–
YMC-Pack Diol-60	4.6	DL06S03-1546WT	DL06S03-2546WT	DL06S03-3046WT	DL06S03-0104GC
	6.0	–	–	DL06S03-3006WT	–
	8.0	DL06S03-1508WT	–	DL06S03-3008WT	DL06S03-0308WTG**
YMC-Pack Diol-120	4.6	DL12S03-1546WT	DL12S03-2546WT	DL12S03-3046WT	DL12S03-0104GC
	6.0	–	–	DL12S03-3006WT	–
	8.0	DL12S03-1508WT	–	DL12S03-3008WT	DL12S03-0308WTG**
YMC-Pack Diol-200	4.6	DL20S03-1546WT	DL20S03-2546WT	DL20S03-3046WT	DL20S03-0104GC
	6.0	–	–	DL20S03-3006WT	–
	8.0	DL20S03-1508WT	–	DL20S03-3008WT	DL20S03-0308WTG**
YMC-Pack Diol-300	4.6	DL30S03-1546WT	DL30S03-2546WT	DL30S03-3046WT	DL30S03-0104GC
	6.0	–	–	DL30S03-3006WT	–
	8.0	DL30S03-1508WT	–	DL30S03-3008WT	DL30S03-0308WTG**

*Guard cartridge holder required, part no. XPGCH-Q1
**no holder required for 30 x 8 mm ID guards
recommended column coupler part no. XRCP1602

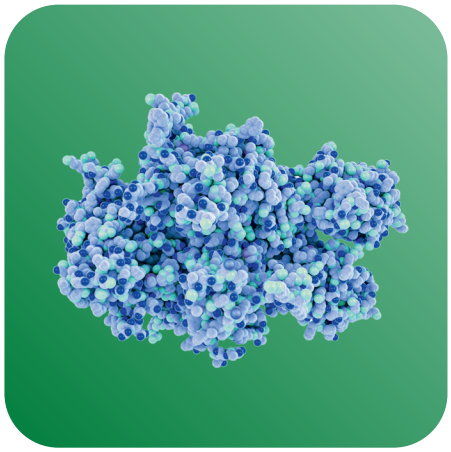
5 µm HPLC columns (max. pressure 20 MPa)

Phase	Column ID [mm]	Column length [mm]			Guard cartridges* with 10/30 mm length
		250	300	500	(pack of 5)
YMC-Pack Diol-60	4.6	DL06S05-2546WT	DL06S05-3046WT	–	DL06S05-0104GC
	6.0	DL06S05-2506WT	DL06S05-3006WT	DL06S05-5006WT	–
	8.0	–	DL06S05-3008WT	DL06S05-5008WT	DL06S05-0308WTG**
	10.0	DL06S05-2510WT	DL06S05-3010WT	DL06S05-5010WT	DL06S05-0310WTG**
YMC-Pack Diol-120	4.6	DL12S05-2546WT	DL12S05-3046WT	–	DL12S05-0104GC
	6.0	DL12S05-2506WT	DL12S05-3006WT	DL12S05-5006WT	–
	8.0	–	DL12S05-3008WT	DL12S05-5008WT	DL12S05-0308WTG**
	10.0	DL12S05-2510WT	DL12S05-3010WT	DL12S05-5010WT	DL12S05-0310WTG**
YMC-Pack Diol-200	4.6	DL20S05-2546WT	DL20S05-3046WT	–	DL20S05-0104GC
	6.0	DL20S05-2506WT	DL20S05-3006WT	DL20S05-5006WT	–
	8.0	–	DL20S05-3008WT	DL20S05-5008WT	DL20S05-0308WTG**
	10.0	DL20S05-2510WT	DL20S05-3010WT	DL20S05-5010WT	DL20S05-0310WTG**
YMC-Pack Diol-300	4.6	DL30S05-2546WT	DL30S05-3046WT	–	DL30S05-0104GC
	6.0	DL30S05-2506WT	DL30S05-3006WT	DL30S05-5006WT	–
	8.0	–	DL30S05-3008WT	DL30S05-5008WT	DL30S05-0308WTG**
	10.0	DL30S05-2510WT	DL30S05-3010WT	DL30S05-5010WT	DL30S05-0310WTG**

*Guard cartridge holder required, part no. XPGCH-Q1
**no holder required for 30 x 8/10 mm ID guards
recommended column coupler part no. XRCP1602 (for 8 mm ID) and XRCP1605 (for 10 mm ID)

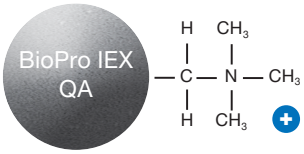


IEX

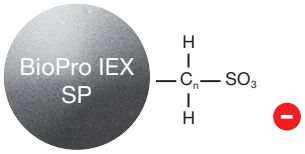


IEX – Bio Pro Series

- **porous or non-porous hydrophilic polymers**
- **high binding capacity and recovery of biomolecules**
- **very high resolution**
- **low nonspecific adsorption**
- **excellent reproducibility**



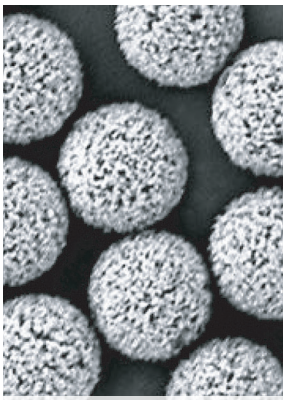
strong anion exchanger



strong cation exchanger

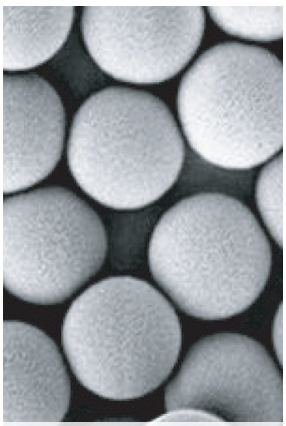
	BioPro IEX QA	BioPro IEX SP
Matrix	hydrophilic polymer (polymethacrylate)	hydrophilic polymer (polymethacrylate)
Particle size / µm	5	5
Pore size / nm	100	100
Charged group	$-CH_2N^+(CH_3)_3$	$-(CH_2)_3SO_3^-$
Counter ion	Cl^-	Na^+
Available pH range	2.0–12.0	2.0–12.0
Temperature range	4–60 °C	
Pressure limit	2.5–3.5 MPa (360–510 psi)	
Column hardware	PEEK	

Also available in 10, 20, 30 or 75 µm for preparative scale



Porous polymer beads

	BioPro IEX QF	BioPro IEX SF
Matrix	hydrophilic polymer (polymethacrylate)	hydrophilic polymer (polymethacrylate)
Particle size / µm	3, 5	3, 5
Pore size / nm	non-porous	non-porous
Charged group	$-CH_2N^+(CH_3)_3$	$-(CH_2)_3SO_3^-$
Counter ion	Cl^-	Na^+
Available pH range	2.0–12.0	2.0–12.0
Temperature range	4–60 °C	
Pressure limit	3 µm: 18–25 MPa (2,610–3,625 psi) 5 µm: 6–12 MPa (870–1,740 psi)	
Column hardware	PEEK	



Non-porous polymer beads

YMC´s BioPro IEX series of ion exchange columns are available in QA and SP chemistries, based on 5 µm porous (QA or SP columns) or on 3 or 5 µm non-porous (QF and SF columns) hydrophilic polymer beads.

The porous materials offer excellent binding capacity with exceptionally high efficiency and low operating pressure, whilst the non-porous particles offer high efficiency, very high resolution and low operating pressures.

High binding capacity and high recovery for porous type

The porous versions of YMC´s BioPro IEX show high dynamic binding capacity and excellent recovery, making them useful for semi-preparative separations of proteins and antibodies.

Comparison of dynamic binding capacity (DBC) for BSA

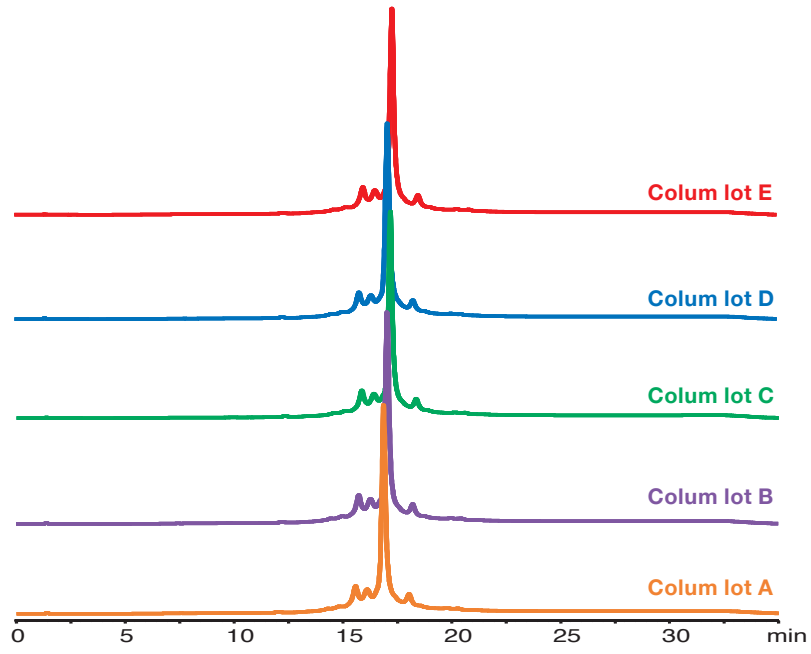
	Dynamic binding capacity (mg/mL-gel, 10% breakthrough)	Eluted amount (mg/mL-gel)	Recovery* (%)
BioPro IEX QA	126	120	95
Mono Q	100	35	35
TSKgel BioAssist Q	73	58	79

High recovery rates for BioPro IEX

* Recovery: (Eluted amount/Dynamic binding capacity) x 100

Compared with conventional porous polymer anion exchange columns, BioPro IEX QA provides higher DBC and recovery rates. This indicates that BioPro IEX has a much lower nonspecific adsorption compared to conventional columns.

Excellent batch-to-batch reproducibility



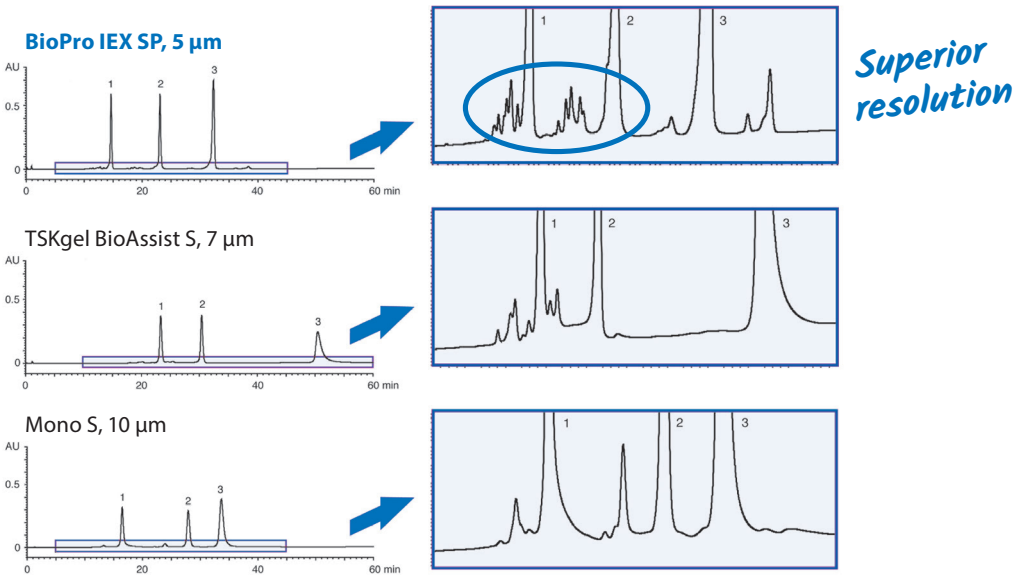
Column:	BioPro IEX SF (5 µm) 100 x 4.6 mm ID	Flow rate:	0.5 mL/min (180 cm/h)
Part No.:	SF00S05-1046WP	Temperature:	25 °C
Eluent:	A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 6.5)	Detection:	UV at 215 nm
	B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 6.5) containing 0.2 M NaCl	Injection:	10 µL
Gradient:	0–50%B (0.5–30 min)	Sample:	monoclonal antibody (IgG1)

BioPro IEX SF columns exhibit excellent batch-to-batch reproducibility for mAb analysis with resolution of peaks for small charge variants. All gel batches are inspected by rigorous quality control tests, including HPLC analysis of mAbs, and must meet the required criteria before release.
BioPro IEX columns are the best choice for the quality control of mAbs and other biopharmaceuticals.

IEX – BioPro IEX: Resolution & throughput

Superior resolution

Comparison of standard protein separation on BioPro IEX SP and commercial S type products

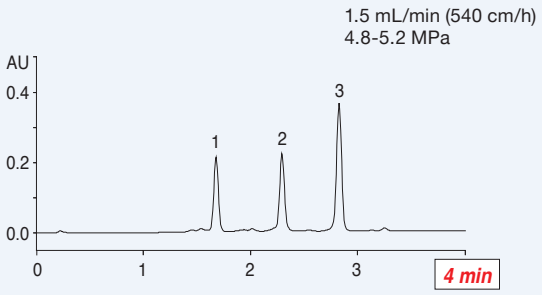


Eluent:	A) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8)		Detection:	UV at 220 nm	
	B) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8) containing 0.5 M NaCl		Injection:	BioPro IEX SP, TSKgel BioAssist S	20 µL
Gradient:	0–100%B (0–60 min)			Mono S	23.6 µL
Flow rate:	BioPro IEX SP, TSKgel BioAssist S	0.5 mL/min	Sample:	1. Ribonuclease A	(0.5 mg/mL)
	Mono S	0.59 mL/min		2. Cytochrome *c*	(0.5 mg/mL)
Temperature:	25 °C			3. Lysozyme	(0.5 mg/mL)

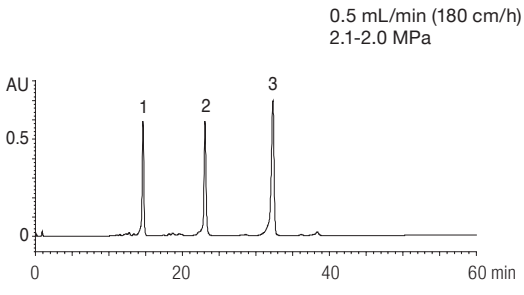
Only BioPro IEX is available in the smaller particle size and is therefore able to provide superior resolution.

Ultra-high-throughput analysis with non-porous BioPro IEX

Non-porous type
BioPro IEX SF (5 µm) 30 x 4.6 mm ID



Porous type
BioPro IEX SP (5 µm) 50 x 4.6 mm ID

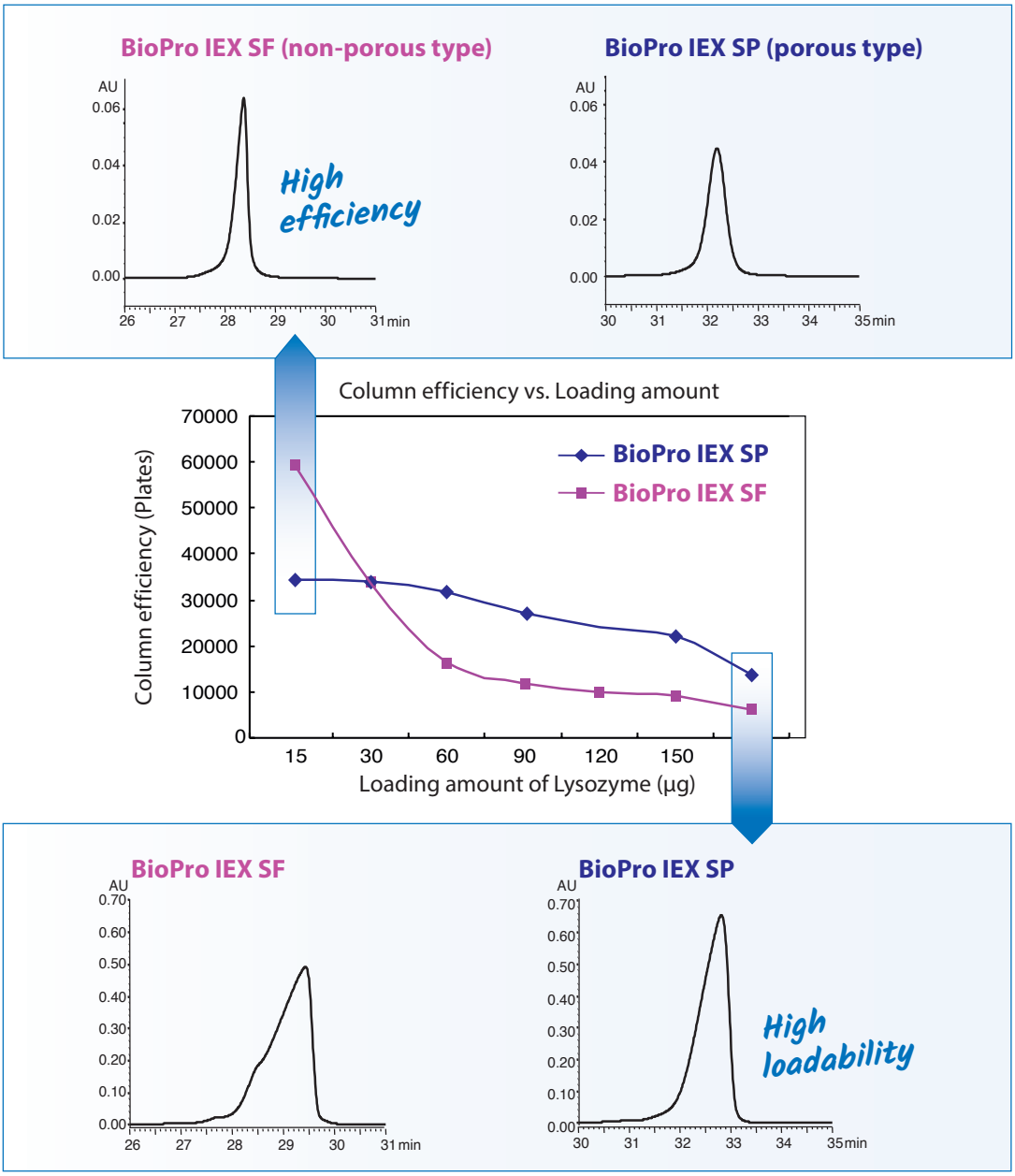


Eluent:	A) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8)		Temperature:	25°C	
	B) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8) containing 0.5 M NaCl		Detection:	UV at 220 nm	
Part Nos.:	SF00S05-0346WP (non-porous)		Injection:	20 µL	
	SPA0S05-0546WP (porous)		Sample:	1. Ribonuclease A	(0.5 mg/mL)
Gradient:	0–100%B (0–4 min) for BioPro IEX SF			2. Cytochrome *c*	(0.5 mg/mL)
	0–100%B (0–60 min) for BioProIEX SP			3. Lysozyme	(0.5 mg/mL)

The high mechanical stability of non-porous polymer beads and the short column length allow faster elution of proteins at a higher flow rate without any loss of resolution.

Column efficiency and loadability

When to use porous and non-porous BioPro IEX



Columns: (5 µm) 50 x 4.6 mm ID
Part Nos.: SF00S05-0546WP
SPA0S05-0546WP
Eluent: A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 6.8)
B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 6.8) containing 0.5 M NaCl
Gradient: 0–100%B (0–60 min)

Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 280 nm
Injection: 100 µL
Sample: Lysozyme

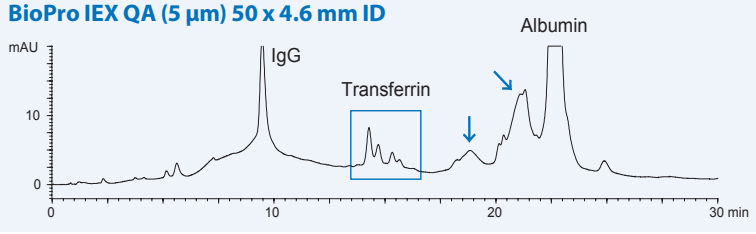
BioPro IEX SF offers outstanding column efficiency at small amount of sample loading. Non-porous type of BioPro IEX columns are especially suitable for microscale analysis which requires higher resolution. BioPro IEX SP maintains the good peak shape even when the loading amount increases. Porous type BioPro IEX columns with high capacity are useful for high-load analytical separations and laboratory-scale purification.

IEX – BioPro IEX: Challenging separations

Protein separation in challenging matrices

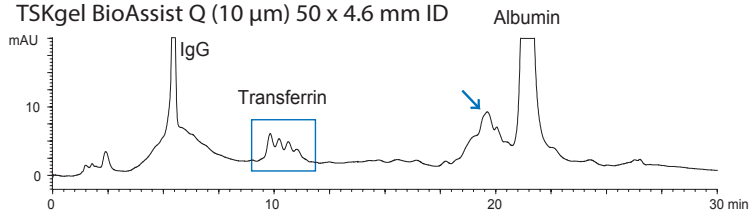
Separation of proteins in human serum on BioPro IEX QA and commercial Q-type products

BioPro IEX QA (5 µm) 50 x 4.6 mm ID

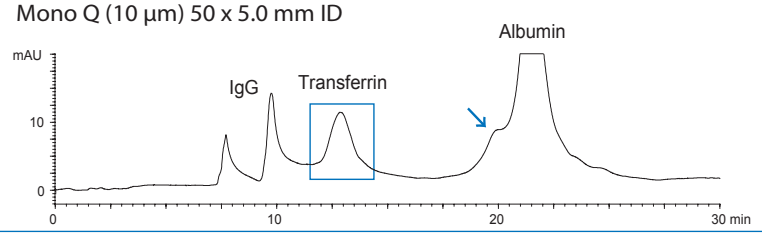


For high resolution porous BioPro IEX QA/SP is recommended!

TSKgel BioAssist Q (10 µm) 50 x 4.6 mm ID



Mono Q (10 µm) 50 x 5.0 mm ID



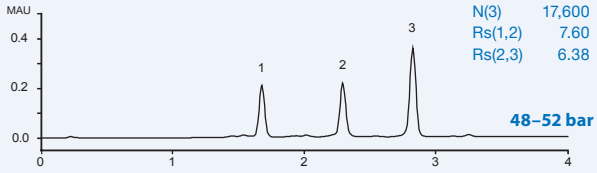
Part No.:	QAA0S05-0546WP	Temperature:	25 °C
Eluent:	A) 20 mM Tris-HCl (pH 8.6)	Detection:	UV at 280 nm
	B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl	Injection:	20 µL
Gradient:	0–30%B (0–15 min), 30–100%B (15–30 min)	Sample:	Human serum (100 µL/mL)
Flow rate:	0.5 mL/min		

Better performance at lower backpressure

Comparison of standard protein separation on BioPro IEX SF and a commercial SP-type product

BioPro IEX SF (5 µm) 30 x 4.6 mm ID

N(1)	7,100
N(2)	12,100
N(3)	17,600
Rs(1,2)	7.60
Rs(2,3)	6.38

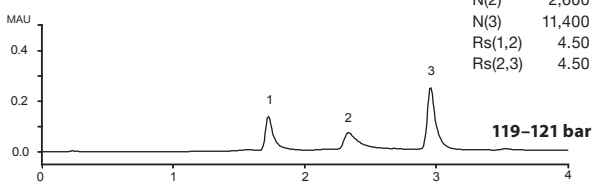


BioPro IEX SF elutes the proteins in sharper peaks without peak-tailing compared to TSKgel SP-NPR.
Despite the larger particle size, the theoretical plate count for BioPro IEX SF is higher than that for TSKgel SP-NPR.

higher plate count

TSKgel SP-NPR (2.5 µm) 35 x 4.6 mm ID

N(1)	4,400
N(2)	2,600
N(3)	11,400
Rs(1,2)	4.50
Rs(2,3)	4.50



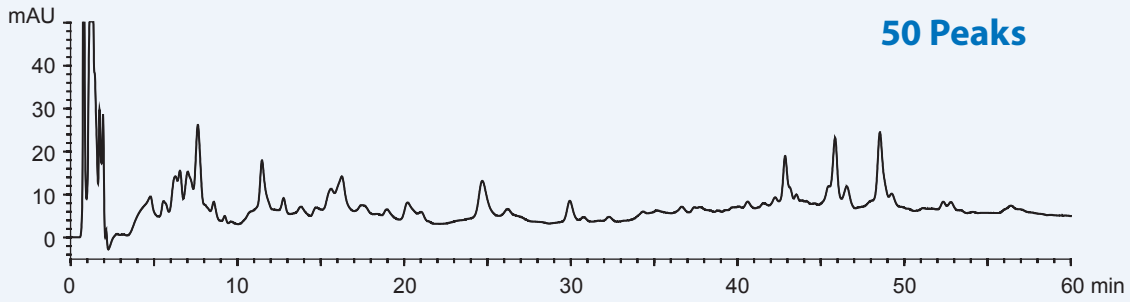
Part No.:	SF00S05-0346WP	
Eluent:	A) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8)	
	B) 20 mM KH_2PO_4-K_2HPO_4 (pH 6.8) containing 0.5 M NaCl	
Gradient:	BioPro IEX SF 0-100%B (0–4 min)	
	TSKgel SP-NPR 0-100%B (0–4.67 min)	
Flow rate:	1.5 mL/min	
Temperature:	25 °C	
Detection:	UV at 220 nm	
Injection:	20 µL	
Sample:	1. Ribonuclease A	(0.1 mg/mL)
	2. Cytochrome *c*	(0.1 mg/mL)
	3. Lysozyme	(0.1 mg/mL)

Compared to the competitor's column, BioPro IEX SF gives higher theoretical plate counts, excellent peak shapes, and lower backpressures. This makes BioPro IEX SF most suitable for high-throughput analysis.

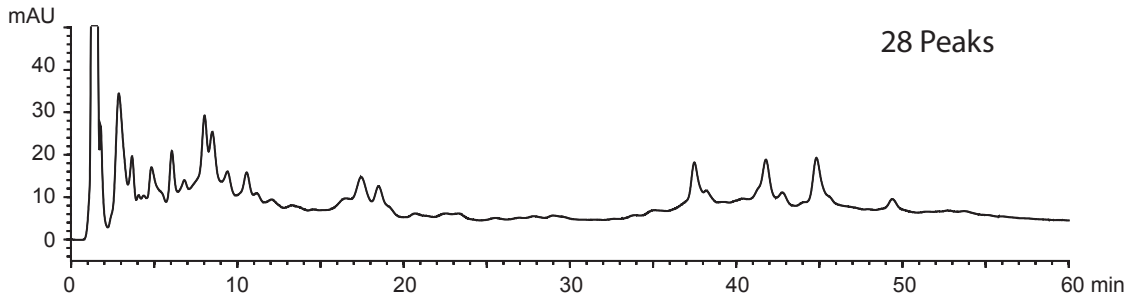
Peptide mapping

Peptide mapping of tryptic digests of BSA with enhanced sensitivity

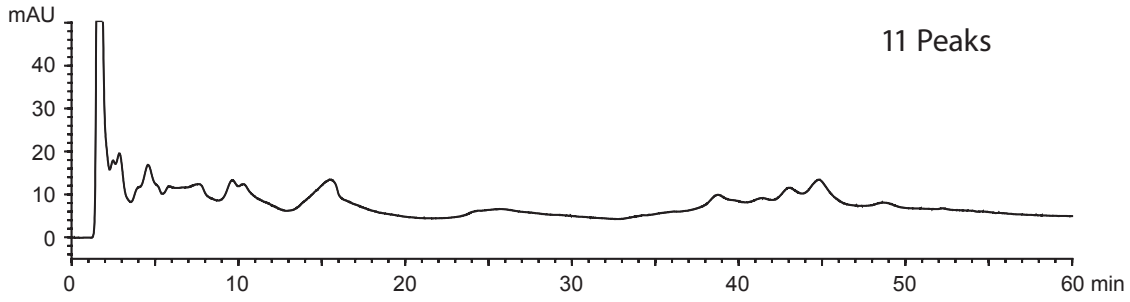
BioPro IEX QA (5 µm) 50 x 4.6 mm ID



TSKgel BioAssist Q (10 µm) 50 x 4.6 mm ID



Mono Q (10 µm) 50 x 5.0 mm ID

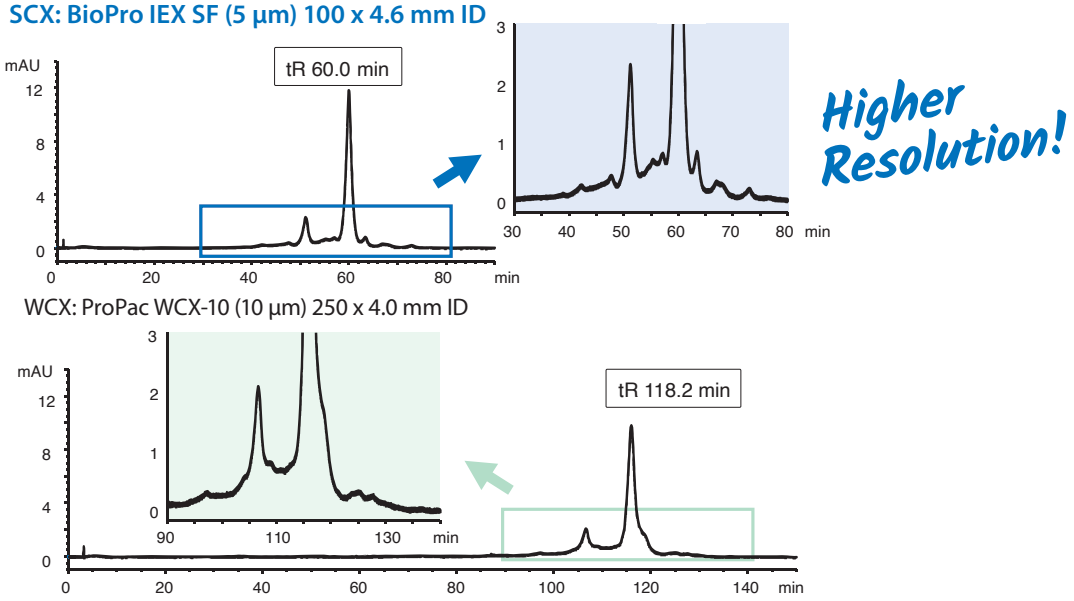


Part No.:	QAA0S05-0546WP
Eluent:	A) 20 mM Tris-HCl (pH 8.6)
	B) 20 mM Tris-HCl (pH 8.6) containing 0.5 M NaCl
Gradient:	0–15%B (0–30 min), 15–60%B (30–60 min)
Flow rate:	0.5 mL/min
Temperature:	25 °C
Detection:	UV at 220 nm
Injection:	20 µL
Sample:	Tryptic digest of BSA

IEX – BioPro IEX: Antibody analysis

Monoclonal antibody analysis with non-porous cation exchange columns

SCX: BioPro IEX SF (5 µm) 100 x 4.6 mm ID



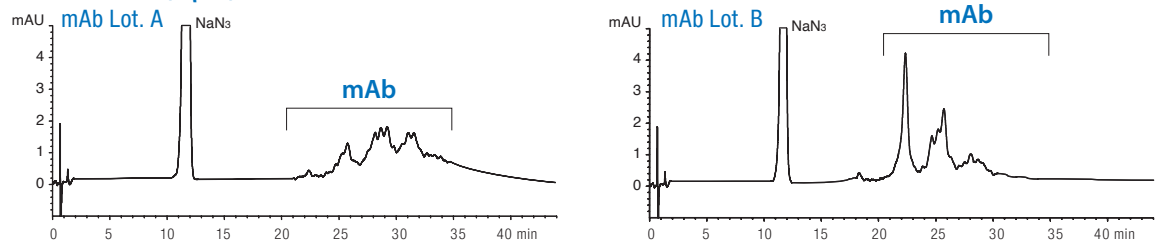
WCX: ProPac WCX-10 (10 µm) 250 x 4.0 mm ID

Part No.: SF00S05-1046WP
Eluent: A) 20 mM MES-NaOH (pH 5.6)
B) 20 mM MES-NaOH (pH 5.6) containing 0.2 M NaCl
Initial ratio: 35%B (70 mM NaCl)
Gradient slope: 0.25%B/min (0.5 mM NaCl)
Flow rate: 180 cm/h (0.5 mL/min for 100 x 4.6 mm ID, 0.378 mL/min for 250 x 4.0 mm ID)
Temperature: 30 °C
Detection: UV at 280 nm
Injection: 10 µL
Sample: Humanized monoclonal IgG1 (1 mg/mL)

The separation of a mAb is compared using a strong cation (BioPro IEX SF) and a weak cation exchange column (ProPac WCX-10) under the same gradient conditions at pH 5.6. BioPro IEX SF can achieve a higher resolution of the mAb than the competitor's column in a shorter analysis time.

QC of monoclonal antibodies with non-porous BioPro IEX QF

BioPro IEX QF (5 µm) 100 x 4.6 mm ID



Part No.: QF00S05-1046WP
Eluent: A) 20 mM Tris-HCl (pH 8.1)
B) 20 mM Tris-HCl (pH 8.1) containing 0.5 M NaCl
Gradient: 10–25%B (0–60 min)
Flow rate: 1.0 mL/min (360 cm/h)
Temperature: 25˚C
Detection: UV at 220 nm
Injection: 14 µL (0.1 mg/mL)
Sample: Mouse monoclonal IgG1 anti-human IgG4 (Purified by DEAE chromatography, containing NaN_3)

Two different batches of a commercially available mAb purified by DEAE chromatography were analysed on a BioPro IEX QF column (100 mm length).
The mAb was separated into several peaks, and the batch-to-batch variability is observed. The BioPro IEX QF/SF 100 mm length columns, which have high efficiency, are ideal for characterisation of glycoproteins, such as monoclonal antibodies, and for quality control assessment of biopharmaceuticals.

Optimisation of oligonucleotide separations on ion exchange chromatography

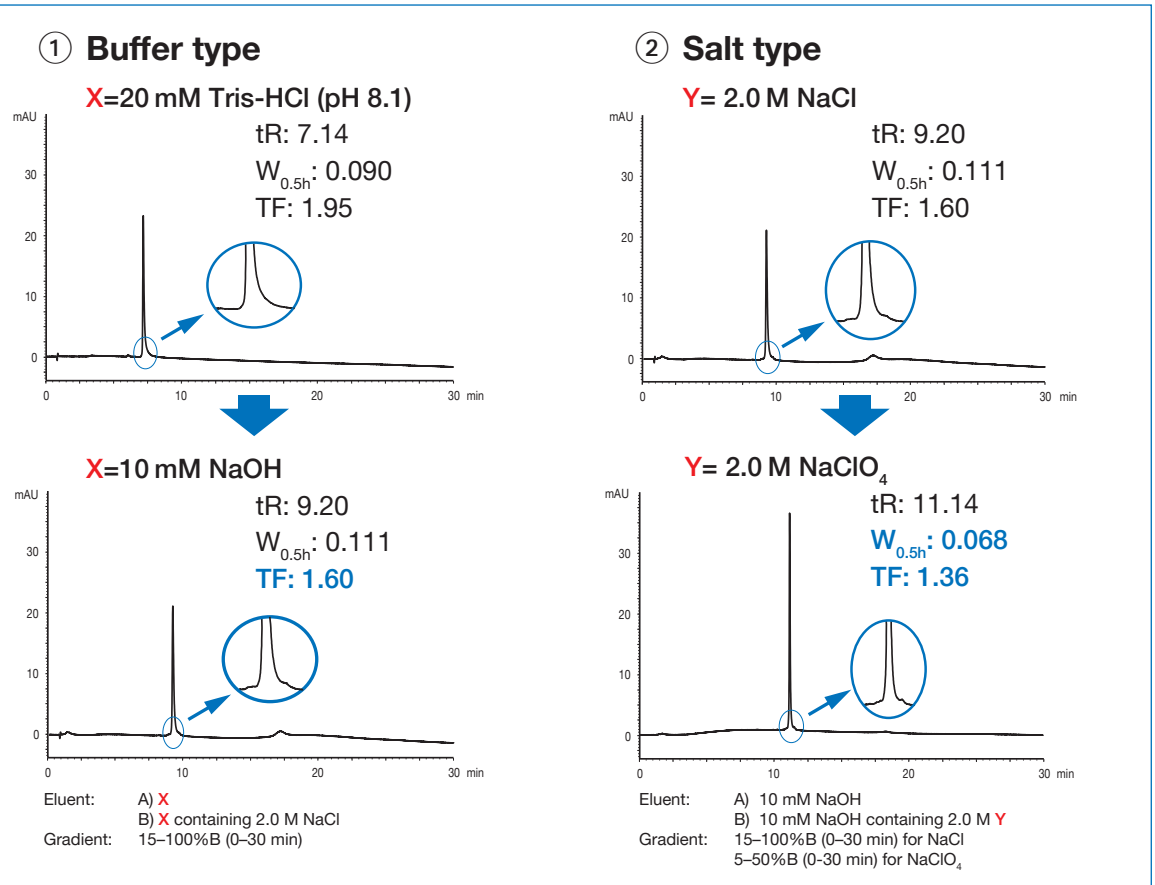
Non-porous anion exchange column is generally suitable for analysis of oligonucleotides. For optimisation of single-stranded DNA and RNA of about 20mer some conditions such as type of mobile phase and column temperature can be changed.

1 Improvement of peak tailing

Sample Group 1 (Phosphodiester oligonucleotides; PO)

Single-stranded RNA (ssRNA)	5′-UCAUCACACUGAAUACCAAU-3′ (RNA 20mer)

By changing the buffer from 20 mM Tris-HCl (pH 8.1) to 10 mM NaOH, the tailing factor for an oligonucleotide is reduced. Furthermore, the peak tailing is further suppressed when $NaClO_4$ was added to 10 mM NaOH instead of NaCl.



Eluent: A) X
B) X containing 2.0 M NaCl
Gradient: 15–100%B (0–30 min)

Eluent: A) 10 mM NaOH
B) 10 mM NaOH containing 2.0 M Y
Gradient: 15–100%B (0–30 min) for NaCl
5–50%B (0-30 min) for $NaClO_4$

Column:	BioPro IEX QF (5 µm) 100 x 4.6 mm ID	Gradient:	15–100%B (0–30 min) for NaCl 5–50%B (0–30 min) for $NaClO_4$
Part No.:	QF00S05-1046WP		
Flow rate:	1.0 mL/min	Temperatur:	25 °C
Eluent:	A) 20 mM Tris-HCl (pH 8.1) or 10 mM NaOH B) 20 mM Tris HCl (pH 8.1) or 10 mM NaOH containing 2.0 M NaCl or $NaClO_4$	Detection:	UV at 260 nm
		Injection:	2 µL (10 nmol/mL)
		Sample:	RNA 20mer

2 Improvement of carry-over

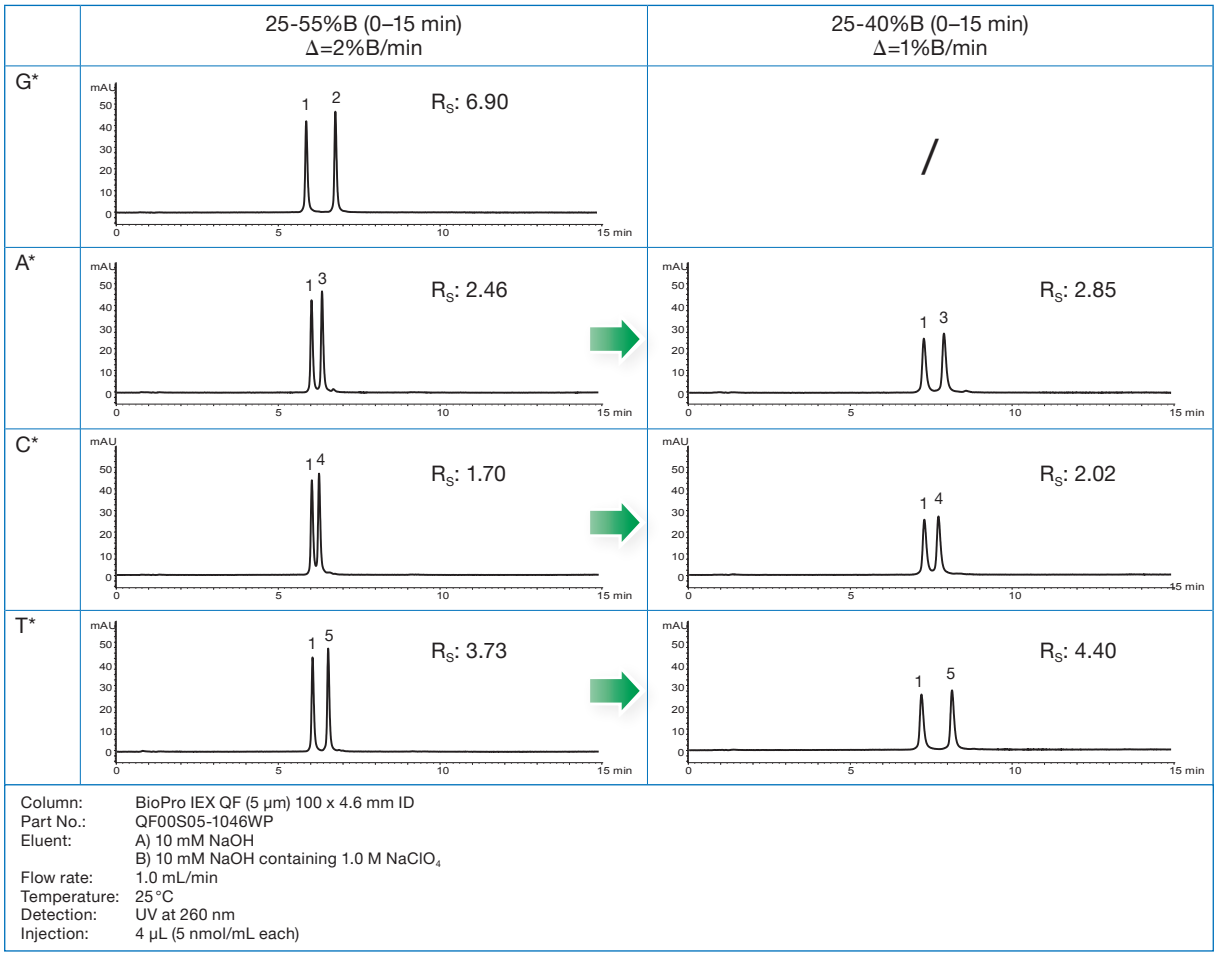
When the initial gradient concentration of NaCl is low (ex. 50 mM), carry-over is observed. By increasing the initial gradient concentration of NaCl up to 400 carry-over can be avoided with good reproducibility.

IEX – Expert Tips: Oligonucleotides

3 Improvement of ssDNA separation with single-base differences (differing in the type of base of 5'end of DNA 21mer)

When ssDNAs with single-base differences (differing in the type of base of 5'end of DNA 21mer) is analysed the degree of separation varies depending on the type of base at the 5' end. If the separation is difficult, it can be improved by making the gradient steeper.

1	Single-stranded DNA	5'-TCATCACACTGAATACCAAT-3' (DNA 20mer)
2		5'-GTCATCACACTGAATACCAAT-3' (DNA 21mer)
3		5'-ATCATCACACTGAATACCAAT-3' (DNA 21mer)
4		5'-CTCATCACACTGAATACCAAT-3' (DNA 21mer)
5		5'-TTCATCACACTGAATACCAAT-3' (DNA 21mer)



Column: BioPro IEX QF (5 µm) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 10 mM NaOH
B) 10 mM NaOH containing 1.0 M $NaClO_4$
Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: UV at 260 nm
Injection: 4 µL (5 nmol/mL each)

*base of 5'end of DNA 21mer

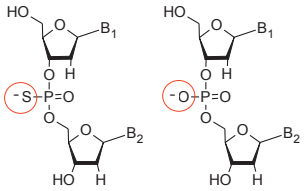
4 Improvement of the separation of phopsphorothioate oligonucleotides with single-base differences in length

Since acidity of all PS is much higher than all PO, a higher salt concentration is required for elution. The peak of all PS is much broader because it is thought that all PS contains as many as 2^{19} (524,288) stereoisomers. A steeper gradient curve, increasing column temperature and adding organic solvent can improve peak separation.

Sample Group 2 (Phosphorothioate oligonucleotides; PS)

1	Single-stranded RNA	5′-U~C~A~U~C~A~C~A~C~U~G~A~A~U~A~C~C~A~A~U-3′ (RNA 20mer All PS)
2		5′-G~U~C~A~U~C~A~C~A~C~U~G~A~A~U~A~C~C~A~A~U-3′ (RNA 21mer All PS)

~=Phosphorothioated



Phosphorothioate Linkage (PS) | Phosphodiester Linkage (PO)

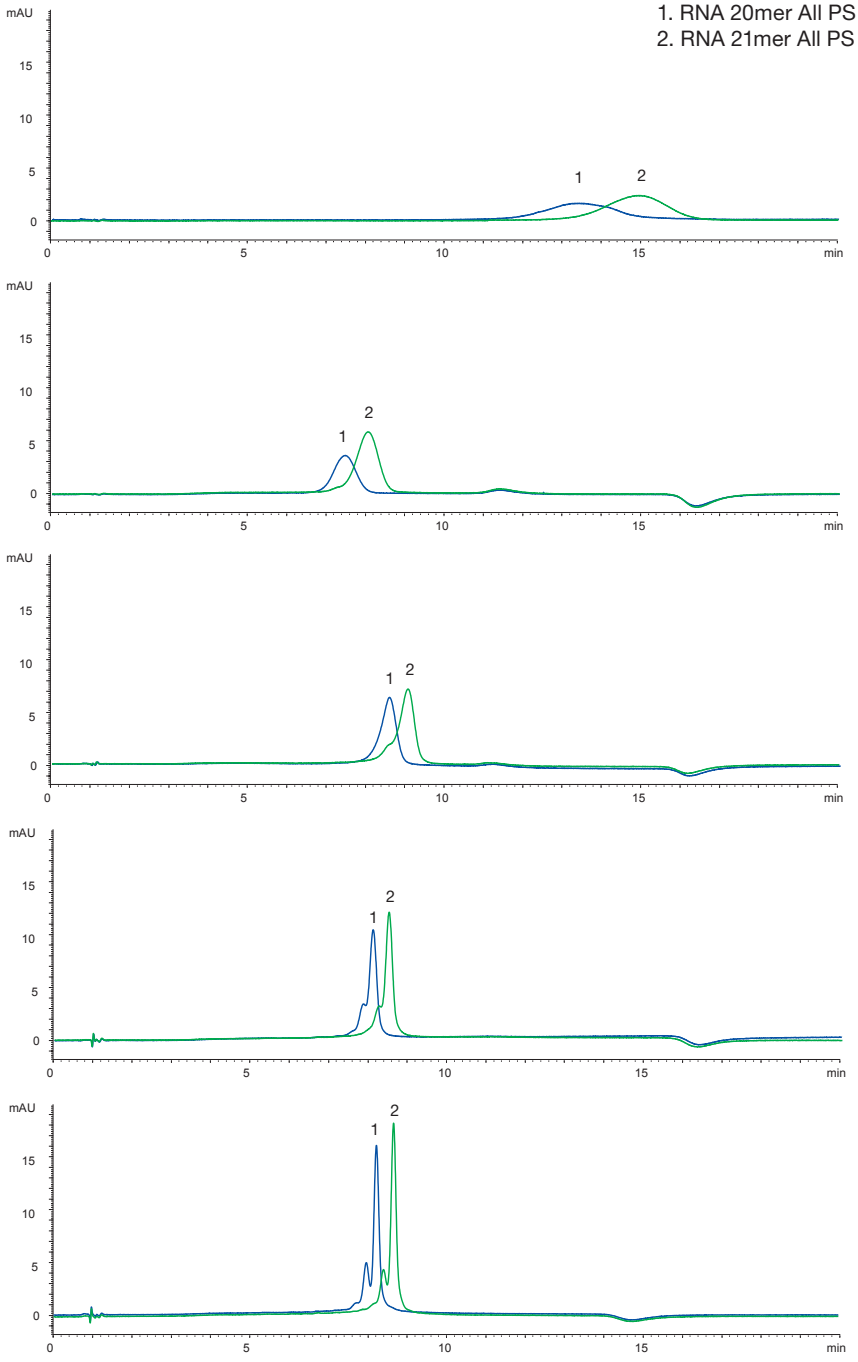
X/Y=100/0
32–80%B (0–24 min)
= Δ20 mM $NaClO_4$/min
25°C

Steep gradient curve
X/Y=100/0
32–80%B (0–8 min)
= Δ60 mM $NaClO_4$/min
25°C

Raising column temperature
X/Y=100/0
32–80%B (0–8 min)
=Δ60 mM $NaClO_4$/min
60 °C

Addition of organic solvent
X/Y=80/20
40–100%B (0–8 min)
= Δ60 mM $NaClO_4$/min
60 °C

Increasing ratio of organic solvent
X/Y=70/30
40–100%B (0–6.3 min)
= Δ60 mM $NaClO_4$/min
60 °C



Column: BioPro IEX QF (5 µm) 100 x 4.6 mm ID
Part No.: QF00S05-1046WP
Eluent: A) 10 mM NaOH/methanol (X/Y)
B) 10 mM NaOH containing 1.0 M $NaClO_4$/methanol (X/Y)

Flow rate: 1.0 mL/min
Detection: UV at 260 nm
Injection: 2 µL (10 nmol/mL)

IEX – Ordering information

3 µm non-porous analytical columns, PEEK hardware (max. pressure 18–25 MPa)

Phase	Column ID [mm]	Column length [mm]				Precolumn filter 2 µm*
		30 (25 MPa)	50 (25 MPa)	100 (25 MPa)	150 (18 MPa)	(pack of 5)
BioPro IEX QF	4.6	QF00S03-0346WP	QF00S03-0546WP	QF00S03-1046WP	QF00S03-1546WP	XRPRCP25
BioPro IEX SF	4.6	SF00S03-0346WP	SF00S03-0546WP	SF00S03-1046WP	SF00S03-1546WP	

5 µm non-porous analytical columns, PEEK hardware (max. pressure 6–12 MPa)

Phase	Column ID [mm]	Column length [mm]				Precolumn filter 2 µm*
		30 (6 MPa)	50 (10 MPa)	100 (12 MPa)	150 (12 MPa)	(pack of 5)
BioPro IEX QF	4.6	QF00S05-0346WP	QF00S05-0546WP	QF00S05-1046WP	QF00S05-1546WP	XRPRCP25
BioPro IEX SF	4.6	SF00S05-0346WP	SF00S05-0546WP	SF00S05-1046WP	SF00S05-1546WP	

5 µm porous analytical columns, PEEK hardware (max. pressure 2.5–3.5 MPa)

Phase	Column ID [mm]	Column length [mm]			Precolumn filter 2 µm*
		30 (2.5 MPa)	50 (3.0 MPa)	100 (3.5 MPa)	(pack of 5)
BioPro IEX QA	4.6	QAA0S05-0346WP	QAA0S05-0546WP	QAA0S05-1046WP	XRPRCP25
BioPro IEX SP	4.6	SPA0S05-0346WP	SPA0S05-0546WP	SPA0S05-1046WP	

* Holder required, part no. XRPRCP02

6 µm non-porous semiprep. columns, stainless steel hardware (max. pressure 3-9 MPa)**

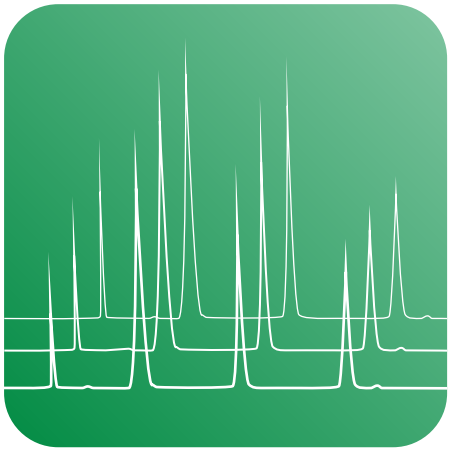
Phase	Column ID [mm]	Column length [mm]
		100
BioPro IEX QF	10 20 30	QF00S06-1010WT QF00S06-1020WT QF00S06-1030WT
BioPro IEX SF	10 20 30	SF00S06-1010WT SF00S06-1020WT SF00S06-1030WT

** optionally bioinert coated stainless steel hardware available

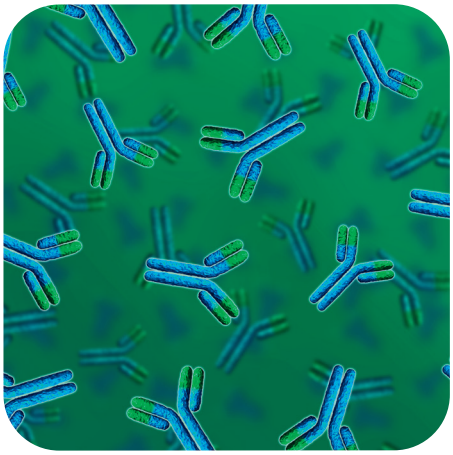
6 µm porous semiprep. columns, stainless steel hardware (max. pressure 4 MPa)

Phase	Column ID [mm]	Column length [mm]
		100
BioPro IEX QA	10 20	QAA0S06-1010WT QAA0S06-1020WT
BioPro IEX SP	10 20	SPA0S06-1010WT SPA0S06-1020WT

Other dimensions on demand




HIC



HIC – BioPro Series

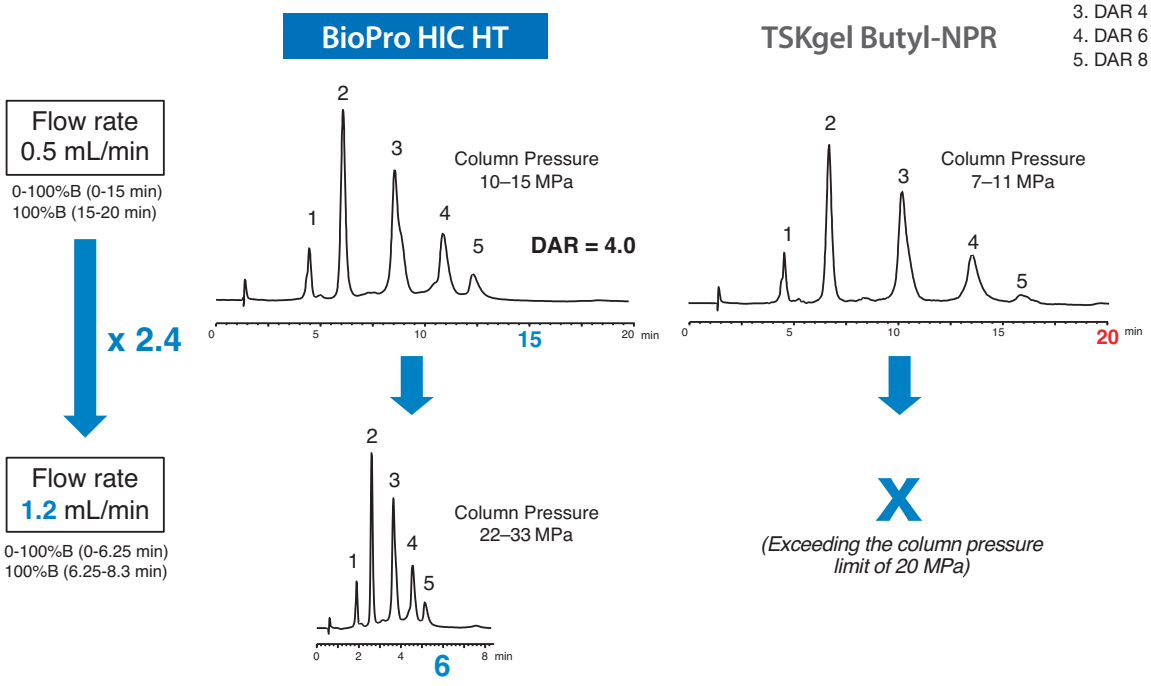
- **Specifically designed for drug-to-antibody conjugates (ADCs) and antibodies**
- **Ideal drug-to-antibody ratio (DAR) analysis**
- **High throughput by reducing analysis time**
- **Excellent batch-to-batch reproducibility**
- **Long term stability**

	BioPro HIC HT	BioPro HIC BF
Base particle	hydrophilic polymer (polymethacrylate)	hydrophilic polymer (polymethacrylate)
Particle size / µm	2.3	4
Pore	non-porous	non-porous
Functional group	butyl	butyl
pH range	2–12	2–12
Pressure limit	40 MPa (5,800 psi)	20 MPa (2,900 psi)
Temperature range	10–60 °C	10–60 °C

High column stability

High throughput by shortening analysis time using high flow rates

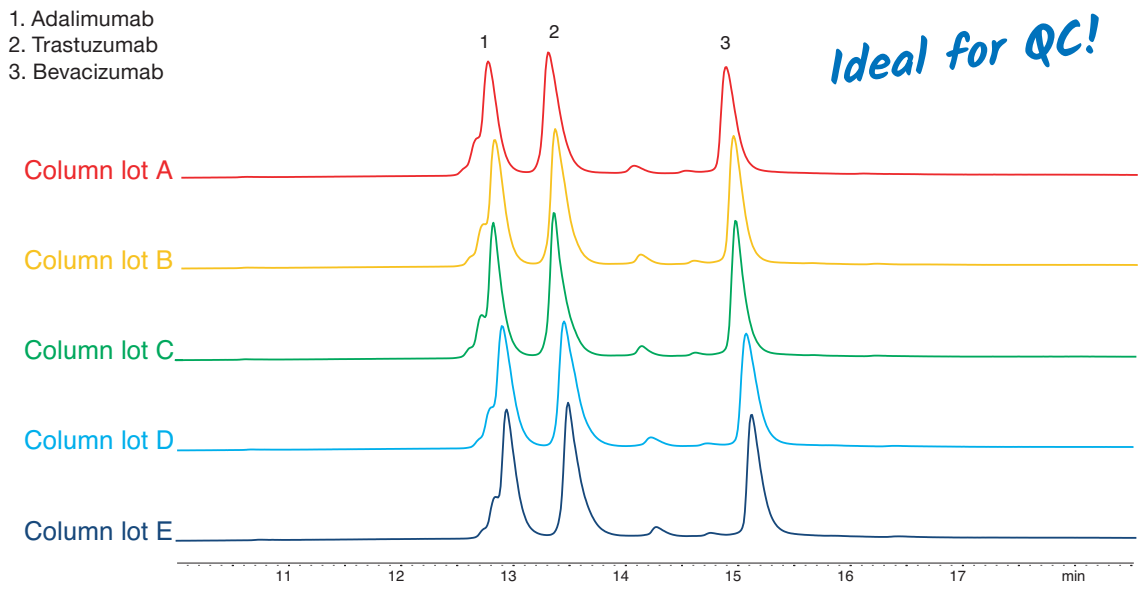
1. DAR 0
2. DAR 2
3. DAR 4
4. DAR 6
5. DAR 8



Column: 100 x 4.6 mm ID
Part No.: BHH00SQ3-1046PTH
Eluent: A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 1.0 M $(NH_4)_2SO_4$
B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)/2-propanol (85/15)
Temperature: 25 °C
Detection: UV at 280 nm
Injection: 10 µL
Sample: Brentuximab vedotin (2.5 mg/mL)

BioPro HIC HT improves analysis throughput of ADCs by 2–3 times with an excellent Drug-to-Antibody Ratio (DAR). The rapid analysis is possible without loss of resolution. Competitor HIC columns fail under these conditions.

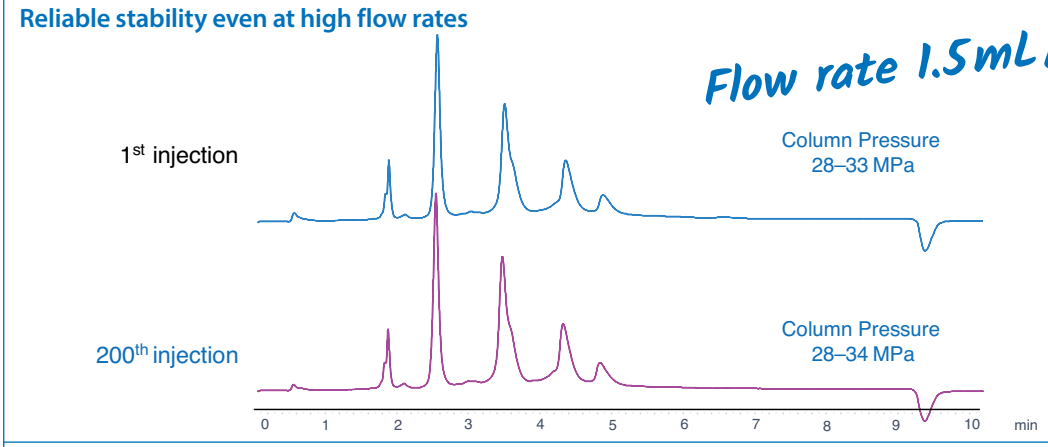
Excellent batch-to-batch reproducibility



Column:	BioPro HIC HT (2.3 µm) 100 x 4.6 mm ID	Flow rate:	0.5 mL/min
Part No.:	BHH00SQ3-1046PTH	Temperature:	25°C
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$	Detection:	UV at 280 nm
	B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Injection:	15 µL
Gradient:	0%B (0–1 min), 0–100%B (1–11 min), 100%B (11–15 min)		

BioPro HIC HT exhibits an excellent batch-to-batch reproducibility making it the ideal choice for quality control analysis of biopharmaceuticals such as mAbs.

Exceptional stability



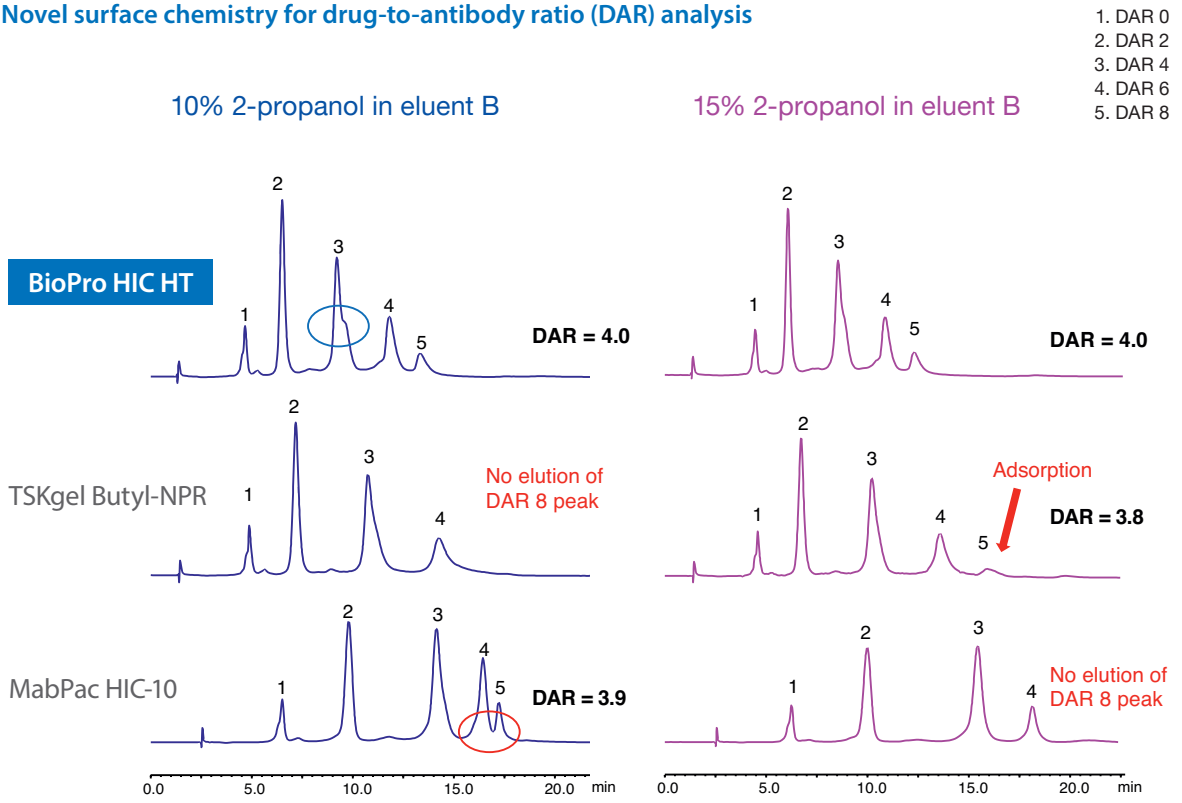
Column:	BioPro HIC HT (2.3 µm) 100 x 4.6 mm ID	Flow rate:	1.5 mL/min
Part No.:	BHH00SQ3-1046PTH	Temperature:	25°C
Eluent:	A) 20 mM NaH_2PO_4-Na_2HPO_4 containing 1.0 M $(NH_4)_2SO_4$ (pH 7.0)	Detection:	UV at 280 nm
	B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)/2-propanol (90/10)	Injection:	10 µL
Gradient:	0–100%B (0–5 min), 100%B (5–8.33 min), 0%B (8.33–12 min)	Sample:	Brentuximab vedotin (2.5 mg/mL)

BioPro HIC HT offers excellent stability under high flow rates / high pressure conditions due to its unique rigid particle and optimised column packing technology.

HIC – BioPro HIC: ADC analysis

Designed for analysis of ADCs

Novel surface chemistry for drug-to-antibody ratio (DAR) analysis



Column:	100 x 4.6 mm ID
Part No.:	BHH00SQ3-1046PTH
Eluent:	A) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 1.0 M $(NH_4)_2SO_4$
	B) 20 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)/2-propanol (90/10) or (85/15)
Gradient:	0–100%B (0–15 min), 100%B (15–20 min), 0%B (20–35 min)
Flow rate:	0.5 mL/min
Temperature:	25°C
Detection:	UV at 280 nm
Injection:	10 µL
Sample:	Brentuximab vedotin (2.5 mg/mL)

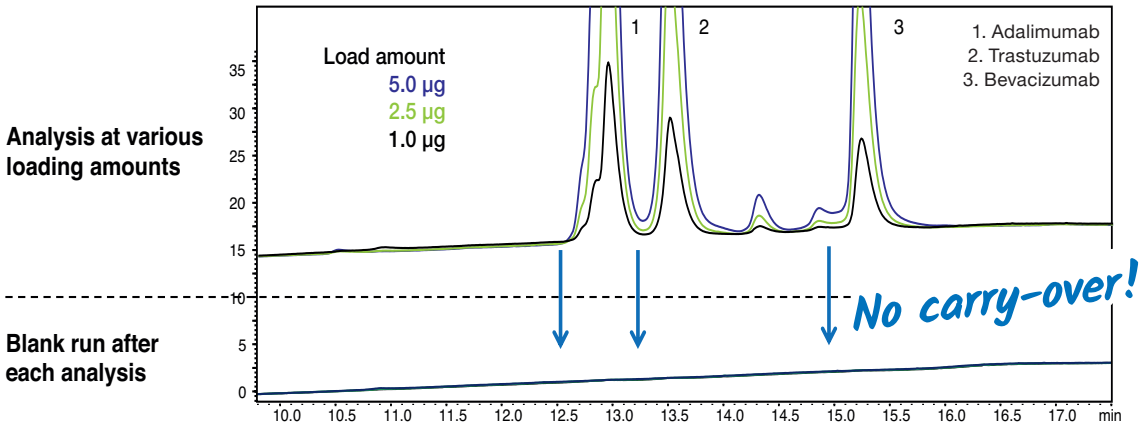
BioPro HIC HT offers higher resolution than conventional HIC columns. Its surface modification suppresses excessive or too strong adsorption of ADCs and results in highly reliable quantification. With varying 2-propanol content, all peaks are completely eluted from the BioPro HIC HT column with high resolution. Another peak is partially separated from peak 3. Additionally, the same DAR values are observed at any content of 2-propanol.

BioPro HIC HT offers:

- **Higher resolution than conventional HIC columns**
- **Highly reliable quantification**
- **Flexible method development**

Excellent recovery and virtually no carry-over

Highly accurate quantification of ADCs and antibodies

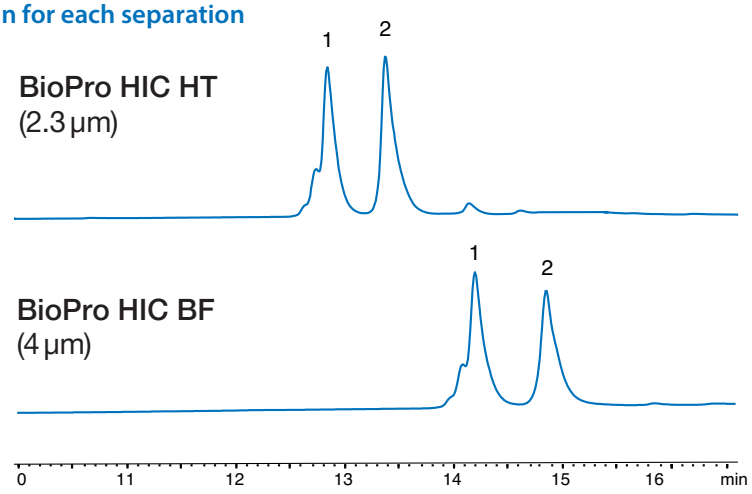


Column: BioPro HIC HT (2.3 µm) 100 x 4.6 mm ID
Part No.: BHH00SQ3-1046PTH
Eluent: A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$
B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)
Gradient: 0%B (0–1 min), 0–100%B (1–11 min), 100%B (11–15 min)
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 280 nm

BioPro HIC HT offers higher linearity over wide loading and virtually no carry-over. This contributes to highly accurate quantitation of ADCs and antibodies.

Different hydrophobicity

The right column for each separation



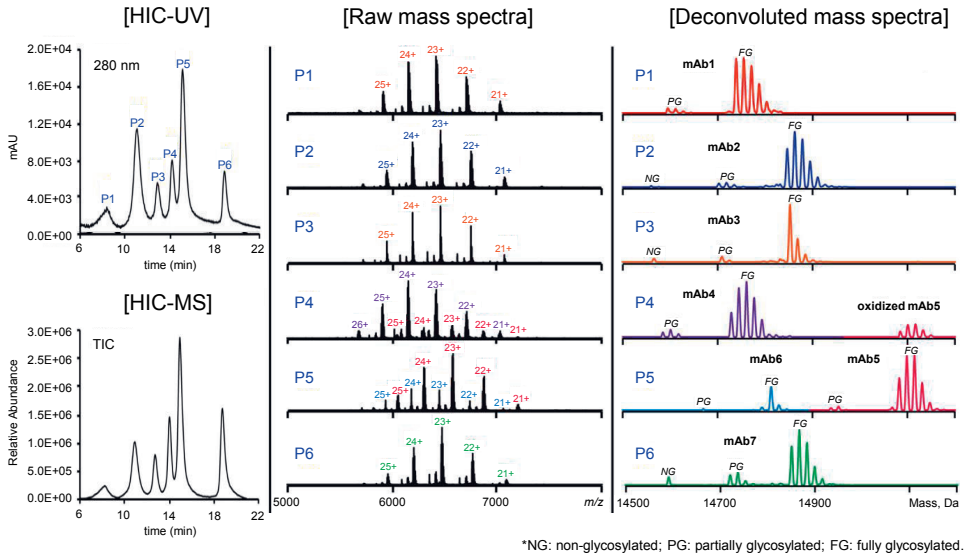
Column: 100 x 4.6 mm ID
Part Nos.: BHH00SQ3-1046PTH
BHB00S04-1046WT
Eluent: A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$
B) 100mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)
Gradient: 0%B (0–1 min), 0–100%B (1–11 min), 100%B (11–15 min)
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV at 280 nm
Injection: 15 µL
Sample: 1. Adalimumab (Humira®; 0.5 mg/mL)
2. Trastuzumab (Herceptin®; 0.5 mg/mL)

BioPro HIC HT is the first choice for ADCs or mAbs. BioPro HIC BF columns show a stronger retention and can therefore be used for the separation of low hydrophobic proteins or especially for the analysis of oxidised mAbs.

HIC – BioPro HIC: Direct HIC-MS coupling

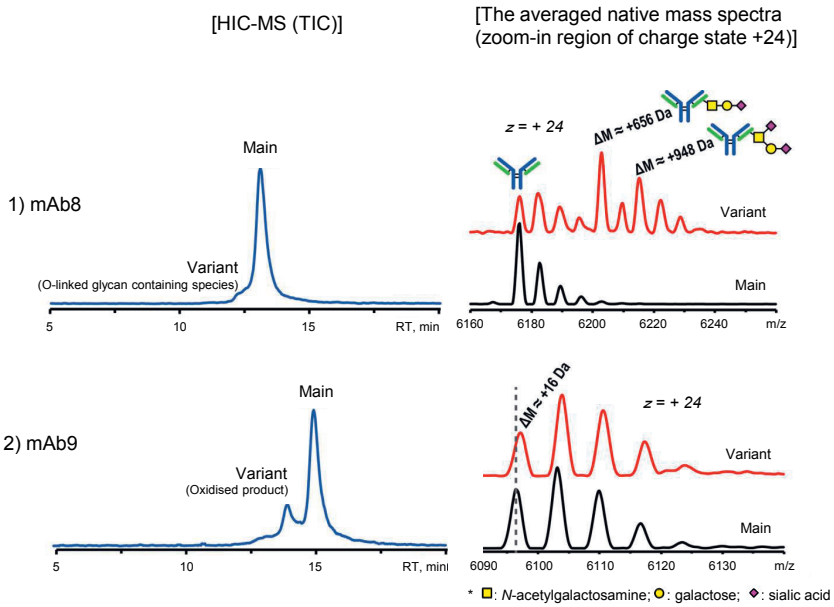
Online native HIC-MS analysis of mAbs and their molecular variants

Separation of an antibody mixture of seven different mAbs



*NG: non-glycosylated; PG: partially glycosylated; FG: fully glycosylated.

Separation of two mAbs from their molecular variants



* □: *N*-acetylgalactosamine; ○: galactose; ◆: sialic acid

Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Injection:	MAb mixture: 3 µL (3–6 µg) MAb 8 and mAb 9: 10 µg each
Part No.:	BHB00S04-1046WT	Sample:	Mixture of 7 in-house mAbs at 1–2 mg/mL each 2 in-house mAbs with molecular variants
Eluent:	A) 3 M ammonium acetate in water B) 100 % water	Setup:	Post-column makeup flow: 100 % water at 1.5 mL/min (reducing salt conc. 6-fold) Splitter to reduce the flow rate to 1–5 µL/min
Gradient:	0 %B (0–2 min), 0–90 %B (2–18 min), 90 %B (18–22 min)		
Flow rate:	0.3 mL/min		
Temperature:	ambient		
Detection:	UV at 280 nm, NSI-MS		

Courtesy by S. Wang, Regeneron Pharmaceuticals Inc.

To enable simultaneous UV and MS detection a post-column makeup flow and a splitter were used. The make-up flow decreases the salt concentration while the splitter reduces the flow rate to enable the coupling to MS. A nanospray ionisation (NSI) was chosen because of its high sensitivity and salt tolerance.

Reference: Y. Yan, T. Xing, S. Wang, T. J. Daly, N. Li, Online coupling of analytical hydrophobic interaction chromatography with native mass spectrometry fo the characterization of monoclonal antibodies and related products, J. Pharm. Biomed. Anal. 186 (2020) 113313.

The influence of salts in HIC separations

The technique known as hydrophobic interaction chromatography is a mode of chromatography that separates proteins by differences in surface hydrophobicity. [1] This method utilises reversible interactions that occur between protein molecules and hydrophobic stationary phase ligands attached to the particle surface.

Certain non-denaturing salts are used to improve the hydrophobic interactions between proteins and the stationary phase. The mobile phase is typically an aqueous solution of salts such as ammonium sulfate or sodium chloride and a buffer to control pH (usually phosphate buffer between pH 6 and 7). The Hofmeister series of lyotropic and chaotropic ions shown below in Fig. 1 provides a template for salt selection. High concentrations of salt, particularly ammonium sulfate, may precipitate proteins; therefore, solubility should be checked under the initial gradient (binding) conditions. The strength of the interaction between the protein and stationary phase decreases with decreasing salt gradient (see Fig. 2). Another option is a change of pH which results in an increase or decrease in the charge on the protein due to the ionisation of acidic or basic groups.

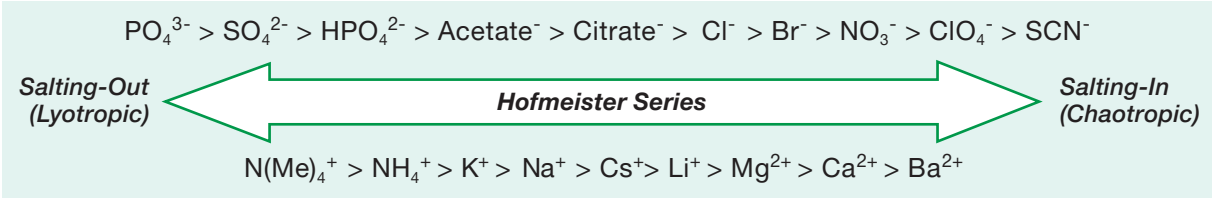


Fig. 1: The Hofmeister Series of lyotropic and chaotropic ions.

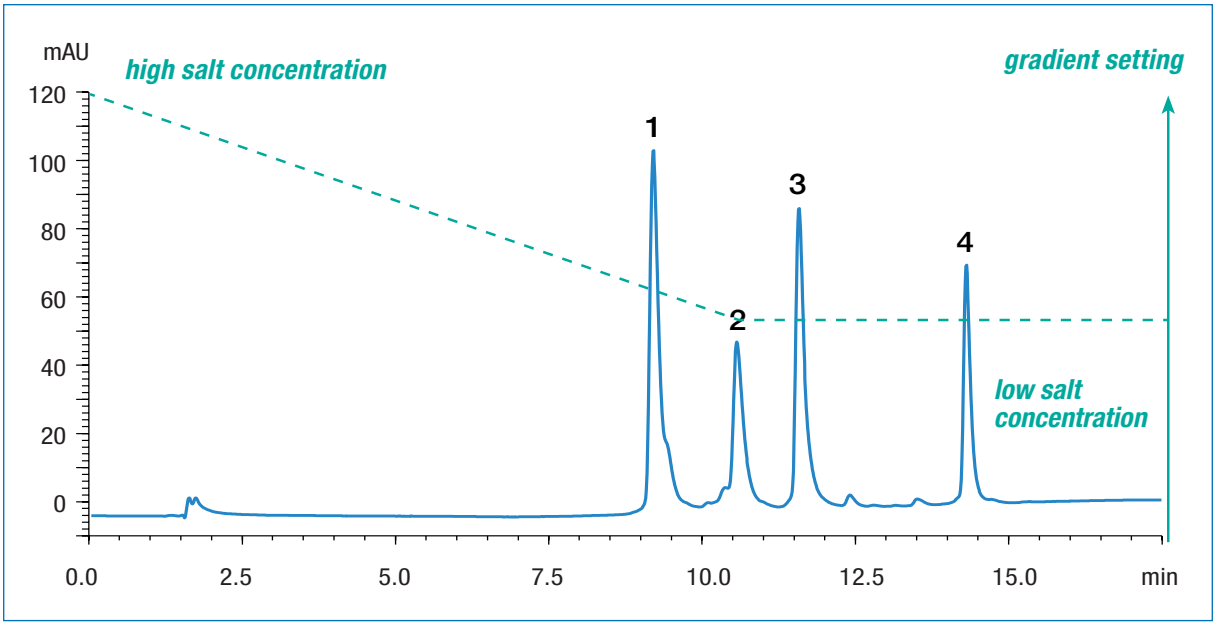


Fig. 2: Method with decreasing salt gradient.

Column:	BioPro HIC BF (100 x 4.6 mm ID)
Part No.:	BHB00S04-1046WT
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$ B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)
Flow rate:	0.5 mL/min
Gradient:	0–100%B (0–11 min), 100%B (11–15 min)
Temperature:	25°C
Detection:	UV at 280 nm
Injection:	15 µL
Samples:	1. Myoglobin (0.73 mg/mL) 2. Ribonuclease A (0.75 mg/mL) 3. Lysozyme (0.25 mg/mL) 4. α-Chymotrypsinogen A (0.25 gm/mL)

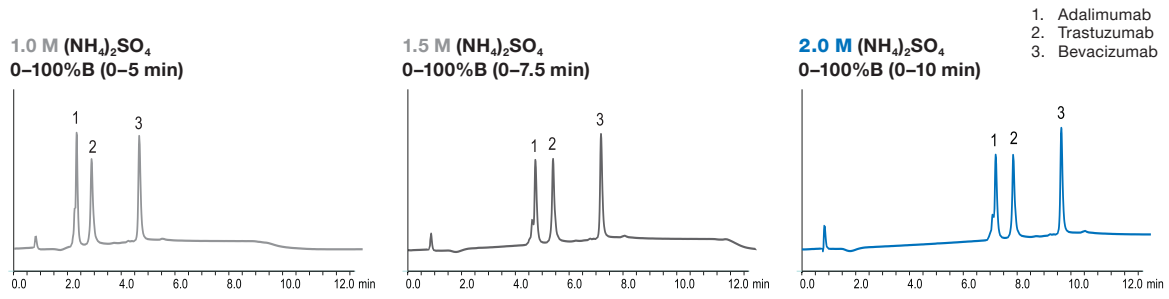
HIC is particularly effective when used to separate proteins and monoconal antibodies. The separation of monoclonal antibodies, mAb aggregates and glycosylated mAbs can be achieved due to their specific hydrophobic properties. It also provides an excellent method for determination of drug-to-antibody ratios in antibody-drug conjugates.

[1] Queiroza, J.A.; Tomaza, C.T.; Cabral, J.M.: Hydrophobic interaction chromatography of proteins, J Biotechnol. 2001, 87, 143-159.

HIC – Expert Tips: Separation factors

Effect of initial salt concentration

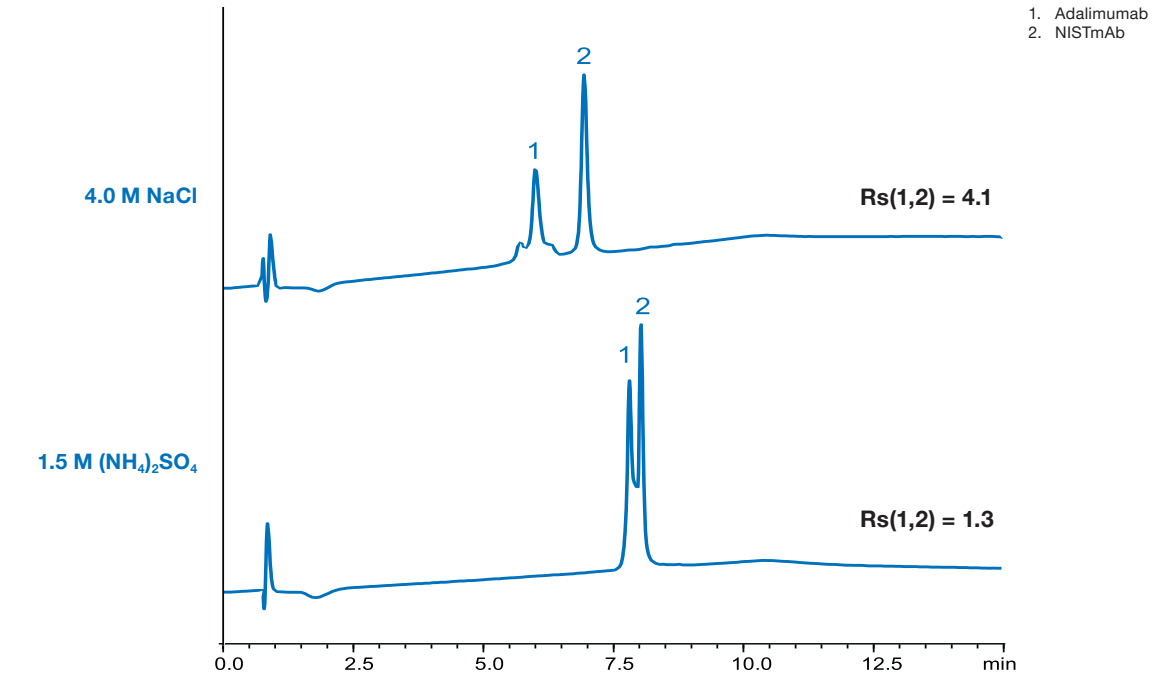
Buffers containing $(NH_4)_2SO_4$ are often used as a mobile phase in HIC mode because $(NH_4)_2SO_4$ has a strong salting-out effect. The higher the initial concentration of $(NH_4)_2SO_4$; the stronger will be the retention of proteins. Therefore, a buffer with a high salt concentration is more suitable for the separation of low hydrophobic proteins with weak retention.



Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Flow rate:	1.0 mL/min
Part No.:	BHB00S04-1046WT	Temperature:	25°C
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing salt	Detection:	UV at 280 nm
	B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Injection:	5 µL (0.5 mg/mL)
Gradient:	0.2 M/min (gradient slope is the same)		

Influence of the type of salt

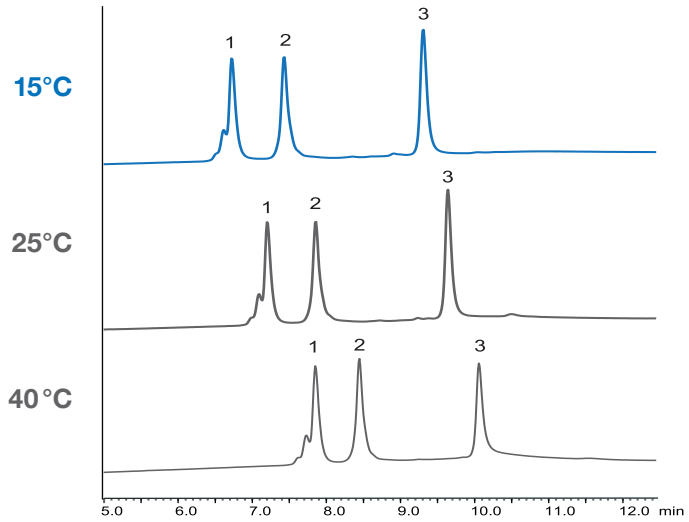
NaCl and CH_3COONH_4 are also used as buffer salts. The separation selectivity varies with the type of salt used in some cases, so changing the type of salt can also be effective when the separation is not sufficient. However, these salts have to be used at very high concentrations to gain retentions comparable to $(NH_4)_2SO_4$. Attention needs to be paid to the prevention of precipitation of salts in the buffer and damage of the LC system.



Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Flow rate:	1.0 mL/min
Part No.:	BHB00S04-1046WT	Temperature:	25°C
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing salt	Detection:	UV at 280 nm
	B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Injection:	10 µL (0.25 mg/mL)
Gradient:	0–100%B (0–10 min), 100%B (10–15 min)		

Temperature influence

In HIC mode, higher temperatures result in longer retention times of proteins. This assumes that the hydrophobic area interacting with the stationary phase becomes larger due to a change in the structure of proteins with increasing temperature, so that the hydrophobic interactions become stronger.

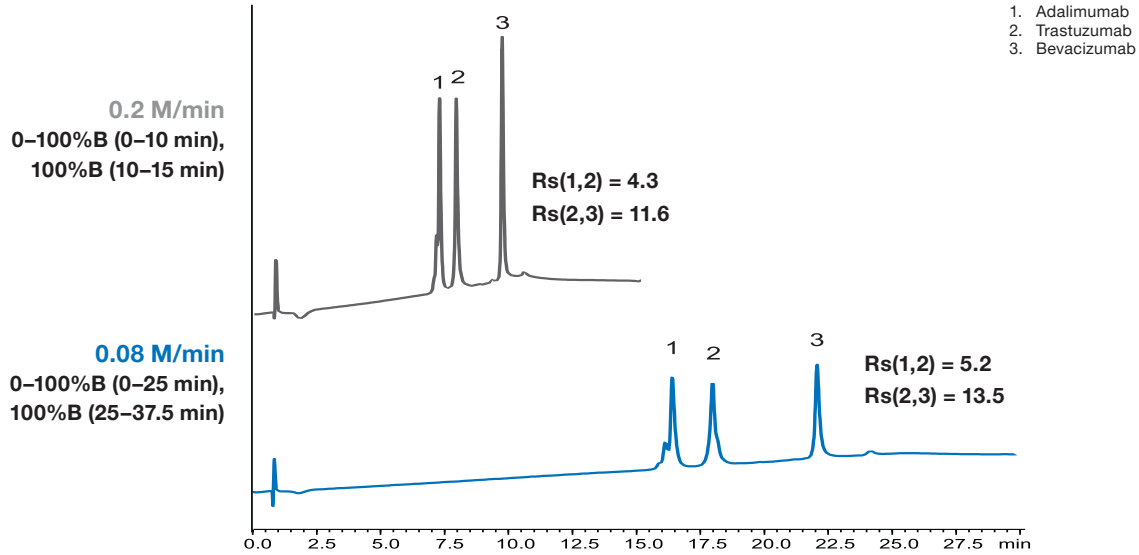


1. Adalimumab
2. Trastuzumab
3. Bevacizumab

Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Gradient:	0–100%B (0–10 min), 100%B (10–15 min)
Part No.:	BHB00S04-1046WT	Flow rate:	1.0 mL/min
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$ B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Detection: Injection:	UV at 280 nm 5 µL (0.5 mg/mL)

Variation of gradient slope

In general, shallower gradients improve the separation and the resulting resolution.



1. Adalimumab
2. Trastuzumab
3. Bevacizumab

Column:	BioPro HIC BF (4 µm) 100 x 4.6 mm ID	Flow rate:	1.0 mL/min
Part No.:	BHB00S04-1046WT	Temperature:	25 °C
Eluent:	A) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0) containing 2.0 M $(NH_4)_2SO_4$ B) 100 mM NaH_2PO_4-Na_2HPO_4 (pH 7.0)	Detection: Injection:	UV at 280 nm 10 µL (0.5 mg/mL)

HIC – Ordering information

Ordering information

Phase	Particle size [µm]	Column ID [mm]	Column length [mm]	Part number	Precolumn filter 2 µm*
					(pack of 5)
BioPro HIC HT (max. pressure 40 MPa)	2.3	4.6	100	BHH00SQ3-1046PTH	XRPRCS35
	2.3	4.6	33	BHH00SQ3-H346PTH	XRPRCS35
BioPro HIC BF (max. pressure 20 MPa)	4	4.6	100	BHB00S04-1046WT	XRPRCS35

*Holder required, part no XRPRCS03
Other dimensions on demand

Substance index

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